

...TENT COOPERATION TREA...

PCTCOMMUNICATION OF
INTERNATIONAL APPLICATIONS

(PCT Article 20)

From the INTERNATIONAL BUREAU

To:

United States Patent and Trademark
Office
(Box PCT)
Crystal Plaza 2
Washington, DC 20231
ETATS-UNIS D'AMERIQUE

in its capacity as designated Office

Date of mailing:

21 August 1997 (21.08.97)

The International Bureau transmits herewith copies of the international applications having the following international application numbers and international publication numbers:

International application no.:

PCT/GB96/03209 L

International publication no.:

WO97 23613

**CORRECTED VERSION
VERSION CORRIGEE**The International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland

Facsimile No.: (41 22) 740 14 35

Authorized officer:

J. Zahra

Telephone No.: (41 22) 338 83 38

PATENT COOPERATION TREATY

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

From the INTERNATIONAL BUREAU

To:

United States Patent and Trademark
Office
(Box PCT)
Crystal Plaza 2
Washington, DC 20231
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year) 08 August 1997 (08.08.97)	
International application No. PCT/GB96/03209	Applicant's or agent's file reference P16589W0/HGH
International filing date (day/month/year) 23 December 1996 (23.12.96)	Priority date (day/month/year) 21 December 1995 (21.12.95)
Applicant BEBBINGTON, Christopher, Robert et al	

1. The designated Office is hereby notified of its election made:

☒ in the demand filed with the International Preliminary Examining Authority on:

18 July 1997 (18.07.97)

☐ in a notice effecting later election filed with the International Bureau on:
2. The election ☒ was
☐ was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Facsimile No.: (41-22) 740.14.35	Authorized officer: Ting Zhao Telephone No.: (41-22) 338.83.38
---	--

PATENT COOPERATION TREATY

WO 97/23613
PCT/GB96/03209

1) DAB
2) HGTH

PCT

From the INTERNATIONAL BUREAU

To:

HALLYBONE, Huw, George
Carpmaels & Ransford
43 Bloomsbury Square
London WC1A 2RA
ROYAUME-UNI

NOTICE INFORMING THE APPLICANT OF THE COMMUNICATION OF THE INTERNATIONAL APPLICATION TO THE DESIGNATED OFFICES

(PCT Rule 47.1(c), first sentence)

Date of mailing (day/month/year) 03 July 1997 (03.07.97)		IMPORTANT NOTICE	
Applicant's or agent's file reference P16589W0/HGH			
International application No. PCT/GB96/03209	International filing date (day month year) 23 December 1996 (23.12.96)	Priority date (day month year) 21 December 1995 (21.12.95)	
Applicant CELLTECH THERAPEUTICS LTD. et al			

1. Notice is hereby given that the International Bureau has communicated, as provided in Article 20, the international application to the following designated Offices on the date indicated above as the date of mailing of this Notice:
AU,BR,CA,CN,CZ,DE,EP,FI,IL,JP,KP,KR,NO,NZ,PL,RO,SK,US

In accordance with Rule 47.1(c), third sentence, those Offices will accept the present Notice as conclusive evidence that the communication of the international application has duly taken place on the date of mailing indicated above and no copy of the international application is required to be furnished by the applicant to the designated Office(s).

2. The following designated Offices have waived the requirement for such a communication at this time:
AL,AM,AP,AT,AZ,BA,BB,BG,BY,CH,CU,DK,EA,EE,ES,GB,GE,HU,IS,KE,KG,KZ,LC,LK,LR,LS,LT,
LU,LV,MD,MG,MK,MN,MW,MX,OA,PT,RU,SD,SE,SG,SI,TJ,TM,TR,TT,UA,UG,UZ,VN

The communication will be made to those Offices only upon their request. Furthermore, those Offices do not require the applicant to furnish a copy of the international application (Rule 49.1(a-bis)).

3. Enclosed with this Notice is a copy of the international application as published by the International Bureau on
03 July 1997 (03.07.97) under No. WO 97/23613

REMINDER REGARDING CHAPTER II (Article 31(2)(a) and Rule 54.2)

If the applicant wishes to postpone entry into the national phase until 30 months (or later in some Offices) from the priority date, a demand for international preliminary examination must be filed with the competent International Preliminary Examining Authority before the expiration of 19 months from the priority date.

It is the applicant's sole responsibility to monitor the 19-month time limit.

Note that only an applicant who is a national or resident of a PCT Contracting State which is bound by Chapter II has the right to file a demand for international preliminary examination.

REMINDER REGARDING ENTRY INTO THE NATIONAL PHASE (Article 22 or 39(1))

If the applicant wishes to proceed with the international application in the national phase, he must, within 20 months or 30 months, or later in some Offices, perform the acts referred to therein before each designated or elected Office.

For further important information on the time limits and acts to be performed for entering the national phase, see the Annex to Form PCT/IB/301 (Notification of Receipt of Record Copy) and Volume II of the PCT Applicant's Guide.

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland	Authorized officer J. Zahra
Facsimile No. (41-22) 740.14.35	Telephone No. (41-22) 338.83.38

PATENT COOPERATION TREATY

PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference P16589WO/HGH	<div style="display: flex; justify-content: space-between;"> <div style="width: 40%;"> FOR FURTHER ACTION </div> <div style="width: 60%; font-size: small;"> see Notification of Transmittal of International Search Report (Form PCT/ISA, 220) as well as, where applicable, item 5 below. </div> </div>	
International application No. PCT/GB 96/03209	International filing date (day, month, year) 23/12/1996	(Earliest) Priority Date (day, month, year) 21/12/1995
Applicant CELLTECH THERAPEUTICS LTD et al.		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.

☒ It is also accompanied by a copy of each prior art document cited in this report.

1. ☐ **Certain claims were found unsearchable** (see Box I).

2. ☐ **Unity of invention is lacking** (see Box II).

3. ☒ The international application contains disclosure of a **nucleotide and/or amino acid sequence listing** and the international search was carried out on the basis of the sequence listing

☐ filed with the international application.
☒ furnished by the applicant separately from the international application,

☐ but not accompanied by a statement to the effect that it did not include matter going beyond the disclosure in the international application as filed.

☐ Transcribed by this Authority

4. With regard to the **title**,

☒ the text is approved as submitted by the applicant.
☐ the text has been established by this Authority to read as follows:

5. With regard to the **abstract**,

☒ the text is approved as submitted by the applicant.
☐ the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this International Search Report, submit comments to this Authority.

6. The figure of the **drawings** to be published with the abstract is:

Figure No. _____ ☐ as suggested by the applicant.
☐ because the applicant failed to suggest a figure
☐ because this figure better characterizes the invention.

☒ None of the figures.

INTERNATIONAL SEARCH REPORT

International Application No.

PC./GB 96/03209

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/12 C07K14/705 C12N15/62 C07K16/00 C12N5/10
A61K35/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 95 02686 A (THE GENERAL HOSPITAL CORPORATION) 26 January 1995 cited in the application see page 12, line 15 - page 16, line 14 see page 36, line 29 - page 51, line 16 ---	1-52
A	WO 93 19163 A (YEDA RESEARCH AND DEVELOPMENT CO. LTD.) 30 September 1993 cited in the application see page 7, line 9 - page 9, line 2 see page 16, line 14 - page 22, line 9 ---	1-52
P,X	WO 96 24671 A (CELL GENESYS, INC.) 15 August 1996 see page 3, line 17 - line 28 see page 10, line 24 - page 26, line 18 see page 28, line 11 - page 29, line 19 see page 30, line 25 - page 34, line 4 --- -/--	1-52

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

* & * document member of the same patent family

Date of the actual completion of the international search

3 July 1997

Date of mailing of the international search report

08.07.97

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+ 31-70) 340-3016

Authorized officer

Montero Lopez, B

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/GB 96/03209

(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	<p>WO 96 23814 A (CELL GENESYS INC) 8 August 1996</p> <p>see page 9, line 28 - page 10, line 31 see page 11, line 20 - page 12, line 9 see page 13, line 11 - page 23, line 26 see page 26, line 1 - line 12 see page 27, line 30 - page 34, line 28; examples</p> <p style="text-align: center;">-----</p>	<p>1-5, 7-13,15, 17-31, 36-52</p>

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/GB 96/03209

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9502686 A	26-01-95	AU 7314094 A	13-02-95
		CA 2166102 A	26-01-95
		CZ 9503408 A	14-08-96
		FI 960178 A	15-01-96
		HU 74252 A	28-11-96
		JP 9500020 T	07-01-97
		NO 960175 A	15-03-96
		ZA 9405204 A	30-05-95

WO 9319163 A	30-09-93	AU 3924393 A	21-10-93
		CA 2132349 A	30-09-93
		EP 0638119 A	15-02-95
		JP 7505282 T	15-06-95

WO 9624671 A	15-08-96	AU 4776196 A	27-08-96

WO 9623814 A	08-08-96	AU 4861396 A	21-08-96

The demand must be filed directly with the competent International Preliminary Examining Authority or, if two or more Authorities are competent, with the one chosen by the applicant. The name or two-letter code of that Authority may be indicated by the applicant on the line below.

IPEA/ **EP**

PCT

CHAPTER II

DEMAND

under Article 31 of the Patent Cooperation Treaty:
The undersigned requests that the international application specified below be the subject of international preliminary examination according to the Patent Cooperation Treaty.

For International Preliminary Examining Authority use only

Identification of IPLA	Date of receipt of DEMAND
------------------------	---------------------------

Box No. I IDENTIFICATION OF THE INTERNATIONAL APPLICATION		Applicant's or agent's file reference P16589WO: HGH
International application No. PCT/GB96/03209	International filing date (day month year) 23 DEC 1996 (23/12/96)	(Earliest) Priority date (day month year) 21 DEC 1995 (21/12/95)

Title of invention

CELL ACTIVATION PROCESS AND REAGENTS THEREFOR

Box No. II APPLICANT(S)	
Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.) CELLTECH THERAPEUTICS LTD. 216 Bath Road Slough Berkshire SL1 4EN UNITED KINGDOM	Telephone No.: Facsimile No.: Teleprinter No.:

State (i.e. country) of nationality: UK	State (i.e. country) of residence: UK
---	---

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.) BEBBINGTON, Christopher, Robert Berry Cottage Westbrook, Boxford, Newbury, Berkshire RG20 8DJ UNITED KINGDOM	
---	--

State (i.e. country) of nationality: UK	State (i.e. country) of residence: UK
---	---

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.) LAWSON, Alastair, David, Griffiths Holden Farm Cheriton Alresford, Hants SO24 0NX UNITED KINGDOM	
---	--

State (i.e. country) of nationality: UK	State (i.e. country) of residence: UK
---	---

<input checked="" type="checkbox"/> Further applicants are indicated on a continuation sheet.

Continuation of Box No. II APPLICANT(S)

If none of the following sub-boxes is used, this sheet is not to be included in the demand.

Name and address: *(Family name followed by given name, for a legal entity full official designation. The address must include postal code and name of country.)*

WEIR, Andrew, Neil, Charles
7 Willow Drive
Twyford
Berkshire RG10 9DD
United Kingdom

State *(i.e. country)* of nationality:

UK

State *(i.e. country)* of residence:

UK

Name and address: *(Family name followed by given name, for a legal entity full official designation. The address must include postal code and name of country.)*

FINNEY, Helene, Margaret
64 Clare Road
Maidenhead
Berkshire SL6 4DQ
UNITED KINGDOM

State *(i.e. country)* of nationality:

UK

State *(i.e. country)* of residence:

UK

Name and address: *(Family name followed by given name, for a legal entity full official designation. The address must include postal code and name of country.)*

State *(i.e. country)* of nationality:

State *(i.e. country)* of residence:

Name and address: *(Family name followed by given name, for a legal entity full official designation. The address must include postal code and name of country.)*

State *(i.e. country)* of nationality:

State *(i.e. country)* of residence:

☐ Further applicants are indicated on another continuation sheet.

Box No. III AGENT OR COMMON REPRESENTATIVE; OR ADDRESS FOR CORRESPONDENCEThe following person is ☒ agent ☐ common representative

- and ☒ has been appointed earlier and represents the applicant(s) also for international preliminary examination.
- ☐ is hereby appointed and any earlier appointment of (an) agent(s)/common representative is hereby revoked.
- ☐ is hereby appointed, specifically for the procedure before the International Preliminary Examining Authority, in addition to the agent(s)/common representative appointed earlier.

Name and address: *(Family name followed by given name for a legal entity, full official designation
The address must include postal code and name of country)*

HALLYBONE, Huw George
Carpmaels & Ransford
43 Bloomsbury Square
LONDON WC1A 2RA
UNITED KINGDOM

Telephone No

0171 242 8692

Facsimile No

0171 405 4166

Teleprinter No

267209

- ☐ Mark this check-box where no agent or common representative is/has been appointed and the space above is used instead to indicate a special address to which correspondence should be sent.

Box No. IV STATEMENT CONCERNING AMENDMENTS

The applicant wishes the International Preliminary Examining Authority*

- (i) ☒ to start the international preliminary examination on the basis of the international application as originally filed.
- (ii) ☐ to take into account the amendments under Article 34 of
- ☐ the description (amendments attached).
 - ☐ the claims (amendments attached).
 - ☐ the drawings (amendments attached).
- (iii) ☐ to take into account any amendments of the claims under Article 19 filed with the International Bureau (a copy is attached).
- (iv) ☐ to disregard any amendments of the claims made under Article 19 and to consider them as reversed.
- (v) ☐ to postpone the start of the international preliminary examination until the expiration of 20 months from the priority date unless that Authority receives a copy of any amendments made under Article 19 or a notice from the applicant that he does not wish to make such amendments (Rule 69.1(d)). *(This check-box may be marked only where the time limit under Article 19 has not yet expired.)*

* Where no check-box is marked, international preliminary examination will start on the basis of the international application as originally filed or, where a copy of amendments to the claims under Article 19 and/or amendments of the international application under Article 34 are received by the International Preliminary Examining Authority before it has begun to draw up a written opinion or the international preliminary examination report, as so amended.

Box No. V ELECTION OF STATES

- ☒ The applicant hereby elects all eligible States *(that is, all States which have been designated and which are bound by Chapter II of the PCT)* except

(If the applicant does not wish to elect certain eligible States, the name(s) or country code(s) of those States must be indicated above.)

Box No. VI CHECK LIST

The demand is accompanied by the following documents for the purposes of international preliminary examination:

- | | | |
|--|---|--------|
| 1. amendments under Article 34 | | |
| description | : | sheets |
| claims | : | sheets |
| drawings | : | sheets |
| 2. letter accompanying amendments under Article 34 | : | sheets |
| 3. copy of amendments under Article 19 | : | sheets |
| 4. copy of statement under Article 19 | : | sheets |
| 5. other (specify): | : | sheets |

For International Preliminary
Examining Authority use only

received

not received

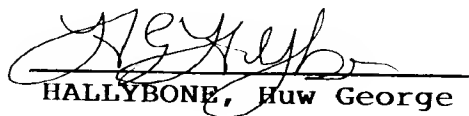
☐☐☐☐☐☐☐☐☐☐☐☐☐☐

The demand is also accompanied by the item(s) marked below:

- | | |
|--|--|
| 1. <input type="checkbox"/> separate signed power of attorney | 4. <input checked="" type="checkbox"/> fee calculation sheet |
| 2. <input type="checkbox"/> copy of general power of attorney | 5. <input type="checkbox"/> other (specify): |
| 3. <input type="checkbox"/> statement explaining lack of signature | |

Box No. VII SIGNATURE OF APPLICANT, AGENT OR COMMON REPRESENTATIVE

Next to each signature, indicate the name of the person signing and the capacity in which the person signs (if such capacity is not obvious from reading the demand).


HALLYBONE, Huw George

For International Preliminary Examining Authority use only

1. Date of actual receipt of DEMAND:

2. Adjusted date of receipt of demand due to CORRECTIONS under Rule 60.1(b):

3. ☐ The date of receipt of the demand is AFTER the expiration of 19 months from the priority date and item 4 or 5, below, does not apply.

☐ The applicant has been informed accordingly.

4. ☐ The date of receipt of the demand is WITHIN the period of 19 months from the priority date as extended by virtue of Rule 80.5.

5. ☐ Although the date of receipt of the demand is after the expiration of 19 months from the priority date, the delay in arrival is EXCUSED pursuant to Rule 82.

For International Bureau use only

Demand received from IPEA on:

PATENT COOPERATION TREATY

PCT

1997


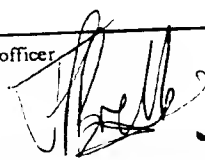
INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference P16589WO: HGH	FOR FURTHER ACTION See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/GB 96/ 03209	International filing date (day/month/year) 23/12/1996	Priority date (day/month/year) 21/12/1995
International Patent Classification (IPC) or national classification and IPC C12N15/12		
Applicant CELLTECH THERAPEUTICS LTD. et al.		

- This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.
- This **REPORT** consists of a total of Six sheets, including this cover sheet.
☐ This report is also accompanied by **ANNEXES**, i.e., sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).
These annexes consists of a total of _____ sheets.

- This report contains indications and corresponding pages relating to the following items:
 - I ☒ Basis of the report
 - II ☐ Priority
 - III ☐ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
 - IV ☐ Lack of unity of invention
 - V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
 - VI ☐ Certain documents cited
 - VII ☐ Certain defects in the international application
 - VIII ☒ Certain observations on the international application

Date of submission of the demand 18/07/1997	Date of completion of this report 02 OCT 1997
Name and mailing address of the IPEA/  European Patent Office D-80298 Munich Tel. (+ 49-89) 2399-0, Tx: 523656 epmu d Fax: (+ 49-89) 2399-4465	Authorized officer  J. Bretherick Telephone No.

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

I. Basis of the report

1. This report has been drawn up on the basis of (Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments.):

☒ the international application as originally filed.

☐ the description, pages _____, as originally filed,
pages _____, filed with the demand,
pages _____, filed with the letter of _____,
pages _____, filed with the letter of _____,

☐ the claims, Nos. _____, as originally filed,
Nos. _____, as amended under Article 19,
Nos. _____, filed with the demand,
Nos. _____, filed with the letter of _____,
Nos. _____, filed with the letter of _____,

☐ the drawings, sheets/fig _____, as originally filed,
sheets/fig _____, filed with the demand,
sheets/fig _____, filed with the letter of _____,
sheets/fig _____, filed with the letter of _____.

2. The amendments have resulted in the cancellation of:

☐ the description, pages _____.
☐ the claims, Nos. _____.
☐ the drawings, sheets/fig _____.

3. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

4. Additional observations, if necessary:

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step and industrial applicability; citations and explanations supporting such statement

1. STATEMENT

Novelty (N)	Claims 1-52_____	YES
	Claims _____	NO
Inventive Step (IS)	Claims 1-52_____	YES
	Claims _____	NO
Industrial Applicability (IA)	Claims 11-52_____	YES
	Claims 1-10 opinion reserved_____	NO

2. CITATIONS AND EXPLANATIONS

1. This Report has been written on the assumption that the entire claimed subject-matter enjoy the priority right assigned to GB application 9526131.9, filed 21/12/95. Were this not the case, then the respective disclosures of WO 96 24671 A, published 15/08/96 and WO 96 23814 A, published 08/08/96, might prove to be relevant within the meaning of Art. 33 PCT with respect to novelty and/or inventive step.
2. The current claims are related to chimeric receptor systems and their encoding DNA in the form of delivery systems which enable an enhancement of the cellular response to the interaction of a cell ligand with its receptor, by virtue of the inclusion of a second not naturally associated intracellular (cytoplasmic) signalling components.

The closest prior art is considered to be WO 95 02686 A, which discloses chimeric receptors wherein the extra-cellular receptor interacts with the normal ligand for

this receptor and internal not naturally associated signal component results in a cellular response. In the detailed exemplification the surface receptor component recognises a ligand associated with a pathogen, target cell or target infective agent, whilst the intracellular signalling component is a protein-tyrosine kinase capable of signalling the therapeutic cell in which the chimeric receptor is expressed to destroy the target cell or target infective agent. This also includes the associated encoding DNA. A similar disclosure is made in WO 93 19163 A. Both documents are cited in the application.

The difference between the subject-matter of claims 11-44 in broad terms is the presence of a second cytoplasmic signalling component which is not naturally linked with the surface receptor component. The technical problem to be solved is thus the provision of alternative constructs for use in obtaining cell activation in response to an extracellular stimulus. It is not indicated in the prior art. The possibility of provision of two internal and possibly independent intracellular signals from one stimulatory interaction is not anticipated nor contemplated in the art cited in the International Search Report.

Novelty and an inventive step under Art. 33 PCT are therefore acknowledged. This applies mutatis mutandis to the subject-matter of claims 1-10, for methods of cell activation and to claims 45-52 for the cells transfected with the corresponding encoding DNA and the DNA itself, respectively.

3. The subject-matter of claims 1-10 when interpreted in the light of the description encompasses methods of treatment of the human or animal body. An assessment of the industrial applicability under Art. 33(1)(4) PCT of

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

said claims is reserved, since there is are no unified
criteria for such an assessment within the PCT system.

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

Claim 52 is for DNA coding for a recombinant chimeric receptor for use in a delivery system according to any one of claims 11 to 44. The delivery systems are for example comprised of separate encoding DNAs in some instances, each having at least one (i.e. one or more) different cytoplasmic signalling components, for example claim 15. Claim 52 might be construed as falling within the disclosure of the prior art cited in Part V, since this deals with individual DNAs encoding chimeric receptors of this type which by definition have only one cytoplasmic signalling component.

PATENT COOPERATION TREATY

From the
INTERNATIONAL PRELIMINARY EXAMINING AUTHORITY

PCT

To:

HALLYBONE, Huw George
CARPMAELS & RANSFORD
43 Bloomsbury Square
London WC1A 2RA
GRANDE BRETAGNE

NOTIFICATION OF TRANSMITTAL OF INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Rule 71.1)

Date of mailing (day, month, year) 02 OCT 1997			
Applicant's or agent's file reference P16589WO: HGH	IMPORTANT NOTIFICATION		
International application No. PCT/GB 96/ 03209 ✓	<table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <td style="width: 50%; padding: 5px;">International filing date (day, month, year) 23/12/1996</td> <td style="width: 50%; padding: 5px;">Priority date (day, month, year) 21/12/1995</td> </tr> </table>	International filing date (day, month, year) 23/12/1996	Priority date (day, month, year) 21/12/1995
International filing date (day, month, year) 23/12/1996	Priority date (day, month, year) 21/12/1995		
Applicant CELLTECH THERAPEUTICS LTD. et al.			

1. The applicant is hereby notified that this International Preliminary Examining Authority transmits herewith the international preliminary examination report and its annexes, if any, established on the international application.

2. A copy of the report and its annexes, if any, is being transmitted to the International Bureau for communication to all the elected Offices.

3. Where required by any of the elected Offices, the International Bureau will prepare an English translation of the report (but not of any annexes) and will transmit such translation to those Offices.

4. **REMINDER**

The applicant must enter the national phase before each elected Office by performing certain acts (filing translations and paying national fees) within 30 months from the priority date (or later in some Offices)(Article 39(1))(see also the reminder sent by the International Bureau with Form PCT/IB.301).

Where a translation of the international application must be furnished to an elected Office, that translation must contain a translation of any annexes to the international preliminary examination report. It is the applicant's responsibility to prepare and furnish such translation directly to each elected Office concerned.

For further details on the applicable time limits and requirements of the elected Offices, see Volume II of the PCT Applicant's Guide.

Name and mailing address of the IPEA:



European Patent Office
D-80298 Munich
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Authorized officer

Peter Ehrenreich

Telephone No.

PATENT COOPERATION TREATY

PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference P16589WO: HGH	FOR FURTHER ACTION See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/GB 96/ 03209	International filing date (day, month, year) 23/12/1996	Priority date (day, month, year) 21/12/1995
International Patent Classification (IPC) or national classification and IPC C12N15/12		
Applicant CELLTECH THERAPEUTICS LTD. et al.		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.


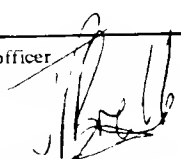
2. This **REPORT** consists of a total of SIX sheets, including this cover sheet.

☐ This report is also accompanied by **ANNEXES**, i.e., sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consists of a total of _____ sheets.

3. This report contains indications and corresponding pages relating to the following items:

- I ☒ Basis of the report
- II ☐ Priority
- III ☐ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
- IV ☐ Lack of unity of invention
- V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- VI ☐ Certain documents cited
- VII ☐ Certain defects in the international application
- VIII ☒ Certain observations on the international application

Date of submission of the demand 18/07/1997	Date of completion of this report 02 OCT 1997
Name and mailing address of the IPEA:  European Patent Office D-80298 Munich Tel. (+ 49-89) 2399-0, Tx: 523656 epmu d Fax: (+ 49-89) 2399-4465	Authorized officer  J. Bretherick Telephone No.

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

I. Basis of the report

1. This report has been drawn up on the basis of (Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments.):

☒ the international application as originally filed.

☐ the description, pages _____, as originally filed,
pages _____, filed with the demand,
pages _____, filed with the letter of _____,
pages _____, filed with the letter of _____.

☐ the claims, Nos. _____, as originally filed,
Nos. _____, as amended under Article 19,
Nos. _____, filed with the demand,
Nos. _____, filed with the letter of _____,
Nos. _____, filed with the letter of _____.

☐ the drawings, sheets/fig _____, as originally filed,
sheets/fig _____, filed with the demand,
sheets/fig _____, filed with the letter of _____,
sheets/fig _____, filed with the letter of _____.

2. The amendments have resulted in the cancellation of:

☐ the description, pages _____.
☐ the claims, Nos. _____.
☐ the drawings, sheets/fig _____.

3. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

4. Additional observations, if necessary:

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step and industrial applicability; citations and explanations supporting such statement

1. STATEMENT

Novelty (N)	Claims 1-52 _____	YES
	Claims _____	NO
Inventive Step (IS)	Claims 1-52 _____	YES
	Claims _____	NO
Industrial Applicability (IA)	Claims 11-52 _____	YES
	Claims 1-10 opinion reserved _____	NO

2. CITATIONS AND EXPLANATIONS

1. This Report has been written on the assumption that the entire claimed subject-matter enjoy the priority right assigned to GB application 9526131.9, filed 21/12/95. Were this not the case, then the respective disclosures of WO 96 24671 A, published 15/08/96 and WO 96 23814 A, published 08/08/96, might prove to be relevant within the meaning of Art. 33 PCT with respect to novelty and/or inventive step.
2. The current claims are related to chimeric receptor systems and their encoding DNA in the form of delivery systems which enable an enhancement of the cellular response to the interaction of a cell ligand with its receptor, by virtue of the inclusion of a second not naturally associated intracellular (cytoplasmic) signalling components.

The closest prior art is considered to be WO 95 02686 A, which discloses chimeric receptors wherein the extracellular receptor interacts with the normal ligand for

this receptor and internal not naturally associated signal component results in a cellular response. In the detailed exemplification the surface receptor component recognises a ligand associated with a pathogen, target cell or target infective agent, whilst the intracellular signalling component is a protein-tyrosine kinase capable of signalling the therapeutic cell in which the chimeric receptor is expressed to destroy the target cell or target infective agent. This also includes the associated encoding DNA. A similar disclosure is made in WO 93 19163 A. Both documents are cited in the application.

The difference between the subject-matter of claims 11-44 in broad terms is the presence of a second cytoplasmic signalling component which is not naturally linked with the surface receptor component. The technical problem to be solved is thus the provision of alternative constructs for use in obtaining cell activation in response to an extracellular stimulus. It is not indicated in the prior art. The possibility of provision of two internal and possibly independent intracellular signals from one stimulatory interaction is not anticipated nor contemplated in the art cited in the International Search Report.

Novelty and an inventive step under Art. 33 PCT are therefore acknowledged. This applies mutatis mutandis to the subject-matter of claims 1-10, for methods of cell activation and to claims 45-52 for the cells transfected with the corresponding encoding DNA and the DNA itself, respectively.

3. The subject-matter of claims 1-10 when interpreted in the light of the description encompasses methods of treatment of the human or animal body. An assessment of the industrial applicability under Art. 33(1)(4) PCT of

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

said claims is reserved, since there is are no unified
criteria for such an assessment within the PCT system.

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

Intern. application No.
PCT/GB96/03209

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

Claim 52 is for DNA coding for a recombinant chimeric receptor for use in a delivery system according to any one of claims 11 to 44. The delivery systems are for example comprised of separate encoding DNAs in some instances, each having at least one (i.e. one or more) different cytoplasmic signalling components, for example claim 15. Claim 52 might be construed as falling within the disclosure of the prior art cited in Part V, since this deals with individual DNAs encoding chimeric receptors of this type which by definition have only one cytoplasmic signalling component.

PCT

REQUEST

The undersigned requests that the present international application be processed according to the Patent Cooperation Treaty.

For receiving Office use only

International Application No.

International Filing Date

Name of receiving Office and "PCT International Application"

Applicant's or agent's file reference
(if desired) (12 characters maximum)

P16589WO/HGH

Box No. I TITLE OF INVENTION

CELL ACTIVATION PROCESS AND REAGENTS THEREFOR

Box No. II APPLICANT

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

CELLTECH THERAPEUTICS LTD
216 Bath Road, Slough,
Berkshire, SL1 4EN,
United Kingdom.

☐ This person is also inventor.

Telephone No.

Facsimile No.

Teleprinter No.

State (i.e. country) of nationality:
UK

State (i.e. country) of residence:
UK

This person is applicant
for the purposes of:

☐ all designated
States

☒ all designated States except
the United States of America

☐ the United States
of America only

☐ the States indicated in
the Supplemental Box

Box No. III FURTHER APPLICANT(S) AND/OR (FURTHER) INVENTOR(S)

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

BEBBINGTON, Christopher Robert
Berry Cottage,
Westbrook, Boxford,
Newbury,
Berkshire, RG20 8DJ,
United Kingdom.

This person is:

☐ applicant only

☒ applicant and inventor

☐ inventor only (If this check-box
is marked, do not fill in below.)

State (i.e. country) of nationality:
UK

State (i.e. country) of residence:
UK

This person is applicant
for the purposes of:

☐ all designated
States

☐ all designated States except
the United States of America

☒ the United States
of America only

☐ the States indicated in
the Supplemental Box

☒ Further applicants and/or (further) inventors are indicated on a continuation sheet.

Box No. IV AGENT OR COMMON REPRESENTATIVE; OR ADDRESS FOR CORRESPONDENCE

The person identified below is hereby/has been appointed to act on behalf
of the applicant(s) before the competent International Authorities as:

☐ agent

☐ common representative

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

HALLYBONE, Huw George
CARPMAELS & RANSFORD
43 Bloomsbury Square
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United Kingdom

Telephone No.

0171 242 8692

Facsimile No.

0171 405 4166

Teleprinter No.

Telex: 267209

☐ Mark this check-box where no agent or common representative is/has been appointed and the space above is used instead to indicate a special address to which correspondence should be sent.

Continuation of Box No. III FURTHER APPLICANTS AND/OR (FURTHER) INVENTORS

If none of the following sub-boxes is used, this sheet is not to be included in the request.

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

LAWSON, Alastair David Griffiths
Holden Farm, Cheriton
Alresford
Hants., SO24 ONX
United Kingdom

This person is:

- ☐ applicant only
☒ applicant and inventor
☐ inventor only (If this check-box is marked, do not fill in below.)

State (i.e. country) of nationality:

UK

State (i.e. country) of residence:

UK

This person is applicant for the purposes of:

- ☐ all designated States ☐ all designated States except the United States of America ☒ the United States of America only ☐ the States indicated in the Supplemental Box

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

WEIR, Andrew Neil Charles
7 Willow Drive
Twyford
Berkshire, RG10 9DD
United Kingdom

This person is:

- ☐ applicant only
☒ applicant and inventor
☐ inventor only (If this check-box is marked, do not fill in below.)

State (i.e. country) of nationality:

UK

State (i.e. country) of residence:

UK

This person is applicant for the purposes of:

- ☐ all designated States ☐ all designated States except the United States of America ☒ the United States of America only ☐ the States indicated in the Supplemental Box

Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

FINNEY, Helene Margaret
64 Clare Road
Maidenhead
Berkshire, SL6 4DQ
United Kingdom

This person is:

- ☐ applicant only
☒ applicant and inventor
☐ inventor only (If this check-box is marked, do not fill in below.)

State (i.e. country) of nationality:

UK

State (i.e. country) of residence:

UK

This person is applicant for the purposes of:

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Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.)

This person is:

- ☐ applicant only
☐ applicant and inventor
☐ inventor only (If this check-box is marked, do not fill in below.)

State (i.e. country) of nationality:

State (i.e. country) of residence:

This person is applicant for the purposes of:

- ☐ all designated States ☐ all designated States except the United States of America ☐ the United States of America only ☐ the States indicated in the Supplemental Box

☐ Further applicants and/or (further) inventors are indicated on another continuation sheet.

Box No.V DESIGNATION OF STATES

The following designations are hereby made under Rule 4.9(a) (mark the applicable check-boxes; at least one must be marked):

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In addition to the designations made above, the applicant also makes under Rule 4.9(b) all designations which would be permitted under the PCT except the designation(s) of

The applicant declares that those additional designations are subject to confirmation and that any designation which is not confirmed before the expiration of 15 months from the priority date is to be regarded as withdrawn by the applicant at the expiration of that time limit. (Confirmation of a designation consists of the filing of a notice specifying that designation and the payment of the designation and confirmation fees. Confirmation must reach the receiving Office within the 15-month time limit.)

Box No. VI PRIORITY CLAIM		Further priority claims are indicated in the Supplemental Box <input type="checkbox"/>	
The priority of the following earlier application(s) is hereby claimed:			
Country (in which, or for which, the application was filed)	Filing Date (day/month/year)	Application No.	Office of filing (only for regional or international application)
item (1) United Kingdom	21st December 1995	9526131.9	
item (2)			
item (3)			
Mark the following check-box if the certified copy of the earlier application is to be issued by the Office which for the purposes of the present international application is the receiving Office (a fee may be required): <input checked="" type="checkbox"/> The receiving Office is hereby requested to prepare and transmit to the International Bureau a certified copy of the earlier application(s) identified above as item(s): <u>9526131.9</u>			
Box No. VII INTERNATIONAL SEARCHING AUTHORITY			
Choice of International Searching Authority (ISA) (If two or more International Searching Authorities are competent to carry out the international search, indicate the Authority chosen; the two-letter code may be used): ISA / _____			
Earlier search Fill in where a search (international, international-type or other) by the International Searching Authority has already been carried out or requested and the Authority is now requested to base the international search, to the extent possible, on the results of that earlier search. Identify such search or request either by reference to the relevant application (or the translation thereof) or by reference to the search request. Country (or regional Office): _____ Date (day/month/year): _____ Number: _____			
Box No. VIII CHECK LIST			
This international application contains the following number of sheets: 1. request : 4 sheets 2. description : 41 sheets 3. claims : 8 sheets 4. abstract : 1 sheets 5. drawings : 40 sheets Total : 94 sheets		This international application is accompanied by the item(s) marked below: 1. <input type="checkbox"/> separate signed power of attorney 2. <input type="checkbox"/> copy of general power of attorney 3. <input type="checkbox"/> statement explaining lack of signature 4. <input type="checkbox"/> priority document(s) identified in Box No. VI as item(s): 5. <input checked="" type="checkbox"/> fee calculation sheet 6. <input type="checkbox"/> separate indications concerning deposited microorganisms 7. <input type="checkbox"/> nucleotide and/or amino acid sequence listing (diskette) 8. <input checked="" type="checkbox"/> other (specify): <u>23/77 & 51/77</u>	
Figure No. _____ of the drawings (if any) should accompany the abstract when it is published.			
Box No. IX SIGNATURE OF APPLICANT OR AGENT			
Next to each signature, indicate the name of the person signing and the capacity in which the person signs (if such capacity is not obvious from reading the request).			
 HALLYBONE, Huw George			

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/12, C07K 14/705, C12N 15/62, C07K 16/00, C12N 5/10, A61K 35/12		A2	(11) International Publication Number: WO 97/23613
			(43) International Publication Date: 3 July 1997 (03.07.97)
(21) International Application Number: PCT/GB96/03209 (22) International Filing Date: 23 December 1996 (23.12.96) (30) Priority Data: ✓ 9526131.9 21 December 1995 (21.12.95) GB (71) Applicant (for all designated States except US): CELLTECH THERAPEUTICS LTD. [GB/GB]; 216 Bath Road, Slough, Berkshire SL1 4EN (GB). (72) Inventors; and (75) Inventors/Applicants (for US only): BEBBINGTON, Christopher, Robert [GB/GB]; Berry Cottage, Westbrook, Boxford, Newbury, Berkshire RG20 8DJ (GB). LAWSON, Alastair, David, Griffiths [GB/GB]; Holden Farm, Cheriton, Alresford, Hants. SO24 0NX (GB). WEIR, Andrew, Neil, Charles [GB/GB]; 7 Willow Drive, Twyford, Berkshire RG10 9DD (GB). FINNEY, Helene, Margaret [GB/GB]; 64 Clare Road, Maidenhead, Berkshire SL6 4DQ (GB). (74) Agent: HALLYBONE, Huw, George; Carpmaels & Ransford, 43 Bloomsbury Square, London WC1A 2RA (GB).			(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i>
(54) Title: CELL ACTIVATION PROCESS AND REAGENTS THEREFOR			
(57) Abstract A cell activation process is described in which an effector cell is transformed with DNA coding for a chimeric receptor containing two or more different cytoplasmic signalling components. The activated cell may be of use in medicine for example in the treatment of diseases such as cancer.			

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CELL ACTIVATION PROCESS AND REAGENTS THEREFOR

5 This invention relates to a process for activating cells, a DNA delivery system for achieving cell activation and the use of activated cells in medicine.

10 The natural T-cell receptor is a complex association of polypeptide chains comprising antigen binding, transmembrane and cytoplasmic components. Binding of antigen to the receptor in the correct context triggers a series of intracellular events leading to activation of the T-cell and for example destruction of the antigen presenting target cell. Before recognition of the antigen can take place, the antigen must be presented in association with MHC molecules.

15 It would be highly desirable if this requirement for MHC could be bypassed by engineering T-cells to become active on binding ligands other than a natural MHC-presented antigen. This would provide a means of avoiding the variability between individuals associated with MHC presentation and
20 would also permit the targeting of more highly expressed surface antigens thereby increasing the efficacy of lymphocyte mediated therapy, for example in tumour therapy.

25 Chimeric receptors have been designed to target T-cells to cells expressing antigen on their cell surface. Such recombinant chimeric receptors include chimeras containing binding domains from antibodies and intracellular signalling domains from the T-cell receptor, termed 'T-bodies' [see for example Published International Patent Specifications Nos. WO 92/10591, WO 92/15322, WO 93/19163 and WO 95/02686].

30 The recombinant chimeric receptors described in the art are composed of a ligand binding component, a transmembrane component and a cytoplasmic component. It has been found however, that transfection of T-cells with these recombinant chimeric receptors does not result in
35 acceptable levels of T-cell activation upon antigen binding unless the T-cell is also co-stimulated by, for example, treatment with high levels of

interleukin 2 [Il-2]. The need for co-stimulation makes the method suitable principally for ex-vivo treatment of patients. This is a lengthy and complicated procedure.

- 5 The present invention offers an alternative to the present ex-vivo approach in that it achieves improved ex-vivo activation without the need for addition of costimulating agents such as Il-2. It also advantageously provides successful in-vivo redirection and activation of T-cells, particularly in response to a single type of extracellular interaction.

10 Essentially the invention provides an effector cell which has been transformed with DNA coding for a chimeric receptor. The chimeric receptor contains two or more different signalling cytoplasmic components which are not naturally linked and which advantageously are chosen to act
15 together cooperatively to produce improved activation of the cell. DNA coding for such recombinant chimeric receptors may be introduced into T-cells or other effector cells in-vivo and/or ex-vivo. Subsequent binding of an effector cell expressing one or more of these chimeric receptors to a target cell elicits signal transduction leading to activation of the effector cell
20 in a process involving clustering or dimerisation of chimeric receptors or allosteric changes in the chimeric receptor or another mechanism for receptor-triggering.

Thus according to one aspect of the invention we provide a method of
25 activating a cell as a result of one type of extracellular interaction between said first cell and a molecule associated with a second target cell characterised in that said first cell is provided with a DNA delivery system comprising DNA coding for one or more recombinant chimeric receptors comprising two or more different cytoplasmic signalling components,
30 wherein said cytoplasmic components are not naturally linked, and at least one is derived from a membrane spanning polypeptide.

The DNA coding for the chimeric receptor(s) is arranged such that when it is expressed, and on the extracellular interaction between the cell and a
35 second target cell, a signal is transduced via the cytoplasmic signalling components to two or more different intracellular signalling messengers.

This results in activation of the cell and elicits a biological response to the target cell. As used herein, cell activation means activation of one or more signal transduction pathways. This may be evidenced by an increase in cell proliferation; expression of cytokines with, for example pro or anti-inflammatory responses; stimulation of cytolytic activity, differentiation or other effector functions; antibody secretion; phagocytosis; tumour infiltration and/or increased adhesion.

The cytoplasmic signalling components are preferably selected such that they are capable of acting together cooperatively. They are "not naturally linked", which term is used herein to denote cytoplasmic signalling components which in nature are not connected to each other on a single polypeptide chain. Particularly useful signalling components include those described hereinafter in relation to other aspects of the invention.

In addition to the cytoplasmic signalling components each recombinant chimeric receptor preferably comprises a binding component capable of recognising a cell surface molecule on a target cell, and a transmembrane component. The DNA coding for these components will additionally code for a signal peptide to ensure that the chimeric receptor(s) once expressed will be directed to the cell surface membrane. All the components may be coded for by a single DNA coding sequence.

Alternatively, each cytoplasmic signalling component may be coded for by two or more separate DNA coding sequences. In this instance each DNA coding sequence may also code for a signal peptide, a transmembrane component and/or a binding component. The binding components may be different, but will generally all be capable of participating in the same type of extracellular event, for example by binding to the same molecule associated with the target cell. In one preference the binding components are the same.

In some of the applications described hereinafter, for example where the binding component is an antibody or an antibody fragment, the DNA coding for the chimeric receptor may comprise two separate DNA coding sequences, one sequence for example coding for part of the binding

component [in the case of an antibody for example a V_H component] linked to the signal peptide, transmembrane and cytoplasmic signalling component(s), and the second sequence coding for the remainder of the binding component [for example a V_L component in the example given].

5

In order to activate a desired cell the DNA coding for the chimeric receptor will first need to be delivered to the cell. Thus according to a second aspect of the invention we provide a DNA delivery system comprising DNA in association with a carrier said DNA coding for a recombinant
10 chimeric receptor capable of one type of extracellular interaction and comprising two or more different cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide.

15 In this aspect of the invention the chimeric receptor may be coded for by a single DNA coding sequence, coding in particular for the two or more different cytoplasmic signalling components. Thus in one preference the invention provides a DNA delivery system comprising DNA in association with a carrier said DNA coding for a recombinant chimeric receptor
20 wherein said DNA codes in reading frame for:

- i) a signal peptide component;
- ii) a binding component capable of recognising a cell surface molecule on a target cell;
- 25 iii) a transmembrane component;
- iv) two or more different cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide, and optionally
- 30 v) one or more spacer regions linking any two or more of said i) to iv) components.

The components of the recombinant chimeric receptor are operatively linked such that the signalling cytoplasmic components are functional in
35 transducing a signal resulting in activation of one or more messenger

systems as a result of recognition of a cell surface molecule on a target cell by the binding component.

Two or more of the components may be linked by one or more spacer regions. The spacer regions may function to facilitate the components adopting the correct conformation for biological activity. The use of a spacer region to link the transmembrane component iii) and the binding component ii) is particularly advantageous.

10 The spacer regions may for example comprise up to 300 amino acids and preferably 20 to 100 amino acids and most preferably 25 to 50 amino acids.

15 Spacers may be derived from all or part of naturally occurring molecules such as from all or part of the extracellular region of CD8, CD4 or CD28; or all or part of an antibody constant region, including the hinge region. All or part of natural spacing components between functional parts of intracellular signalling molecules for example spacers between ITAMS (immunoreceptor tyrosine based activation motifs) may also be used.

20 Alternatively the spacer may be a non-naturally occurring sequence.

The binding component ii) may be any molecule capable of interacting with cell surface molecules and may be chosen to recognise a surface marker expressed on cells associated with a disease state such as for
25 example those associated with virally infected cells; bacterially infected cells; cancer cells, such as the bombesin receptor expressed on lung tumour cells, carcinoembryonic antigen, polymorphic epithelial mucin, and CD33; peptide hormones, adhesion molecules, inflammatory cells present in autoimmune disease, or a T-cell receptor or antigen giving rise to
30 autoimmunity.

Suitable binding components for use in the chimeric receptors of the invention also include all or part of receptors associated with binding to cell surface associated molecules; the T-cell receptor; CD4; CD8; CD28;
35 cytokine receptors e.g. an interleukin receptor, TNF receptor, or interferon receptor e.g. γ -IFN; receptors for colony stimulating factors e.g. GMCSF;

antibodies and antigen binding fragments thereof including for example Fab, Fab', F(ab')₂, single chain Fv, Fv, and V_H or V_L components which may be in association with C_H and C_L domains. The antibodies or fragments may be murine, human, chimeric or engineered human antibodies and fragments. As used herein the term engineered human antibody or fragment is intended to mean an antibody or fragment which has one or more CDR's and one or more framework residues derived from one antibody, e.g. a murine antibody embedded in an otherwise human framework. Such antibodies are well known and may be prepared by a number of methods for example as described in International Patent Specification No. WO91/09967.

Particularly useful binding components include Fab' fragments or, especially, single chain Fv fragments.

When the binding component is an antibody or antibody fragment other than a single chain Fv or V_H or V_L component which contains separate binding chains it will be necessary to include a second separate DNA coding sequence in the delivery system according to the invention to code for the second binding chain. In this instance the first DNA sequence containing the cytoplasmic signalling components and one chain of the antibody or fragment will be coexpressed with the second DNA sequence coding for a signal peptide and the second chain of the antibody or fragment so that assembly of the antibody binding component can occur.

Transmembrane components iii) may be derived from a wide variety of sources such as all or part of the alpha, beta or zeta chain of the T-cell receptor, CD28, CD8, CD4, a cytokine receptor, e.g. an interleukin receptor, TNF receptor, or interferon receptor, or a colony stimulating factor receptor e.g. GMCSF.

The binding and transmembrane components may be linked directly or, preferably, by a spacer region. The spacer region may be one or more of the regions described above. Where more than one region is present, for example two regions, these are preferably different regions, for example

an antibody hinge region linked to all or part of the extracellular region of CD28.

5 The spacer and transmembrane components are advantageously chosen such that they have free thiol groups thereby providing the chimeric receptor with multimerisation, particularly dimerisation capacity. Receptors of this type, especially dimers, are particularly preferred and include those which have CD28 components, the zeta chain of the natural T-cell receptor, and/or antibody hinge sequences.

10 The transmembrane component may or may not be naturally linked to the cytoplasmic component to which it is attached either directly or by means of a spacer.

15 The cytoplasmic signalling components iv) can for example transduce a signal which results in activation of one or more intracellular messenger systems. It is preferred that each of the cytoplasmic components activates a different messenger system. The intracellular messenger systems which may be activated either directly or indirectly include, for example,
20 one or more kinase pathways such as those involving tyrosine kinase, PKC or MAP kinase; G-protein or phospholipase mediated pathways; calcium mediated pathways; and pathways involving synthesis of a cytokine such as an interleukin e.g. IL-2, including NFAT, and cAMP mediated pathways.

25 Examples of suitable cytoplasmic components iv) include, for example those derived from the T-cell receptor such as all or part of the zeta, eta or epsilon chain; CD28; the γ chain of a Fc receptor; or signalling components from a cytokine receptor e.g. interleukin, TNF and interferon
30 receptors, a colony stimulating factor receptor e.g. GMCSF, a tyrosine kinase e.g. ZAP-70, fyn, lyk, Itk and syk; an adhesion molecule e.g. LFA-1 and LFA-2, B29, MB-1, CD3 delta, CD3 gamma, CD5 or CD2. The signalling cytoplasmic components are preferably ITAM containing cytoplasmic components

35

The cytoplasmic signalling components are preferably selected so that they act cooperatively. They may be in any orientation relative to one another. Particularly useful components include all or part of the signalling component of CD28 or the zeta chain of the T-cell receptor.

5

The signal component may be that naturally associated with the binding component or may be derived from other sources.

10

Examples of suitable signal peptide components i) include immunoglobulin signal sequences.

15

The signal component, binding component, transmembrane component, and cytoplasmic components are preferably derived from or based on human sequences.

20

Homologues of the individual components of the chimeric receptor may be used and the invention is to be understood to extend to such use. The term homologue as used herein with respect to a particular nucleotide or amino acid sequence coding for a component of the chimeric receptor represents a corresponding sequence in which one or more nucleotides or amino acids have been added, deleted, substituted or otherwise chemically modified provided always that the homologue retains substantially the same function as the particular component of the chimeric receptor. Homologues may be obtained by standard molecular biology and/or chemistry techniques e.g. by cDNA or gene cloning, or by use of oligonucleotide directed mutagenesis or oligonucleotide directed synthesis techniques or enzymatic cleavage or enzymatic filling in of gapped oligonucleotides.

25

30

Fragments of the individual components may also be used wherein one or more nucleotides has been deleted provided that the fragment retains substantially the same function as the starting component of the chimeric receptor.

35

The DNA for use in this and other aspects of the invention may be obtained from readily available DNA sources using standard molecular

biology and/or chemistry procedures, for example by use of oligonucleotide directed mutagenesis or oligonucleotide directed synthesis techniques, enzymatic cleavage or enzymatic filling in of gapped oligonucleotides. Such techniques are described by Maniatis *et al* in
5 Molecular Cloning, Cold Spring Harbor Laboratory, New York 1989, and in particular in the Examples hereinafter.

The carrier for use in the DNA delivery systems according to the invention may be a vector or other carrier suitable for introduction of the DNA *ex-*
10 *vivo* or *in-vivo* into target cells and/or target host cells. Examples of suitable vectors include viral vectors such as retroviruses, adenoviruses, adenoassociated viruses, EBV, and HSV, and non-viral vectors, such as liposomal vectors and vectors based on DNA condensing agents. Alternatively the carrier may be an antibody. Where appropriate, the
15 vector may additionally include promoter/regulatory sequences and/or replication functions from viruses such as retrovirus LTRs, AAV repeats, SV40 and hCMV promoters and/or enhancers, splicing and polyadenylation signals; EBV and BK virus replication functions. Tissue specific regulatory sequences such as the TCR- α promoter, E-selectin
20 promoter and the CD2 promoter and locus control region may also be used.

Where two or more DNA molecules are used in the DNA delivery system they may be incorporated into the same or different carriers as described
25 above.

For *ex-vivo* use, the DNA delivery system of the invention may be introduced into effector cells removed from the target host using methods well known in the art e.g. transfection, transduction, biolistics, protoplast
30 fusion, calcium phosphate precipitated DNA transformation, electroporation, cationic lipofection, or targeted liposomes. The effector cells are then reintroduced into the host using standard techniques.

A wide variety of target hosts may be employed according to the present
35 invention such as, for example, mammals and, especially, humans.

Examples of suitable effector cells include cells associated with the immune system such as lymphocytes e.g. cytotoxic T-lymphocytes, tumour infiltrating lymphocytes, natural killer cells, neutrophils, basophils or T-helper cells; dendritic cells, B-cells, haematopoietic stem cells, macrophages, monocytes or NK cells. The use of cytotoxic T-lymphocytes is especially preferred.

The DNA delivery system according to the invention is particularly suitable for *in vivo* administration. It may be in one preferred example in the form of a targeted delivery system in which the carrier is capable of directing the DNA to a desired effector cell. Particular examples of such targeted delivery systems include targeted-naked DNA, targeted liposomes encapsulating and/or complexed with the DNA, targeted retroviral systems and targeted condensed DNA such as protamine and polylysine condensed DNA.

Targeting systems are well known in the art and include using, for example, antibodies or fragments thereof against cell surface antigens expressed on target cells *in vivo* such as CD8; CD16; CD4; CD3; selectins e.g. E-selectin; CD5; CD7; CD34; activation antigens e.g. CD69 and IL-2R. Alternatively, other receptor - ligand interactions can be used for targeting e.g. CD4 to target HIV_{gp160} - expressing target cells.

In general the use of antibody targeted DNA is preferred, particularly antibody targeted naked DNA, antibody targeted condensed DNA and especially antibody targeted liposomes. Particular types of liposomes which may be used include for example pH-sensitive liposomes where linkers cleaved at low pH may be used to link the antibody to the liposome. Cationic liposomes which fuse with the cell membrane and deliver the recombinant chimeric receptor DNA according to the invention directly into the cytoplasm may also be used. Liposomes for use in the invention may also have hydrophilic groups attached to their surface to increase their circulating half-life such as for example polyethylene glycol polymers. There are many examples in the art of suitable groups for attaching to liposomes or other carriers; see for example International Patent

91/05546, WO 93/19738, WO 94/20073 and WO 94/22429. The antibody or other targeting molecule may be linked to the DNA, condensed DNA or liposome using conventional readily available linking groups and reactive functional groups in the antibody e.g. thiols, or amines and the like, and in the DNA or DNA containing materials.

Non-targeted delivery systems may also be used and in these targeted expression of the DNA is advantageous. Targeted expression of the DNA may be achieved for example by using T-cell specific promoter systems such as the zeta promoter and CD2 promoter and locus control region, and the perforin promoter.

The aspect of the invention described above advantageously utilises a single DNA sequence to code for the chimeric receptor. It will be appreciated however that the invention may be extended to DNA delivery systems in which the chimeric receptor is coded for by two or more separate DNA coding sequences. Thus in one example, a first and second separate DNA coding sequence may be present in the delivery system each of which codes for components i) to iv) and optionally v) in the same reading frame as described above but which differ from each other in that the cytoplasmic signalling component iv) is not the same. The two DNA coding sequences may each code for more than one signalling component providing that at least one component on the first DNA is different to any other signalling component on the second DNA. As above, the signalling components are advantageously selected to act cooperatively and the remaining components may be any of those previously described for the single DNA embodiment. The binding component iv) coded for by the first DNA will preferably be the same as that coded for by the second DNA. Advantageously the binding component coded by the first DNA will be separated from the transmembrane component by a different spacer region to that coded by the second DNA.

The delivery system may be used *ex vivo* and in a further aspect the invention provides effector cells transfected with a DNA delivery system according to the invention. The effector cells may be any of those

previously described above which are suitable for ex vivo use and are preferably T-cells most preferably cytotoxic T-cells.

- 5 The DNA delivery system may take the form of a pharmaceutical composition. It may be a therapeutic or diagnostic composition and may take any suitable form suitable for administration. Preferably it will be in a form suitable for parenteral administration e.g. by injection or infusion, for example by bolus injection or continuous infusion. Where the composition is for injection or infusion, it may take the form of a suspension, solution or emulsion in an oily or aqueous vehicle and it may contain formulatory agents such as suspending, preservative, stabilising and/or dispersing agents. Alternatively, the composition may be in dry form, for reconstitution before use with an appropriate sterile liquid.
- 10
- 15 If the composition is suitable for oral administration the formulation may contain, in addition to the active ingredient, additives such as: starch - e.g. potato, maize or wheat starch or cellulose - or starch derivatives such as microcrystalline cellulose; silica; various sugars such as lactose; magnesium carbonate and/or calcium phosphate. It is desirable that, if the formulation is for oral administration it will be well tolerated by the patient's digestive system. To this end, it may be desirable to include in the formulation mucus formers and resins. It may also be desirable to improve tolerance by formulating the compositions in a capsule which is insoluble in the gastric juices. It may also be preferable to include the composition in a controlled release formulation.
- 20
- 25

- The DNA delivery system according to the invention is of use in medicine and the invention extends to a method of treatment of a human or animal subject, the method comprising administering to the subject an effective amount of a DNA delivery system described above. The exact amount to be used will depend on the ages and condition of the patient, the nature of the disease or disorder and the route of administration, but may be determined using conventional means, for example by extrapolation of animal experiment derived data. In particular, for ex vivo use the number of transfected effector cells required may be established by ex vivo transfection and re-introduction into an animal model of a range of effector
- 30
- 35

cell numbers. Similarly the quantity of DNA required for *in vivo* use may be established in animals using a range of DNA concentrations.

The DNA delivery system according to the invention may be useful in the treatment of a number of diseases or disorders. Such diseases or disorders may include those described under the general headings of infectious diseases, e.g. HIV infection; inflammatory disease/autoimmunity e.g. rheumatoid arthritis, osteoarthritis, inflammatory bowel disease; cancer; allergic/atopic diseases e.g. asthma, eczema; congenital e.g. cystic fibrosis, sickle cell anaemia; dermatologic, e.g. psoriasis; neurologic, e.g. multiple sclerosis; transplants e.g. organ transplant rejection, graft-versus-host disease; metabolic/idiopathic disease e.g. diabetes.

DNA coding for a chimeric receptor as described herein also forms a feature of the invention, particularly for use in a delivery system described herein.

The invention is further illustrated in the following non-limiting Examples and Figures in which:

- Figure 1 shows: diagrammatic representation of recombinant chimeric receptor constructs cloned into pBluescript SK+
- Figure 2 shows: diagrammatic representation of recombinant chimeric receptor constructs cloned into pBluescript SK+
- Figure 3 shows: oligonucleotide sequences for recombinant chimeric receptor construction
- Figure 4 shows: nucleotide and amino acid sequence of an hCTMO1/CD8/zeta recombinant chimeric receptor
- Figure 5 shows: nucleotide and amino acid sequence of an hCTMO1/CD8/zeta-CD28 recombinant chimeric receptor fusion
- Figure 6 shows: nucleotide and amino acid sequence of an hCTMO1/CD8/CD28 recombinant chimeric receptor
- Figure 7 shows: nucleotide and amino acid sequence of an CTMO1/G1/zeta recombinant chimeric receptor

- Figure 8 shows: nucleotide and amino acid sequence of an hCTMO1/G1/zeta-CD28 recombinant chimeric receptor fusion
- Figure 9 shows: nucleotide and amino acid sequence of an hCTMO1/h/CD28 recombinant chimeric receptor
- 5 Figure 10 shows: histogram representation of IL2 production by cell lines TB3.2, 3.13 and 3.24 when stimulated with an anti-idiotypic antibody alone or in combination with an anti-CD28 antibody
- 10 Figure 11 shows: histogram representation of the production of IL2 by cell line TB3.13 when stimulated with antigen expressing tumour cells, shown with and without co-stimulation using an anti-CD28 antibody.
- Figure 12 shows: histogram representation of IL-2 production by HGT1.2 and HGT1.4 in response to various stimuli
- 15 Figure 13 shows: histogram representation of IL-2 production by HGT2.4 incubated with various combinations of antibodies.
- Figure 14 shows: schematic representation of recombinant chimeric receptor constructs.
- 20 Figure 15 shows: schematic representation of recombinant chimeric receptor constructs
- Figure 16 shows: schematic representation of recombinant chimeric receptor constructs.
- 25 Figure 17 shows: schematic representation of recombinant chimeric receptor constructs
- Figure 18 shows: histogram representation of levels of expression of CD28 chimeras in Jurkat cells
- 30 Figure 19 shows: histogram representation of IL-2 production by Jurkat cells expressing two different chimeric receptors in response to target cells.

Figure 20 shows: Graph showing Cytolysis of target cells by CD8+ve human CTL cells infected with recombinant adenoviruses

5 **EXAMPLE 1**

Construction of chimeric receptor genes

Each component of the chimeric receptor constructs was either PCR cloned or PCR assembled by standard techniques (PCR Protocols, Innis et al, 1990, Academic Press inc.) and sub-cloned in a cassette format into
10 pBluescript SK+ (Stratagene), see figure 1, 2, 2b and 2c. Oligonucleotides are described in Figure 3.

1. **Single chain Fv cassettes**

hCTMO1

15 An scFv from the engineered human CTMO1 antibody was constructed as follows. Leader sequence and hCTMO1 VI was PCR cloned from plasmid pAL 47 (International Patent Specification No. WO 93/06231) with oligos R6490 and R6516 (Oligo sequences are shown in Figure 3). R6490 introduces 5' Not I and Hind III sites and R6516 forms part of the
20 (Gly4Ser)₅ linker. hCTMO1 Vh was PCR cloned from plasmid pAL 52 (WO 93/06231) with oligos R6515 (forms part of linker) and R6514 (introduces 3' Spe I site. Leader / VI and Vh fragments were then PCR spliced together and the PCR product was restricted with Not I and Spe I and sub-cloned into pBluescript SK+.

25

hP67.6

An scFv from another engineered human antibody, hP67.6, engineered according to WO91/09967, was similarly prepared and subcloned into
30 pBluescript SK+.

30

2. **CD8 hinge spacer cassette**

The CD8 hinge spacer for hCTMO1 TCR Zeta chimeric receptor and hCTMO1 TCR Zeta-CD28 fusion chimeric receptor (which includes a small part of 5' Zeta) was PCR assembled using overlapping oligos:
35 R6494, R6495, R6496 and R6497. The CD8 hinge spacer for hCTMO1 CD28 chimeric receptor was PCR assembled using overlapping oligos:

R6494, R6495, R6496 and R6506. Both PCR products were restricted with Spe I and BamH I and sub-cloned into pBluescript SK+.

3. Human TCR Zeta cassette

- 5 Human Zeta transmembrane and intracellular components were PCR cloned from human leukocyte cDNA (Clonotech) with oligos R6488 (introducing a 5' BamH I site) and R6489 (introducing a 3' EcoR I site). PCR product was restricted with BamH I and EcoR I and sub-cloned into pBluescript SK+.

10

4. Human CD28 cassette

- Human CD28 transmembrane and intracellular components were PCR cloned from human leukocyte cDNA (Clonotech) with oligos P3240 (introducing a 5' BamH I site) and P3241 (introducing a 3' EcoR I site).
15 PCR product was restricted with BamH I and EcoR I and sub-cloned into pBluescript SK+.

5. Hinge-CD28 cassette

- Human CD28 extracellular, transmembrane and intracellular components
20 were PCR cloned from human leukocyte cDNA (Clonotech) with oligos S0146 (introducing a 5' Spe I site) and P3241 (introducing a 3' EcoR I site). S0146 also constitutes residues 234 to 243 of human IgG1 hinge. The product of the PCR reaction was digested with restriction enzyme Spe1 and EcoR1 and sub-cloned into pBluescriptSK+.

25

6. Zeta-CD28 fusion cassette

- The 3' end of Zeta, starting at a naturally occurring Sty I site and the intracellular component of human CD28 were PCR assembled such that the Zeta stop codon was removed and an inframe fusion protein would be
30 translated. PCR assembly carried out with overlapping oligos: P3301, P3302, P3303, P3304, P3305 and P3306. PCR product was restricted with Sty I and EcoR I and sub-cloned into pBluescriptSK+ containing the hCTMO1 TCR Zeta chimeric receptor construct, replacing the 3' end of Zeta.

35

7. Human IgG1 spacer cassette

Human IgG1 hinge, CH2 and CH3 were PCR cloned from IgG1 cDNA clone (A. Popplewell) with oligos S0060 (introducing a 5' Spe I site) and S0061 (introducing residues L, D, P, and K constituting a 3' BamH I site). PCR product was restricted with Spe I and BamH I and sub-cloned into pBluescriptSK+.

8. **h.28 spacer cassette**

Human IgG1 hinge and part of human CD28 extracellular component were PCR cloned from a scFv/h/CD28 plasmid with oligos T4057 and T4058. T4057 introduces a 5' Spe I site and T4058 introduces residues L, D, P, and K constituting a 3' BamH I site. PCR product was restricted with Spe I and BamH I and sub-cloned into pBluescriptSK+.

9. **CD28-Zeta fusion cassette**

Human CD28 transmembrane and intracellular components were PCR cloned from a scFv/h/CD28 plasmid with oligos T7145 and T4060. T7145 introduces residues L, D, P, and K constituting a 3' BamH I site. T4060 comprises a 3' overhang compatible with the 5' end of human Zeta intracellular component.

Human Zeta intracellular component was PCR cloned from a scFv/G1/Zeta plasmid with oligos T4387 and S4700. T4387 comprises a 5' overhang compatible with the 3' end of human CD28 intracellular component. S4700 introduces a 3' EcoR I site.

CD28 transmembrane and intracellular components were then PCR spliced to Zeta intracellular component with oligos T7145 and S4700. PCR product was restricted with BamH I and EcoR I and sub-cloned into pBluescriptSK+.

10. **CD28-Zeta-CD28 fusion cassette**

A Pst I restriction site in human Zeta was used to subclone the 3' end of Zeta intracellular component and the CD28 intracellular component on a Pst I to EcoR I fragment from the Zeta-CD28 fusion cassette into the CD28-Zeta fusion cassette, replacing the 3' end of Zeta. This generates a CD28-Zeta-CD28 fusion cassette with a 5' BamH I site and 3' EcoR I site.

All of the above cassettes were completely sequenced (Applied Biosystems, Taq DyeDeoxy Terminator Cycle Sequencing, Part Number 901497) in pBluescriptSK+ prior to cloning into the expression vectors.

- 5 These cassettes were assembled to construct chimeric receptors with the specificity of the engineered human antibodies hCTMO1, directed against human polymorphic epithelial mucin (PEM) or hP67.6, directed against human CD33, by assembling the appropriate cassettes using standard molecular biology techniques. The following chimeric receptors were
10 constructed; see Table 2 and Figures 14 - 17 in which potential di-sulphide bonds are indicated by a horizontal line between the two sub-units (not all di-sulphide bonds may form in 100% of the molecules).

1) **scFv / CD8 / Zeta Chimeric Receptor (Figure 14)**

- 15 The scFv / CD8 / Zeta chimeric receptor consists of a single chain Fv (scFv) linked to an extracellular spacer in the form of part of human CD8 hinge, linked to the extracellular, transmembrane and intracellular components of the human T-cell receptor Zeta chain (TCR).
- 20 The scFv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 98 to 142 of the hinge region of human CD8 (Zamoyska *et al* : Cell 43,153-163, 1985
25). This is linked to residues 6 to 142 of human TCR Zeta comprising extracellular (part), transmembrane and intracellular regions (Weissman *et al* : PNAS 85, 9709-9713, 1988. Moingeon *et al*: Eur. J. Immunol. 20, 1741-1745, 1990).

30 2) **scFv / CD8 / CD28 Chimeric Receptor (Figure 14)**

- The CD8 hinge/CD28 chimeric receptor consists of a scFv linked to an extracellular spacer in the form of part of human CD8 hinge, linked to the transmembrane and intracellular component of human CD28.
- 35 The scFv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅

linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 98 to 142 of the hinge region of human CD8 (Zamoyska *et al* : Cell 43 153-163, 1985). This is linked to residues 132 to 202 of human CD28 comprising the transmembrane and intracellular components (Aruffo & Seed : PNAS 84, 8573-8577).

3) **scFv /CD8 / Zeta-CD28 Fusion Chimeric Receptor (Figure 14)**

The scFv /CD8 / Zeta-CD28 Fusion chimeric receptor consists of a single chain Fv linked to an extracellular spacer in the form of part of human CD8 hinge, linked to the extracellular, transmembrane and intracellular components of human TCR Zeta fused to the intracellular component of human CD28.

The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extra cellular spacer consists of residues 98 to 142 of the hinge region of human CD8 (Zamoyska *et al* : Cell, 43,153-163, 1985). This is linked to residues 6 to 142 of human TCR Zeta comprising extracellular (part), transmembrane and intracellular components (Weissman *et al* : PNAS 85,9709-9713, 1988 Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990). This is linked to residues 162 to 202 comprising the intracellular component of human CD28.

4) **scFv / G1 / Zeta Chimeric Receptor (Figure 15)**

The scFv / G1 / Zeta chimeric receptor consists of a single chain Fv linked to an extracellular spacer comprising human IgG 1 hinge, CH2 and CH3, linked to the transmembrane and intracellular regions of human TCR Zeta.

The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge, 244 to 360 of CH2 and 361 to 478 of

CH3 (Kabat *et al* Sequences of proteins of immunological interest, 1987). This is linked to residues 6 to 142 of human TCR Zeta comprising extracellular (part), transmembrane and intracellular regions (Weissman *et al* : PNAS 85 ,9709-9713, 1988. Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990).

5) **scFv / G1 / Zeta-CD28 fusion Chimeric Receptor (Figure 15)**

The scFv / G1 / Zeta chimeric receptor consists of a single chain Fv linked to an extracellular spacer comprising human IgG 1 hinge, CH2 and CH3, linked to the transmembrane and intracellular regions of human Zeta fused to the intracellular region of human CD28.

The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge, 244 to 360 of CH2 and 361 to 478 of CH3 (Kabat *et al* Sequences of proteins of immunological interest, 1987). This is linked to residues 6 to 142 of human TCR Zeta comprising extracellular (part), transmembrane and intracellular regions (Weissman *et al* : PNAS 85 ,9709-9713, 1988. Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990).

This is linked to residues 162 to 202 comprising the intracellular component of human CD28 (Aruffo & Seed : PNAS 84, 8573-8577).

6) **scFv / h / CD28 Chimeric Receptor (Figure 15)**

The scFv / h / CD28 chimeric receptor consists of a single chain Fv linked to an extracellular spacer consisting of human IgG1 hinge and part of the extracellular region of human CD28, linked to the transmembrane and intracellular regions of human CD28.

The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge and residues 118 to 134 of human CD28.

This is linked to residues 135 to 202 of human CD28 comprising the transmembrane and intracellular regions (Aruffo & Seed : PNAS 84, 8573-8577).

5 7) **scFv / G1 / CD28 Chimeric Receptor (Figure 16)**

The scFv / G1 / Zeta chimeric receptor consists of a single chain Fv linked to an extra cellular spacer comprising human IgG 1 hinge, CH2 and CH3, linked to the transmembrane and intracellular regions of human CD28.

- 10 The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge, 244 to 360 of CH2 and 361 to 478 of
- 15 CH3 (Kabat *et al* Sequences of proteins of immunological interest, 1987). This is linked via residues L, D, P and K to residues 135 to 202 comprising the transmembrane and intracellular components of human CD28 (Aruffo & Seed : PNAS 84, 8573-8577).

20 8) **scFv / G1 / CD28 -Zeta fusion Chimeric Receptor (Figure 16)**

The scFv / G1 / Zeta chimeric receptor consists of a single chain Fv linked to an extracellular spacer comprising human IgG 1 hinge, CH2 and CH3, linked to the transmembrane and intracellular regions of human CD28 fused to the intracellular region of human Zeta.

- 25 The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues
- 30 234 to 243 of human IgG1 hinge, 244 to 360 of CH2 and 361 to 478 of CH3 (Kabat *et al* Sequences of proteins of immunological interest, 1987). This is linked via residues L, D, P and K to residues 135 to 202 comprising the transmembrane and intracellular components of human CD28.

- 35 This is linked to residues 31 to 142 of human TCR Zeta, the intracellular region (Weissman *et al* : PNAS 85 ,9709-9713, 1988. Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990).

9) scFv / G1 / CD28 -Zeta -CD28 fusion Chimeric Receptor (Figure 16)

5 The scFv / G1 / Zeta chimeric receptor consists of a single chain Fv linked to an extracellular spacer comprising human IgG 1 hinge, CH2 and CH3, linked to the transmembrane and intracellular regions of human CD28 fused to the intracellular region of human Zeta fused to the intracellular region of CD28.

10 The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge, 244 to 360 of CH2 and 361 to 478 of CH3 (Kabat *et al* Sequences of proteins of immunological interest, 1987).
15 This is linked via residues L, D, P and K to residues 135 to 202 comprising the transmembrane and intracellular components of human CD28.

This is linked to residues 31 to 142 of human TCR Zeta, the intracellular region (Weissman *et al* : PNAS 85 ,9709-9713, 1988. Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990).
20 This is linked to residues 162 to 202 comprising the intracellular component of human CD28.

10) scFv / h.28 / Zeta Chimeric Receptor (Figure 17)

25 The scFv / h / CD28 chimeric receptor consists of a single chain Fv linked to an extracellular spacer consisting of human IgG1 hinge, part of the extracellular region of human CD28 and 4 amino acid residues, linked to the transmembrane and intracellular regions of human TCR Zeta.

30 The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge and residues 118 to 134 of human CD28.

35 This is linked via residues L, D, P and K to residues 10 to 142 of human TCR Zeta comprising the transmembrane and the intracellular region

(Weissman *et al* : PNAS 85 ,9709-9713, 1988. Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990).

11) **scFv / h.28 / Zeta-CD28 fusion Chimeric Receptor (Figure 17)**

5 The scFv / h / CD28 chimeric receptor consists of a single chain Fv linked to an extracellular spacer consisting of human IgG1 hinge, part of the extracellular region of human CD28 and 4 amino acid residues, linked to the transmembrane and intracellular regions of human Zeta fused to the intracellular region of human CD28.

10

The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues
15 234 to 243 of human IgG1 hinge and residues 118 to 134 of human CD28. This is linked via residues L, D, P and K to residues 10 to 142 of human TCR Zeta comprising transmembrane and intracellular regions (Weissman *et al* : PNAS 85 ,9709-9713, 1988. Moingeon *et al*:Eur. J. Immunol. 20, 1741-1745, 1990).

20 This is linked to residues 162 to 202 comprising the intracellular component of human CD28.

12) **scFv / h.28 / CD28-Zeta fusion Chimeric Receptor (Figure 17)**

25 The scFv / h / CD28 chimeric receptor consists of a single chain Fv linked to an extracellular spacer consisting of human IgG1 hinge, part of the extracellular region of human CD28 and 4 amino acid residues, linked to the transmembrane and intracellular regions of human CD28 fused to the intracellular region of human Zeta.

30 The single chain Fv consists of the leader sequence and variable component of the light chain of the engineered human antibody linked via a (Gly4Ser)₅ linker to the variable component of the heavy chain of the engineered human antibody. The extracellular spacer consists of residues 234 to 243 of human IgG1 hinge and residues 118 to 134 of human CD28.

35 This is linked via residues L, D, P and K to residues 135 to 202 comprising the transmembrane and intracellular components of human CD28.

This is linked to residues 31 to 142 of human TCR Zeta, the intracellular region (Weissman *et al*: PNAS 85, 9709-9713, 1988. Moingeon *et al*: Eur. J. Immunol. 20, 1741-1745, 1990).

- 5 Table 1 shows a number of preferred recombinant chimeric receptors which may be made in an analogous way by following the above teaching and methods.

- 10 Table 2 gives details of the chimeric receptor constructs and cell line nomenclature used.

EXAMPLE 2

Analysis of hCTMO1-chimeric receptor constructs expressed in Jurkat cells

- 15 Chimeric receptor constructs were sub-cloned from pBluescriptSK+ into the expression vectors pEE6hCMV.ne and pEE6hCMV.gpt (Bebbington (1991), Methods 2, 136-145) on a Hind III to EcoR I restriction fragment. The hCTMO1/CD8/ Zeta chimeric receptor construct was cloned into pEE6hCMVne and the hCTMO1 / CD8 /CD28 and hCTMO1 Zeta-CD28
20 fusion chimeric receptor constructs were cloned into pEE6hCMVgpt.

- Plasmids were linearised and transfected into Jurkat E6.1 cells (ECACC) by electroporation using a Bio-Rad Gene Pulser using the method of Rigley *et al* (J. Immunol. (1995) 154, 1136-1145). Chimeric - receptor
25 expressing colonies were selected in media either containing the drug G418 (2 mg/ml) for Neo vectors or Mycophenolic acid for Gpt vectors as described (Rigley *et al* *ibid.*). After approximately four weeks colonies were visible. Colonies were screened by analysis of surface expression of single chain Fv.

30

Antibodies

- Anti-idiotypic antibodies are purified antisera from rabbits immunised with hCTMO1. Anti-Id antibodies were purified initially on Protein A-Sepharose, absorbed out against human IgG-Sepharose and finally
35 affinity purified on hCTMO1. OKT3 recognises an extracellular component of human CD3 ϵ (ATCC). Anti-CD28 used in these

experiments was a rat IgG2b monoclonal antibody (clone YTH 913.12) directed against the extracellular component of human CD28 (Cymbus Bioscience). FITC labelled donkey anti-rabbit Ig recognises rabbit heavy and light chains (Jackson Research Laboratories).

5

Analysis of surface expression of scFv

Approximately 5×10^5 cells were stained with saturating concentrations of anti-idiotypic (10 µg/ml), then incubated with fluorescein-conjugated donkey anti-rabbit antibody. Fluorescence was analysed by a FACScan
10 cytometer (Beckton Dickinson).

Anti-Id stimulation

1 $\times 10^6$ Jurkat transfectants were incubated in a 96 well plate (Nunc) previously coated with / without a saturating concentration of anti-idiotypic
15 antibody at 37°C / 5% CO₂ in non-selective media. Additional stimuli of anti-CD28 and OKT3 were added in solution to a final concentration of 5 µg/mL. After 18 to 20 hours cells were centrifuged and supernatant assayed for human IL-2 (Quantikine kit, R & D Systems).

Antigen expressing cell stimulation

20 1 $\times 10^6$ Jurkat transfectants were incubated with 1 $\times 10^5$ MCF-7 cells (P.E.M. antigen expressing) in a 96 well plate (Falcon) overnight at 37°C / 5% CO₂.

25 Additional stimulus of anti-CD28 was added in solution to a final concentration of 5 µg/mL. After 18 to 20 hours cells were centrifuged and supernatant assayed for human IL-2 (Quantikine kit, R & D Systems).

RESULTS

Cross-linking the T-cell receptor with anti-CD3 antibodies can be used to stimulate human T-cell lines such as Jurkat E6.1 to produce cytokines including IL-2. The expression of IL-2 can be further enhanced by co-stimulation by means of antibodies to the CD28 cell surface molecule in
35 this cell line. This therefore provides a convenient model system to

evaluate chimeric receptors for the ability to deliver signals which are co-stimulatory for T-cell activation.

1. Enhancement of IL2 production by a Jurkat E6.1 cell line transfected with an hCTM01 scFv-CD8- TCR ζ chimeric receptor (plasmid pTB3 in response to antigen or anti-idiotypic antibody by co-stimulation with an anti-CD28 antibody.

The cell lines TB 3.2, 3.13 and 3.24 were stable cell lines derived from Jurkat E6.1 transfected with CTM01hscFv/CD8/Zeta. Figure 10 shows IL2 production by these cell lines when stimulated with an anti-CTM01 idiotypic antibody alone or in combination with an anti-CD28 antibody. In each case the co-stimulation with anti CD-28 results in a greater than 2-fold stimulation of IL2 production compared to stimulation with anti-CTM01 idio type antibody alone. Incubation of these cell lines with anti-CD28 alone did not result in stimulation of IL2.

Figure 11 shows the production of IL2 by one of the above cell lines (TB 3.13) when stimulated with antigen expressing tumour cells. As in figure 10 this is shown with and without co-stimulation using anti-CD28 antibody and indicates that co-stimulation can enhance IL-2 production when stimulation of the chimeric receptor is mediated by antigen.

2. Construction and testing of a chimeric receptor designed to generate a response analogous to CD28 stimulation on interaction with the extracellular scfv component.

Having established that co-stimulation via the CD28 molecule could result in enhancement of the response of a T cell transfectant to a tumour associated antigen a chimeric receptor incorporating the CD28 transmembrane and cytoplasmic components was constructed. This hCTM01/CD8/CD28 chimeric receptor (pHMF332) (HGT1) was transfected into Jurkat E6.1 cells to generate stable cell lines. Two of these lines HGT 1.2 and 1.4 were incubated in the presence of various combinations of stimulating antibodies as shown in figure 12 (see materials and methods for experimental procedure), and anti-idiotypic antibody was used to stimulate the chimeric receptor.

Incubation of the cell lines shown with an anti-CD3 antibody resulted in a low level of IL2 production. This stimulation could be enhanced by co-stimulating with an anti-CD28 antibody (column 5 figs. 12a and 12b).

- 5 Incubation with the anti-CD28 alone as expected did not result in IL2 production.

Similarly incubation with the anti-idiotypic antibody alone (stimulating the chimeric CD28 receptor) resulted in no IL2 production. However, by
10 analogy with the combined anti-CD3 and anti-CD28 stimulation, incubation with anti-CD3 and anti-idiotypic resulted in IL2 production enhanced over CD3 stimulation alone. This demonstrates that a chimeric receptor could be constructed that responds via stimulation of extracellular scFv to
15 activate.

3. **Provision of both primary and accessory stimulation in the same effector cell.**

In order to provide both primary (for example TCR ζ mediated) and co-
20 stimulatory (for example CD28 mediated) activation of the effector cell via interaction of a chimeric receptor with a defined ligand or antigen a fusion receptor incorporating two different signalling components was constructed. This chimeric receptor hCTM01/CD8/TCRZeta-CD28 (pHMF334) was transfected into Jurkat E6.1 cells and stable lines
25 selected. One of these lines (HGT 2.4) was incubated with various combinations of antibodies and IL2 production measured (see Fig. 13).

The anti-CD3 and anti-CD28 antibodies individually and in combination resulted in a similar relative stimulation of IL2 production to that seen with
30 the other transfected cell lines. However, with the construct HGT2 the anti-idiotypic antibody alone resulted in a level of IL2 production greater than achieved with the combined anti-CD3 and anti-CD28 antibodies. Furthermore, the stimulation achieved with the single anti-idiotypic interaction could not be enhanced by further co-stimulation with anti-CD3,
35 anti-CD28 or combinations of these.

EXAMPLE 3**Analysis of single gene hP67.6-chimeric receptor constructs expressed in Jurkat cells**

5 In order to confirm the results obtained with the hCTMO1 fusion receptor for a different antibody scFv, and to evaluate additional fusion receptors, a number of different chimeras based on the hP67.6 scFv were introduced into Jurkat cells.

10 Chimeric receptor constructs hP67.6 / G1 / Zeta (HGT16), hP67.6 / G1 / Zeta-CD28 (HGT17), hP67.6 / G1 / CD28-Zeta (HGT21), hP67.6 / G1 / CD28-Zeta-CD28 (HGT26), hP67.6 / h.28 / Zeta-CD28 (HGT20) and hP67.6 / h.28 / CD28-Zeta (HGT22) chimeric receptor constructs were sub-cloned from pBluescriptSK+ into the expression vector pEE6hCMV.ne as described in Example 2. Expression plasmids were transfected into
15 Jurkat E6.1 and permanent cell lines expressing chimeric receptors on their cell surfaces were identified as described above (Example 2) but using a purified rabbit anti-p67.6 idiotype antiserum prepared as described for hCTMO1 anti-idiotype. Alternatively, cells were stained with purified recombinant CD33 extracellular domain conjugated to FITC (10 µg/ml)
20 and analysed directly on the cytometer.

Western blot analysis was carried out on representative clones for each construct to confirm that chimeric receptors of the expected size were expressed. Approximately 10^7 cells were lysed in lysis buffer (1%
25 NP40, 150mM NaCl, 10mM NaF, 0.4mM EDTA, 1mM Na vanadate, 1 mg/ml Pefabloc, 10 µg/ml Pepstatin, 10 µg/ml Leupeptin, 20 µg/ml Aprotinin) and samples subjected to SDS-PAGE with or without reduction of cystine residues with β-mercaptoethanol. Western blots were probed with rabbit anti-P67.6 idiotype followed by horseradish - peroxidase (HRP)
30 conjugated donkey anti-rabbit Ig or HRP-conjugated rabbit anti-human Fc antisera according to standard techniques.

A comparison of the apparent molecular weights of the chimeric receptors in reduced and non-reduced samples indicated that the zeta-chain
35 chimera in cell line HGT16.1 and the fusion receptor in HGT17.39 were present as di-sulphide linked homodimers. The CD28 chimera in HGT14.1

is present as approximately 50% disulphide-linked homodimers and approximately 50% of the molecules are not disulphide linked. At least 50% of molecules are disulphide - linked in the case of the fusion receptors in HGT20, HGT21 and HGT22 cell lines.

5

A panel of independent transfectant clones for each construct were analysed for IL-2 production in response to cells which express CD33 (HL60 cells) or are CD33 negative (eg Jurkat E6.1). It is important to analyse a number of clones expressing each construct since individual clones vary substantially in the level of expression of chimeric receptor. Moreover, even clones expressing similar levels of receptor show different capacities to produce IL-2. Each transfectant was mixed with an equal number of target cells (eg 10^5 cells of each cell type per well of a 96-well plate) and co-cultured for approximately 20 h. The concentration of IL-2 in the supernatant was then determined using a Quantikine human IL-2 ELISA (R&D Systems).

Cell lines containing construct HGT 16 produce levels of IL-2 in response to HL60 cells of up to approximately 200 pg/ml and do not produce detectable IL-2 when stimulated with CD33 - negative cells. Cell lines expressing fusion receptors HGT17, 20, 21, 22 and 26 also produce IL-2, specifically in response to CD33 positive target cells, indicating that the zeta-chain signalling capacity is intact in the fusion proteins. In fact cells expressing the fusion receptors at comparable levels on the cell surface produce on average more IL-2 in response to HL60 cells than HGT16 cell lines (from 50% more to 7-fold more), consistent with their capacity to provide both primary and co-stimulatory signals.

The function of the CD28 signalling domain can be confirmed by assaying for recruitment of downstream signalling components to the CD28 intracellular domain in response to receptor ligand binding. The association of the regulatory (p85) sub-unit of PI3-kinase with phosphorylated ITAM motifs of the sequence YMXM (single-letter amino acid code) in the CD28 intracellular domain in response to CD28 stimulation is well documented (eg Stein et al., 1994 Mol. Cell. Biol. 14: 3392-3402). CD28 also associates specifically with the tyrosine kinase ITK

on activation (August et al. 1994 Proc. Natl. Acad. Sci. USA 91: 9347-9351).

Association of p85 with the receptor chimeras is analysed by immunoprecipitation of the receptor and detection of bound p85 protein by Western blotting as follows. Approximately 5×10^7 cells are washed once with PBS and activated in 0.5 ml PBS containing 10 μ g/ml rabbit anti-P67.6 idiotype antibody at 37°C for various times from 0 - 10 mins. Cells are then washed twice with ice-cold PBS and lysed in 1 ml lysis buffer as described above. Lysates are centrifuged at 15000 rpm in an Eppendorf micro-centrifuge for 10 min. and the supernatants immunoprecipitated with 100 μ l protein A - sepharose beads (Pharmacia) at room temperature for 30 min. (This immunoprecipitation procedure also serves to immunoprecipitate chimeric receptors containing antibody constant regions from cells which have not been stimulated with anti-idiotype antibody to act as a negative control). The beads are then washed 3 times with fresh lysis buffer, resuspended in 50 μ l SDS loading buffer and subjected to SDS-PAGE and Western blotting. Blots are probed with mouse anti-p85 monoclonal antibody and HRP-conjugated rabbit anti-mouse Ig according to standard techniques.

This showed that p85 can associate with fusion receptors but not with the zeta chain chimera in cell line HGT16.1 thus confirming that p85 associates specifically with CD28 and not zeta and that CD28 signalling is retained in fusion chimeras.

Association of ITK with CD28 intracellular components is detected using published methods (August et al. 1994 Proc. Natl. Acad. Sci. USA 91: 9347-9351).

EXAMPLE 4**Expression of two hP67.6 - chimeric receptors in the same cell .**

In order to express both a zeta chimeric receptor and a CD28 co-stimulatory receptor chimera in the same cell, stably transfected Jurkat cell
5 lines expressing CD28 receptor chimeras were infected with recombinant adenovirus encoding the hP67.6 / G1 / Zeta chimeric receptor.

The hP67.6/h.28/CD28 construct was sub-cloned into pEE6hCMV.gpt and transsfectated into Jurkat E6.1 cells as described in Example 2. Cell line
10 HGT14.1 is a Jurkat transfectant expressing this construct. The hP67.6/G1/CD28 construct was cloned into pEE6hCMV.ne and Jurkat clones HGT23.11 and HGT23.16 expressing this construct were isolated as in Example 2. The levels of expression of the CD28 chimeras on the surface of the transfected cells, determined by FAC-analysis with FITC-
15 CD33 as described in Example 3, is shown in Figure 18.

In order to transiently express a uniform amount of the zeta-chain chimera hP67.6/G1/ zeta in each of these CD28-chimera cell lines, a recombinant adenovirus vector expressing the zeta chimera was constructed as
20 follows. The hP67.6/G1/zeta coding sequence from pHMF342 (Example 1 and Table 2) was excised as a Not1 - Kpn1 fragment and inserted into the adenovirus-5 transfer vector pAL119 (provided by G. Wilkinson, Department of Medicine, University of Wales, Cardiff; unpublished) between the Not1 and BamH1 sites, after insertion of a Kpn1 - BamH1
25 adaptor oligonucleotide, to form pAL119-342. In this plasmid, the chimeric receptor coding sequences are expressed under the control of the hCMV-MIE promoter-regulatory region and polyadenylation signal (Wilkinson and Akrigg 1992 Nucl. Acids Res.20: 2233-2239).

30 Suitable alternative adenovirus transfer vectors containing the hCMV-MIE promoter include pCA3 and pCA4 (Hitt et al. 1995 in Methods in Molecular Genetics, K.W. Adolph (ed) Academic Press, Orlando.) Alternative adenovirus transfer vectors can be used such as pAC (Gerard and Meidell
1995 In DNA Cloning: a practical approach (2nd edition) Volume 4 ed
35 Glover and Hames, IRL Press) which does not contain a promoter. In this case, one of many other heterologous promoters, such as the RSV-LTR

promoter or T-cell specific promoters, may be introduced upstream of the chimeric receptor coding sequence prior to insertion into the transfer vector. Additional RNA processing signals are also desirable, such as a polyadenylation signal (eg from SV40 Virus) and an intron (e.g. from the hCMV-MIE gene) (Bebbington (1991), Methods 2, 136-145).

Approximately 5 μ g pAl119-342 was co-transfected with 5 μ g pJM17 (Microbix Biosystems Inc., McGrory et al. 1988 Virology 163: 614-617) into the human embryonic kidney cell line, 293 (ATCC CRL 1573) by calcium phosphate-mediated transfection, according to standard procedures for construction of adenovirus recombinants (Lowenstein et al 1996 in Protocols for gene transfer in Neuroscience, P.R. Lowenstein and L.W. Enquist (eds) Wiley and Sons). This generated recombinant virus RAd160 containing the chimeric receptor cDNA under the control of hCMV - MIE gene regulatory regions. Large scale preparations of RAd160 were prepared (Lowenstein et al *ibid.*) with titres of greater than 10^{10} pfu/ml and stored at -70°C in small aliquots.

Recombinant adenoviruses containing coding sequences for CD28 chimeric receptors are prepared in the same way after insertion of the desired coding sequence into pAl119 or another adenovirus transfer vector.

RAd160 was added to Jurkat E6.1 cells or transfectants expressing CD28 receptor -chimeras at a multiplicity of infection (MOI) of up to 400 pfu/cell with 2 μ g/ml DEAE - Dextran and incubated for 24h at a cell concentration of 10^6 cells/ml in the presence of virus. Samples of cells were infected with a recombinant adenovirus expressing an irrelevant β -galactosidase protein RAd35 (Wilkinson and Akrigg 1992 Nucl. Acids Res.20: 2233-2239) in the same way to act as a negative control. Infected cells were then washed once in fresh growth medium, expanded in culture for a further 6 days and assayed for IL-2 production in response to target cells. The results are shown in Figure 19. Jurkat cells infected with RAd160 produce essentially undetectable levels of IL-2 in response to HL60-cell stimulation (less than 10 pg/ml) unless co-stimulated with 10 μ g/ml anti-CD28 antibody 15E8 (Caltag) which leads to low levels of IL-2 production

specifically in response to HL60 cells and not in response to a cell line which does not express human CD33, the murine SP2/0 cell line. In contrast, RAd160-infected HGT14.1 cells, which express a CD28 chimeric receptor, produce significant levels of IL-2 specifically in response to HL60 target cells even in the absence of anti-CD28 antibody. This indicates that the CD28-chimeric receptor hP67.6/h.28/CD28 is able to contribute the requisite co-stimulation to the zeta chimera. Cell lines expressing the alternative CD28 chimeric receptor, hP67.6/G1/CD28, 23.11 and 23.16 show markedly reduced levels of IL-2 production compared with 14.1. Indeed, 23.16, the cell line expressing the highest level of this CD28 chimera produces no detectable IL-2 at all. The CD28 signalling pathway was shown to be intact in this cell line since stimulation through CD3 (using anti-CD3 antibody) in 23.16 yields very high levels of IL-2 (results not shown). Thus the signalling defect in cell lines expressing the hP67/G1/CD28 chimera appears to be due to interference with zeta-chain signalling. The mechanism responsible is likely to be related to the use of the same extracellular domain in the zeta and CD28 chimeric receptors. This will allow heterodimerisation of the two receptors and this appears to interfere with zeta-chain signalling. This hypothesis is supported by the fact that 23.16, expressing high levels of the CD28 chimera, shows greater interference with zeta-chain signalling than 23.11, expressing very low levels of the CD28 chimera (Figure 18).

This experiment shows that it is possible to use the same scFv region to stimulate two chimeric receptor molecules in the same cell, one to provide a primary stimulus in response to antigen and the other receptor to provide a co-stimulatory signal. This leads to efficient IL-2 production specifically in response to antigen - expressing target cells provided that the two receptors are prevented from heterodimerisation, for instance by using different dimerisation domains on the two receptors. It is envisaged that additional pairs of dimerisation domains will be compatible. For instance the scFv/h.28/zeta chimeric receptor (Example 1; Figure 17) could provide the primary signal and the scFv/G1/CD28 receptor (Example 1; Figure 16) would provide the co-stimulatory signal.

EXAMPLE 5**Identification of additional co-stimulatory cell-surface receptors using anti-receptor antibodies.**

5 5×10^5 HGT16.1 cells expressing the hP67.6 scFv/G1/zeta chimeric receptor (Example 3) were incubated for 16h with an equal number of HL60 cells in the presence of various mouse monoclonal antibodies directed against human T-cell surface markers. The bivalent antibodies were included at 10 $\mu\text{g/ml}$ to test for their ability to co-stimulate the zeta - chain chimera. The antibodies used in this experiment were: anti-CD2
10 RPA2.10 (Pharmingen), anti-CD3 OKT3 (ATCC), anti-CD4 OKT4 (ATCC), anti-CD5 UCHT2 (Pharmingen), anti-CD28 15E8 (Caltag) and a control antibody MOPC21 (ATCC). IL-2 accumulated in the supernatant at the end of the incubation was measured by Quantikine IL-2 ELISA (R&D Systems).

15 The results indicate that anti-CD2, anti-CD5 and anti-CD28 co-stimulate production of IL-2 in HGT16.1 cells in response to HL60 target cells hence confirming CD2, CD5 and CD28 as co-stimulatory receptors compatible with zeta-chain chimera signalling. From experiments designed in this
20 way, it would be possible to determine the co-stimulatory activity of other cell surface molecules. The intracellular domains can then be included in chimeric receptors as described in Example 1 and evaluated as described in Examples 2, 3 and 4.

25 EXAMPLE 6**Introduction of chimeric receptors into primary human CTLs.**

In order to establish an assay for co-stimulation of cytolytic T-cell function, a zeta-chain chimera was introduced into primary human T-cells using recombinant adenovirus vectors. Peripheral blood mononuclear cells
30 (PBMC) were isolated from healthy volunteers using centrifugation over Ficoll-Hypaque (Pharmacia) according to the manufacturer's instructions and cultured in RPMI-1640 medium with 10% FCS in 175-cm² tissue culture flasks. Non-adherent cells were transferred to fresh tissue culture flasks after 24h and phytohaemagglutinin (PHA) was added to a final
35 concentration of 2 $\mu\text{g/ml}$ and human recombinant IL-2 at 50ng/ml. After 6 days, CD4 - positive cells were removed using anti-CD4 antibody

immobilised on magnetic Dynabeads (Becton - Dickinson) to leave a population of cells at least 95% CD8 - single positive (CTL cells). The cells were washed by centrifugation and resuspended in fresh medium +10% FCS at 10^6 cells /ml.

5

Recombinant adenovirus RAd160 (expressing the hP67.6/G1/zeta chimeric receptor, Example 4) or the control virus RAd35 was added to the cells at a multiplicity of infection (MOI) of up to 400 pfu/cell with 2 μ g/ml DEAE-Dextran and incubated for 24h. Samples of cells were then fixed in
10 1% glutaraldehyde in PBS and infection rates measured by staining RAd35 - infected cells for β -galactosidase activity using 5-Bromo-4-chloro-3-indolyl β -D-galactoside (X-gal; Promega, according to the manufacturer's instructions). By this method, infection frequencies were determined to be at least 80%. Infected cells were expanded in culture for
15 a further 6 days in medium containing 50 ng/ml human IL-2. In some experiments, 2mM sodium butyrate was added to infected CTL cells to induce expression from the hCMV-MIE promoter.

Cytolytic activity against the CD33-expressing tumour cell line HL60 was
20 detected in recombinant adenovirus - infected CD8-positive cells incubated for 6 days in IL-2 and 2mM butyrate using standard 6h ^{51}Cr release assays. 2×10^7 HL60 target cells were labelled by incubation with 25MBq ^{51}Cr (CJS4 Amersham) for 45 min. at 37°C in T-cell growth medium. After washing, 1.5×10^4 labelled HL60 cells were transferred
25 into each well of a 96-well microtitre plate in the presence of RAd - infected CD8-positive effector cells at ratios in the range 100 to 0.1 effector:target cells. Cells were incubated for 6h in T-cell growth medium before centrifuging the plates and removal of the supernatant for counting. Cytolysis was expressed as the amount of ^{51}Cr released into the medium
30 compared to that released by detergent treatment of target cells. In the experiment illustrated (Figure 20) specific lysis was mediated by RAd 160 - infected effector cells but not by CD8-positive cells infected with RAd35. The degree of specific lysis is increased with increased E:T ratio.

35 This assay is useful for determining the effects of co-stimulation on cytolytic function using anti-receptor antibodies, co-stimulatory cytokines

or co-stimulatory chimeric receptors. Cells starved of IL-2 for various lengths of time can also be used to increase the sensitivity of assays designed to evaluate co-stimulatory activities. CD28 chimeric receptors can be introduced by co-infection of recombinant adenovirus with RAd160.

- 5 Alternatively a fusion receptor containing both zeta and CD28 signalling domains can be introduced using a single recombinant adenovirus. Anti-receptor antibodies which may be screened in this assay include anti-CD2 and anti-CD5 (see Example 5).

10 **EXAMPLE 7**

Analysis of co-stimulatory activities in Macrophages and Monocytes.

- Human monocytes were isolated from peripheral blood as follows. PBMC were isolated as described above and adherent cells obtained by settling on to plastic tissue culture flasks for 24 h before washing extensively with
15 fresh medium.

- Primary macrophages were isolated from the peritoneal cavity of Wistar rats 5 days after i.p. injection of 5 ml 3% thioglycollate (Sigma T-9032) in saline according to the method of Argys (Argys 1967, J.Immunol. 99:744-
20 750) or 3 ml mineral oil (heavy white oil; Sigma 400-5). Peritoneal lavage was carried out with 20ml RPMI 1640 medium + 10% FCS and 3.15% sodium citrate. Greater than 60% of the cells in the peritoneal lavage were mononuclear phagocytes as defined by flow cytometry using FITC-conjugated mouse anti-rat macrophage antibody ED2 (Serotec) and
25 morphological characteristics. Adherent cells were enriched by applying cells to plastic flasks or 6-well plates in RPMI 1640 medium + 10% FCS and culturing for 2 days. Non-adherent cells were then removed by extensive washing with fresh medium. Alternatively, macrophages were purified by Percoll density centrifugation (Lawson and Stevenson 1983 Br.
30 J. Cancer 48: 227-237.)

- Monocytes and macrophages were maintained in culture for 48h and infected with recombinant adenoviruses at a MOI of up to 200 pfu/cell for 16h in the presence of 2 µg/ml DEAE-Dextran, after which the virus was
35 removed by washing with fresh medium. Up to 80% of human peripheral - blood monocytes and rat peritoneal macrophages were infectable using

this procedure, as determined using X-gal staining of cells infected with RAd35. The use of higher concentrations of virus increased the percentage of cells infected but led to a significant reduction in cell viability.

5

The recombinant adenovirus RAd160 can be used to provide a human CD33-specific primary stimulus to cells of the rat or mouse monocyte - macrophage lineage. Since human monocytes express the CD33 antigen, for the analysis of chimeric receptor function in human monocytic phagocytes, it may be more appropriate to use an alternative binding specificity such as the hCTMO1scFv - containing chimeric receptor, constructed as in Example 1 and inserted into a recombinant adenovirus vector. Additionally, the zeta chain sequences of the chimeric receptor may be substituted with the transmembrane and intracellular domain of a FcRIII γ chain (Park et al 1993, J. Clin. Invest. 92: 2073-2079).

10
15

Rat peritoneal macrophages infected with RAd160 at an MOI of 100 pfu/cell, expressed high levels of chimeric receptor on their surfaces 48h post-infection as determined by staining with FITC-CD33 and analysis by a FACScan flow cytometer.

20

The response of monocytes and macrophages expressing the appropriate chimeric receptor to stimulation with specific antigen or antigen-expressing cells recognised by the scFv is measured in standard ^{51}Cr release assays (Example 6). Alternatively, phagocytosis and cytostasis assays (Lawson and Stevenson 1983 Br. J. Cancer 48: 227-237) or assays for the release of cytokines are carried out eg human TNF ELISA (R&D Systems) or rat TNF ELISA (Biosource).

25

Identification of appropriate receptor intracellular domains to provide a co-stimulatory signal can be accomplished by incubation of macrophages expressing the chimeric receptor with a source of the specific antigen and with cross-linking antibodies or natural ligands specific for individual cell surface receptors present on monocytes and macrophages as described in Example 5. Suitable receptors include the IL-2 receptor, the CSF-1 receptor, the IFN- γ receptor, the GM-CSF receptor and TNF receptors.

30
35

- Natural ligands which can be used for human monocytes / macrophages include recombinant human IL-2, human CSF-1 (M-CSF), human IFN γ , human GM-CSF and human TNF α (all from Genzyme). Ligands which can be used for rat or mouse macrophages include recombinant rat or human
- 5 IL-2, human CSF-1 (M-CSF), mouse IFN γ , mouse GM-CSF and mouse TNF α (Genzyme). Species-specific antibodies which cross-link and stimulate the chosen receptors can be raised using standard techniques or can be identified by screening commercially available antibodies.
- 10 Those antibodies or natural ligands which co-stimulate macrophage responses to CD33 identify candidate receptors whose intracellular domains or associated signalling molecules, such as receptor - associated tyrosine kinases, can be used to produce chimeric co-stimulatory receptors or fusion receptors containing both co-stimulatory and primary
- 15 signalling domains as described in Example 1. The intracellular components which may be used in these chimeric receptors include the following. The intracellular domains of the GM-CSF receptor β chain can be used as part of a di-sulphide linked homodimeric receptor or in combination with an intracellular component from the α chain (Muto et al.
- 20 1996, J. Exp. Med. 183: 1911-1916). The intracellular domains of the IFN γ -receptor α and β chains can be used (Bach et al., 1996.. Mol. Cell. Biol. 16: 3214-3221.), as can the intracellular domains of the IL-2 receptor, particularly the β and γ chains. One or more intracellular tyrosine kinase components can be used such as the jak1, jak2 and jak3 kinases or the
- 25 intracellular domain of the CSF-1 receptor tyrosine kinase (Carlberg and Rohrschneider 1994 Mol. Biol. Cell 5:81-95). If these tyrosine kinases are used, the receptors containing them are preferably constructed so that they are presented on the cell surface as monomers which oligomerise on binding of the scFv component to the target antigen, for instance using a
- 30 scFv coupled to a CD8 hinge extracellular component, coupled to a CD28 transmembrane component (see Example 1) which is coupled to the tyrosine kinase component.

EXAMPLE 8

- 35 Analysis of co-stimulatory activities in other cells of the immune system

- Additional immune cell types such as CD4-positive T-cells, B-cells, NK cells, basophils, neutrophils, haematopoietic stem cells are isolated from human peripheral blood, mouse or rat blood or peritoneal cavity or other sources by published procedures (Current Protocols in Immunology ed Coligan et al. John Wiley and Sons). Established cell lines which retain the differentiated functions of various immune cell types can also be used eg the human NK-like cell line YT2C2 (Roger et al 1996 Cellular Immunol. 168: 24-32.) A chimeric receptor capable of delivering a primary stimulus such as the hP67.6/G1/zeta chimera described above is introduced into the isolated immune cell type, eg by infection with recombinant adenovirus RAd160, and cross-linking antibodies or natural ligands of cell surface receptors are used to identify cell-surface molecules capable of providing co-stimulatory signals as described in Example 7.
- 15 Chimeric receptors containing appropriate cytoplasmic components to provide suitable co-stimulatory functions are then constructed as described in Example 1. The function of the chimeric receptors in the chosen cell types can be analysed using recombinant adenovirus vectors.

TABLE 1

POSSIBLE CHIMERIC RECEPTOR COMBINATIONS

LIGAND BINDING	SPACER	TRANS MEMBRANE	SPACER	CYTOSOLIC COMPONENT	SPACER	CYTOSOLIC COMPONENT	SPACER	CYTOSOLIC COMPONENT	* CYTOSOL SPACERS
A	TAA SCFV	G1	TCR ZETA	OPT**	OPT	TCR ZETA	OPT	OPT	OPT
	TAA SCFV	h	CD28	OPT	OPT	CD28	OPT	OPT	OPT
B	TAA SCFV	CD8	TCR ZETA	OPT	OPT	TCR ZETA	OPT	OPT	OPT
	TAA SCFV	h	CD28	OPT	OPT	CD28	OPT	OPT	OPT
C	TAA SCFV	G1	TCR ZETA	OPT	OPT	TCR ZETA	OPT	OPT	OPT
	TAA SCFV	G1	IL2 R β	OPT	OPT	IL2 R β	OPT	IL2 R γ	OPT
D	TAA SCFV	G1	TCR ZETA	OPT	OPT	TCR ZETA	OPT	CD28	OPT
E	TAA SCFV	h	TCR ZETA	OPT	OPT	TCR ZETA	OPT	CD28	OPT
F	TAA SCFV	G1	TCR ZETA	OPT	OPT	TCR ZETA	OPT	IL2 R β	IL2 R γ

A,B and C describe pairs of genes coding for pairs of chimeric receptors
D,E and F describe fusion chimeric receptors, as shown in C one of a pair of receptors may be a fusion receptor

TAA SCFV denotes a single chain FV to a Tumour associated antigen

For a pair of chimeric receptors the SCFVs may bind the same or different epitopes of the same antigen or different antigens on the same or different cells.

G1 is the IgG CH₃ CH₂ HINGE spacer construct described in the text

h denotes the IgG hinge plus part of the CD28 extracellular component described in the text

* one or more further cytosolic and or spacer components

** OPT = optional

TABLE 2

5

**CHIMERIC RECEPTOR CONSTRUCTS AND CELL LINE
NOMENCLATURE**

10	CONSTRUCT	CONSTRUCTION PLASMID	EXPRESSION PLASMID	CELL LINES
15	hCTMO1 scFv / CD8 / TCR zeta	pBS3	pTB3	TB3
	hP67.6 scFv / CD8 / TCR zeta	pBS5	pTB5	TB5
	hCTMO1 scFv / CD8 / CD28	pHMF 320	pHMF 332	HGT 1
20	hCTMO1 scFv / CD8 / TCR zeta-CD28	pHMF 326	pHMF 334	HGT 2
	hP67.6 scFv / G1 / TCR zeta	pHMF 342	pHMF 351	HGT 6 & 16
25	hP67.6 scFv / G1 / TCR zeta-CD28	pHMF 354	pHMF 355	HGT 7 & 17
	hP67.6 scFv / h / CD28	pHMF 350	pHMF 353	HGT 8 & 14
	hP67.6 scFv / G1 / CD28	pHMF 375	pHMF 376	HGT 23
30	hP67.6 scFv / G1 / CD28-TCR zeta	pHMF 372	pHMF 373	HGT 21
	hP67.6 scFv / G1 / CD28-TCR zeta-CD28	pHMF 379	pHMF 380	HGT 26
35	hP67.6 scFv / h.28 / TCR zeta	pHMF 377	pHMF 378	HGT 24
	hP67.6 scFv / h.28 / TCR zeta - CD28	pHMF 363	pHMF 364	HGT 20
	hP67.6 scFv / h.28 / CD28 - TCR zeta	pHMF 369	pHMF 371	HGT 22

40

G1 is the IgG hinge CH2 CH3 spacer

45

h is the IgG hinge component plus part of CD28 extracellular domain spacer.

h.28 is the IgG hinge component plus part of CD28 extracellular domain and amino acid residues L, D, P & K spacer.

50

Expression plasmids pTB3 and pTB5, pHMF 334, 351, 355, 378 and 364 include the TCR zeta transmembrane domain.

Expression plasmids pHMF 332, 353, 376, 373, 380 and 371 include the CD28 transmembrane domain.

55

CLAIMS

1. A method of activating a cell as a result of one type of extracellular interaction between said first cell and a molecule associated with a second target cell characterised in that said first cell is provided with a DNA delivery system comprising DNA coding for one or more recombinant chimeric receptors comprising two or more different cytoplasmic signalling components, wherein said cytoplasmic components are not naturally linked, and at least one is derived from a membrane spanning polypeptide.
2. A method according to Claim 1 wherein the cytoplasmic signalling components are capable of acting together cooperatively.
3. A method according to Claim 1 or Claim 2 wherein said DNA additionally codes for signal peptide, binding and/or transmembrane components of said one or more chimeric receptors, wherein the binding component is capable of recognising a cell surface molecule on a target cell.
4. A method according to Claim 3 wherein the signal peptide, transmembrane and cytoplasmic signalling components and all or part of the binding component are coded for by a single DNA coding sequence.
5. A method according to Claim 3 wherein each cytoplasmic signalling component is coded for by a separate DNA coding sequence, each of DNA sequence additionally coding for a signal peptide, a transmembrane component and all or part of a binding component.
6. A method according to Claim 4 or Claim 5 wherein said DNA codes for part of said binding component and an additional separate DNA coding sequence codes for the remainder of the binding component.
7. A method according to Claim 5 or Claim 6 wherein the binding component coded for by one DNA sequence is capable of

participating in the same type of extracellular binding event as the binding component coded for by any other DNA sequence.

- 5 8. A method according to Claim 7 wherein each binding component binds to the same molecule associated with the target cell.
9. A method according to Claim 8 wherein each binding component is the same.
- 10 10. A method according to any one of Claims 1 to 9 wherein the one or more recombinant chimeric receptors are capable of recognising a viral or cell surface molecule on a target cell.
- 15 11. A DNA delivery system comprising DNA in association with a carrier said DNA coding for a recombinant chimeric receptor capable of one type of extracellular interaction and comprising two or more different cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide.
- 20 12. A DNA delivery system comprising DNA in association with a carrier said DNA coding for two or more recombinant chimeric receptors each capable of the same one type of extracellular interaction and wherein each of said receptors comprises one or more different cytoplasmic signalling components which are not naturally linked, and
- 25 wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide.
- 30 13. A DNA delivery system according to Claim 11 wherein said DNA codes in reading frame for:
 - i) a signal peptide component;
 - ii) a binding component capable of recognising a cell surface molecule on a target cell;
 - iii) a transmembrane component;
 - 35 iv) two or more different cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic

components is derived from a membrane spanning polypeptide; and optionally

v) one or more spacer regions linking any two or more of said i) to iv) components.

5

14. A DNA delivery system according to Claim 11 wherein said DNA comprises 1) a first DNA which codes in reading frame for:

i) a signal peptide component;

ii) part of a binding component;

10 iii) a transmembrane component;

iv) two or more cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide; and optionally

15 v) one or more spacer regions linking any two or more of said i) to iv) components; and 2) a second separate DNA which codes in reading frame for a signal peptide component and a further part of the binding component ii) coded for by said first DNA, such that the binding component parts together are capable of recognising a cell
20 surface molecule on a target cell.

15. A DNA delivery system according to Claim 12 wherein said DNA comprises a first and a second separate DNA each of which codes in reading frame for:

25 i) a signal peptide component;

ii) a binding component capable of recognising a cell surface molecule on a target cell;

iii) a transmembrane component;

30 iv) one or more different cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide; and optionally

v) one or more spacer regions linking any two or more of said i) to iv) components; provided that said first DNA codes for at least one
35 signalling component iv) that is not coded for by said second DNA.

16. A DNA delivery system according to Claim 12 wherein said DNA comprises 1) a first and a second separate DNA each of which codes in reading frame for:
- i) a signal peptide component;
 - 5 ii) one part of a binding component;
 - iii) a transmembrane component;
 - iv) one or more different cytoplasmic signalling components which are not naturally linked, and wherein at least one of said cytoplasmic components is derived from a membrane spanning polypeptide; and
 - 10 optionally
 - v) one or more spacer regions linking any two or more of said i) to iv) components; provided that said first DNA codes for at least one signalling component iv) that is not coded for by said second DNA; and 2) a separate third and fourth DNA each of which codes in
 - 15 reading frame for a signal peptide component and a further part of the binding component ii) coded for by said first and second DNA respectively, such that the binding component parts together provided by the first and third DNA and together provided by the second and fourth DNA are each capable of recognising a cell
 - 20 surface molecule on a target cell.
17. A DNA delivery system according to Claims 13 to 16 wherein each signal peptide component is an immunoglobulin signal sequence.
- 25 18. A DNA delivery system according to Claims 15 to 17 wherein the binding component coded for by said first DNA is the same as the binding component coded for by said second DNA.
- 30 19. A DNA delivery system according to Claims 13 to 18 wherein the binding component is an antibody or an antigen binding fragment thereof.
- 35 20. A DNA delivery system according to Claim 19 wherein the antibody or fragment thereof is an engineered human antibody or antigen binding fragment thereof.

21. A DNA delivery system according to Claims 18 to 20 wherein the binding component is a single chain Fv fragment.
22. A DNA delivery system according to Claims 18 to 20 wherein the binding component is a Fab' fragment.
23. A DNA delivery system according to any one of Claims 13 to 22 wherein the transmembrane component is derived from all or part of the alpha, beta or zeta chain of the T-cell receptor, CD28, CD8, CD4, a cytokine receptor or a colony stimulating factor receptor.
24. A DNA delivery system according to Claim 23 wherein the transmembrane component is derived from all or part of CD28.
25. A DNA delivery system according to any one of Claims 11 to 24 wherein the cytoplasmic signalling components are capable of acting together cooperatively.
26. A DNA delivery system according to any one of Claims 13 to 25 wherein the cytoplasmic signalling components are derived from all or part of the cytoplasmic domains of a zeta, eta or epsilon chain of the T-cell receptor, CD28, the γ chain of a Fc receptor, a cytokine receptor, a colony stimulating factor receptor, a tyrosine kinase or an adhesion molecule, B29, MB-1, CD3 delta, CD3 gamma, CD5 or CD2.
27. A DNA delivery system according to Claim 26 wherein the cytoplasmic signalling components are ITAM containing cytoplasmic components.
28. A DNA delivery system according to Claim 26 or Claim 27 wherein the cytoplasmic signalling components are derived from all or part of CD28 and/or the zeta chain of the T-cell receptor.

29. A DNA delivery system according to any one of Claims 11 to 28 wherein the cytoplasmic signalling components are in any orientation relative to one another.
- 5 30. A DNA delivery system according to any one of Claims 13 to 29 wherein said DNA coding for components i) to iv) additionally codes for one or more spacer regions linking the binding component ii) and the transmembrane component iii).
- 10 31. A DNA delivery system according to Claim 30 wherein two or more different spacer regions link the binding component ii) and the transmembrane component iii), both regions either being coded for by one DNA sequence or when a first and second DNA sequence is present one region being coded for by said first DNA and the other
- 15 different region being coded for by said second DNA.
32. A DNA delivery system according to Claims 30 or Claim 31 wherein the spacer region is selected to provide one or more free thiol groups.
- 20 33. A DNA delivery system according to Claims 30 to 32 wherein the spacer region is derived from all or part of the extracellular region of CD8, CD4 or CD28 .
34. A DNA delivery system according to Claims 30 or Claim 32 wherein
- 25 the spacer region is all or part of an antibody constant region.
35. A DNA delivery system according to Claims 30 to 32 wherein the spacer region is derived from all or part of an antibody hinge region linked to all or part of the extracellular region of CD28.
- 30 36. A DNA delivery system according to any one of Claims 11 to 35 wherein the carrier is a viral vector or a non-viral vector.
37. A DNA delivery system according to Claim 36 wherein the non-viral
- 35 vector is a liposomal vector.

38. A DNA delivery system according to Claim 37 wherein the carrier is a targeted non-viral vector.
- 5 39. A DNA delivery system according to Claim 38 wherein the targeted vector is an antibody targeted liposome.
40. A DNA delivery system according to Claim 38 wherein the targeted vector is an antibody targeted condensed DNA.
- 10 41. A DNA delivery system according to Claim 40 wherein the targeted vector is an antibody targeted protamine or polylysine condensed DNA.
- 15 42. A DNA delivery system according to Claim 38 wherein the targeted vector is antibody targeted naked DNA.
43. A DNA delivery system according to Claims 39 to 42 wherein the antibody is a whole antibody or an antigen binding fragment thereof.
- 20 44. A DNA delivery system according to Claim 43 wherein the antibody is an engineered human antibody or an antigen binding fragment thereof.
- 25 45. An effector cell transfected with a DNA delivery system according to any one of Claims 1 to 44.
46. An effector cell according to Claim 45 which is a lymphocyte, a dendritic cell, a B-cell, a haematopoietic stem cell, a macrophage, a monocyte or a NK cell.
- 30 47. An effector cell according to Claim 46 which is a cytotoxic T-lymphocyte.
- 35 48. A DNA delivery system according to any one of Claims 11 to 47 for use in the treatment of infectious disease, inflammatory disease,

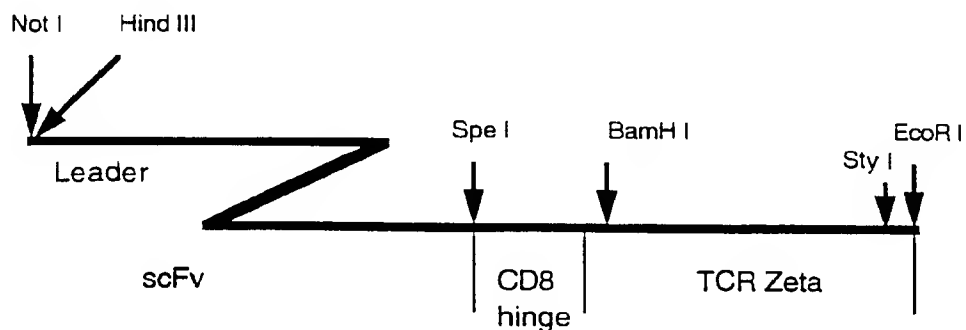
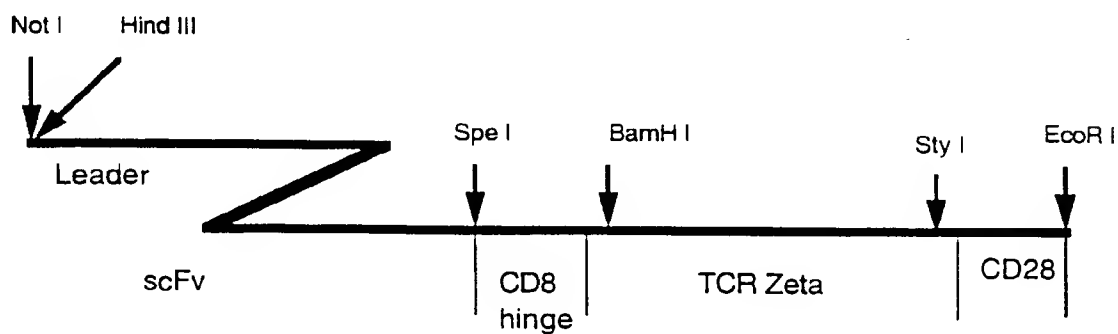
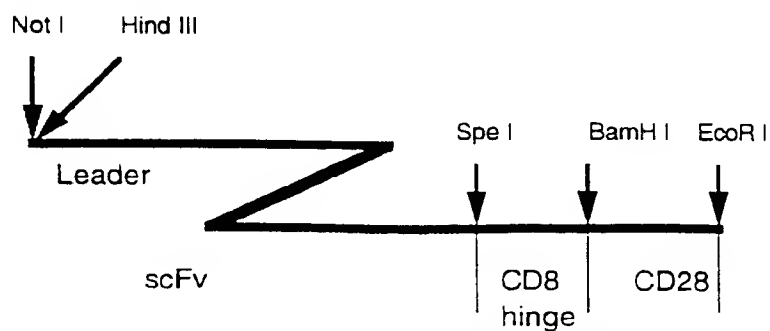
cancer, allergic/atopic disease, congenital disease, dermatologic disease, neurologic disease, transplants and metabolic/idiopathic disease.

- 5 49. A DNA delivery system according to Claim 48 for use in the treatment of rheumatoid arthritis, osteoarthritis, inflammatory bowel disease, asthma, eczema, cystic fibrosis, sickle cell anaemia, psoriasis, multiple sclerosis, organ or tissue transplant rejection, graft-versus-host disease or diabetes.
- 10 50. A pharmaceutical composition comprising a DNA delivery system according to any one of Claims 11 to 44 together with one or more formulatory agents.
- 15 51. A pharmaceutical composition according to Claim 50 wherein the formulatory agent is a suspending, preservative, stabilising and/or dispersing agent.
- 20 52. DNA coding for a recombinant chimeric receptor for use in a delivery system according to any one of Claims 11 to 44.

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FIG. 1

Construct cassettes cloned into pBluescript SK +

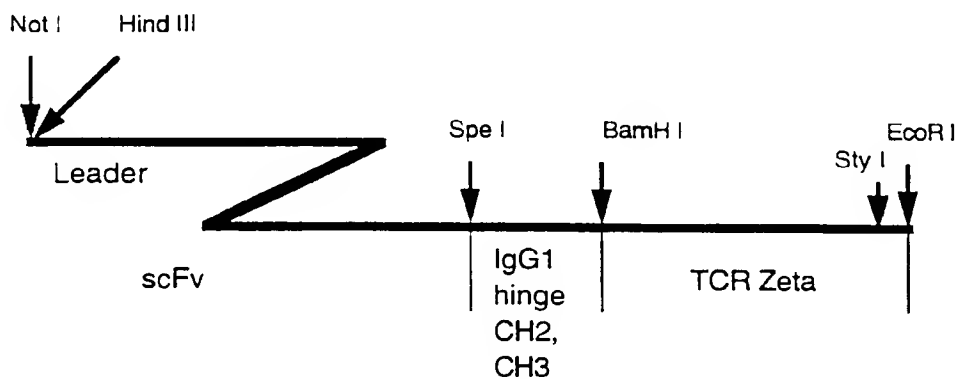
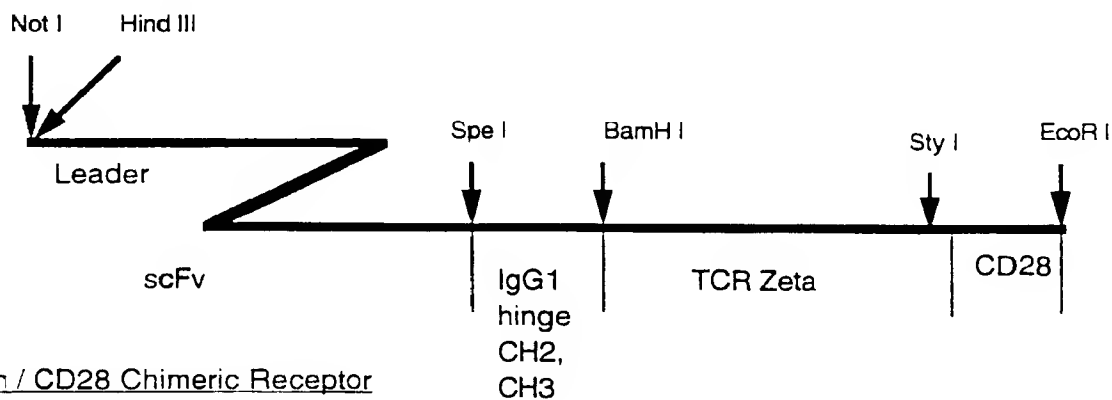
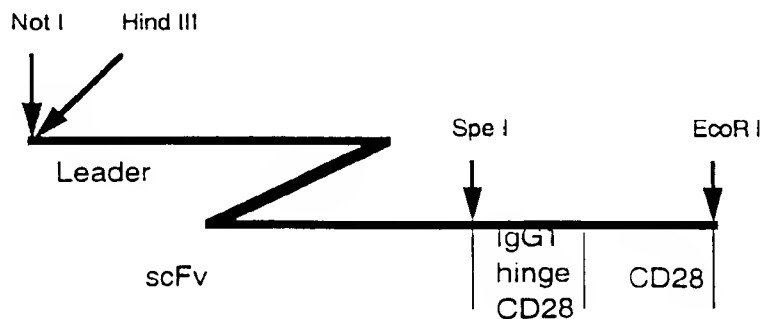
scFv / CD8 / Zeta Chimeric ReceptorscFv / CD8 / Zeta-CD28 fusion Chimeric ReceptorscFv / CD8 / CD28 Chimeric Receptor

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FIG. 2a

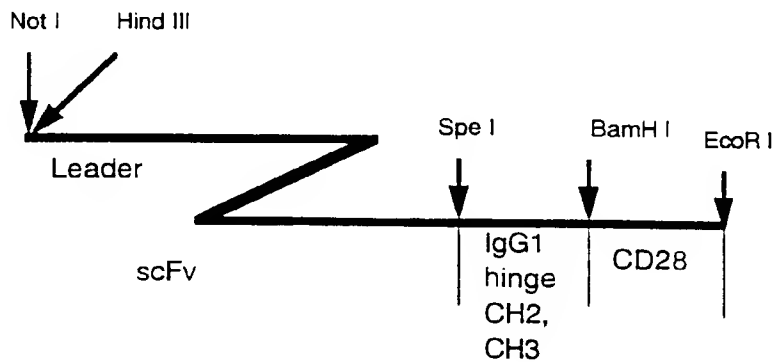
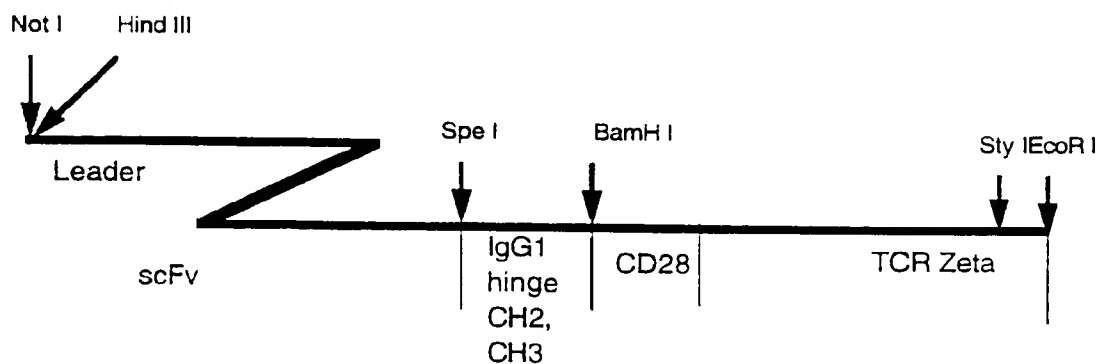
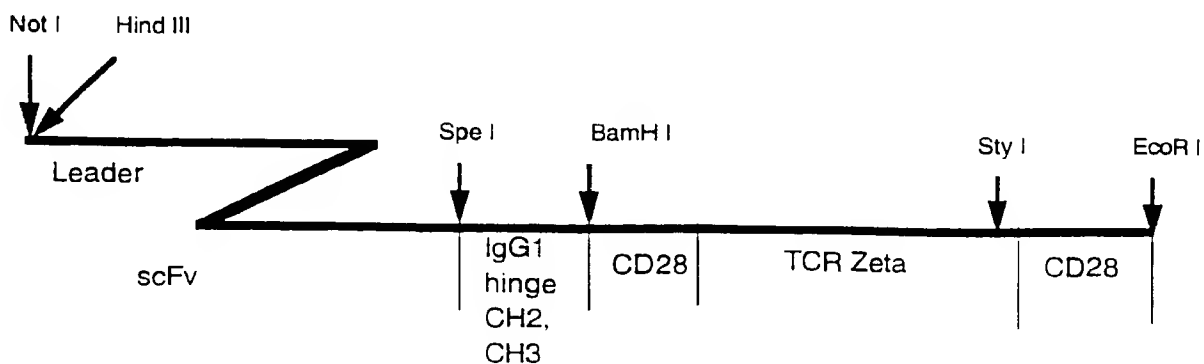
Construct cassettes cloned into pBluescript SK +

scFv / G1 / Zeta Chimeric ReceptorscFv / G1 / Zeta-CD28 fusion Chimeric ReceptorscFv / h / CD28 Chimeric Receptor

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FIG. 2b

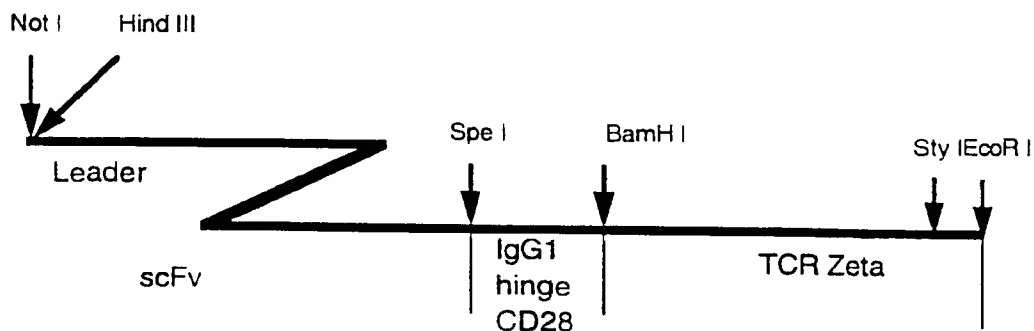
Construct cassettes cloned into pBluescript SK +

scFv /G1 /CD28 Chimeric ReceptorscFv /G1 /CD28-Zeta fusion Chimeric ReceptorscFv /G1 /CD28-Zeta-CD28 fusion Chimeric Receptor

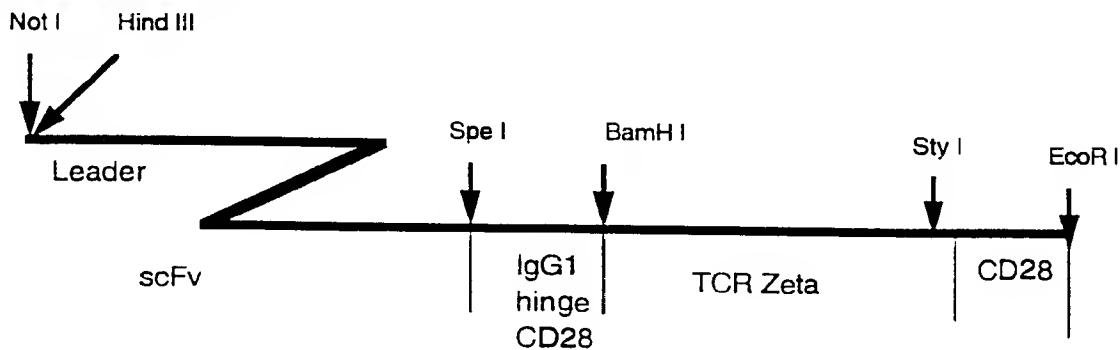
4 / 40

FIG. 2c
Construct cassettes cloned into pBluescript SK +

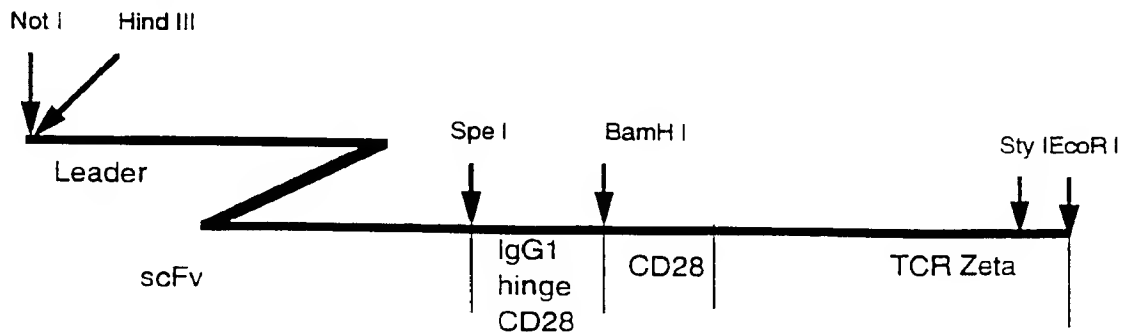
scFv / h.28 / Zeta Chimeric Receptor



scFv / h.28 / Zeta - CD28 fusion Chimeric Receptor



scFv / h.28 / CD28-Zeta fusion Chimeric Receptor



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FIG. 3
OLIGONUCLEOTIDE SEQUENCES FOR T-BODY CONSTRUCTION

All oligos listed in the 5' to 3' orientation.

R6490 : ATA TAG CGG CCG CAA GCT TCC ACC ATG TCT GTC CCC ACC CAA
GTC CTC

R6491 : TGA CCC TCC GCC ACC TGA CCC TCC GCC ACC TGA CCC TCC GCC
ACC TGA CCC TCC GCC ACC TGA CCC TCC GCC ACC TTT TAC TTC TAC TTT AGT ACC

R6492 : GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA
GGG TCA GGT GGC GGA GGG TCA GAG GTG CAG CTG GTG CAG TCT

R6493 : TAT ATA CTA GTA GAA GAC ACT GTC ACC AGT

R6516 : TGA CCC TCC GCC ACC TGA CCC TCC GCC ACC TGA CCC TCC GCC
ACC TGA CCC TCC GCC ACC CGT ACG TTT TAC TTC TAC TTT

R6515 : GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA
GGG TCA GGT GGC GGA GGG TCA CAG ATT CAG CTG GTG CAG TCT

R6514 : TAT ATA CTA GTC GGG CCC TTC GTT GAG GCA

R6494 : ATA TAA CTA GTA ACT CCA TCA TGT ACT TCA GCC ACT TCG TGC
CGG TCT TCC TGC CAG CG

R6495 : CGG TGT TGG TGG TCG CGG CGC TGG CGT CGT GGT GGG CTT CGC
TGG CAG GAA GAC CGG CAC

R6496 : GCG CCG CGA CCA CCA ACA CCG GCG CCC ACC ATC GCG TCG CAG
CCC CTG TCC CTG CGC CCA

R6497 : TAT ATG GAT CCA GCA GGC CAA AGC TCT GCG CCT CTG GGC GCA
GGG ACA GGG GCT G

R6506 : TAT ATG GAT CCC GCC TCT GGG CGC AGG GAC AGG GGC TG

R6488 : ATA TAG GAT CCC AAA CTC TGC TAC CTG CTG

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FIG. 3 (contd.)

R6489 : TAT ATG AAT TCT TAG CGA GGG GGC AGG GCC TGC AT

P3240 : TAT GGA TCC AAG CCC TTT TGG GTG CTG GTG GTG

P3241 : TAT GAA TTC TCA GGA GCG ATA GGC TGC GAA

P3301 : GCC ACC AAG GAC ACC TAC GAC GC

P3302 : CCC CCT CGC AGG AGT AAG AGG AGC AGG CTC CTG CAC AGT GAC

TAC ATG AAC ATG ACT CCC C

P3303 : CAA GCA TTA CCA GCC CTA TGC CCC ACC ACG CGA CTT CGC AGC

CTA TCG CTC CTG AGA ATT CAT A

P3304 : TAT GAA TTC TCA GGA GCG ATA G

P3305 : GCA TAG GGCTGG TAA TGC TTG CGG GTG GGC CCG GGG CGG CGG

GGA GTC ATG TTC ATG TAG T

P3306 : CTC TTA CTC CTG CGA GGG GGC AGG GCC TGC ATG TGA AGG GCG

TCG TAG GTG TCC TTG GTG GC

S0146 : CGA CTA GTG ACA AAA CTC ACA CAT GCC CAC CGT GCC CAA AAG

GGA AAC ACC TTT GTC CAA GGT CCC

S0060 : CGA CTA GTG ACA AAA CTC ACA CAT GCC CAC CG

S0061 : TTG GGA TCC AGT TTA CCC GGA GAC AGG GAG AGG CT

T4057 : CTA CTA GTG ACA AAA CTC ACA C

T4058 : TTG GGA TCC AGG GGC TTA GAA GGT CCG GGA AAT AG

T7145 : CTG GAT CCC AAA TTT TGG GTG CTG GTG GTG GTT G

T4060 : GCT CCT GCT GAA CTT CAC TCT GGA GCG ATA GGC TGC GAA GTC G

T4387 : GCG ACT TCG CAG CCT ATC GCT CCA GAG TGA AGT TCA GCA GGA

GCG

S4700 : TAT GAA TTC TTA GCG AGG GGG CAG GGC CTG CAT G

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FIG. 4

SEQUENCE OF hCTMO1 / CD8 / ZETA RECOMBINANT CHIMERIC RECEPTOR

```

      10      20      30      40
      *      *      *      *
ATG TCT GTC CCC ACC CAA GTC CTC GGA CTC CTG CTG CTG TGG
TAC AGA CAG GGG TGG GTT CAG GAG CCT GAG GAC GAC GAC ACC
M   S   V   P   T   Q   V   L   G   L   L   L   L   W>

      50      60      70      80
      *      *      *      *
CTT ACA GAT GCC AGA TGC GAT ATC CAG ATG ACT CAG AGT CCA
GAA TGT CTA CGG TCT ACG CTA TAG GTC TAC TGA GTC TCA GGT
L   T   D   A   R   C   D   I   Q   M   T   Q   S   P>

      90      100     110     120
      *      *      *      *
AGT ACT CTC AGT GCC AGT GTA GGT GAT AGG GTC ACC ATC ACT
TCA TGA GAG TCA CGG TCA CAT CCA CTA TCC CAG TGG TAG TGA
S   T   L   S   A   S   V   G   D   R   V   T   I   T>

      130     140     150     160
      *      *      *      *
TGT AGG AGT AGT AAA AGT CTC CTC CAT AGT AAC GGT GAC ACC
ACA TCC TCA TCA TTT TCA GAG GAG GTA TCA TTG CCA CTG TGG
C   R   S   S   K   S   L   L   H   S   N   G   D   T>

      170     180     190     200     210
      *      *      *      *      *
TTC CTC TAT TGG TTC CAG CAG AAA CCA GGT AAA GCC CCA AAG
AAG GAG ATA ACC AAG GTC GTC TTT GGT CCA TTT CGG GGT TTC
F   L   Y   W   F   Q   Q   K   P   G   K   A   P   K>

      220     230     240     250
      *      *      *      *
CTC CTC ATG TAT AGG ATG AGT AAC CTC GCC AGT GGT GTA CCA
GAG GAG TAC ATA TCC TAC TCA TTG GAG CGG TCA CCA CAT GGT
L   L   M   Y   R   M   S   N   L   A   S   G   V   P>

      260     270     280     290
      *      *      *      *
TCT AGA TTC AGT GGT AGT GGT AGT GGT ACT GAG TTC ACT CTC
AGA TCT AAG TCA CCA TCA CCA TCA CCA TGA CTC AAG TGA GAG
S   R   F   S   G   S   G   S   G   T   E   F   T   L>

      300     310     320     330
      *      *      *      *
ACT ATC AGT AGT CTC CAG CCA GAT GAT TTC GCC ACT TAT TAT
TGA TAG TCA TCA GAG GTC GGT CTA CTA AAG CGG TGA ATA ATA
T   I   S   S   L   Q   P   D   D   F   A   T   Y   Y>

      340     350     360     370
      *      *      *      *
TGT ATG CAG CAT CTC GAA TAT CCA TTC ACT TTC GGT CAG GGT
ACA TAC GTC GTA GAG CTT ATA GGT AAG TGA AAG CCA GTC CCA
C   M   Q   H   L   E   Y   P   F   T   F   G   Q   G>

      380     390     400     410     420
      *      *      *      *      *
ACT AAA GTA GAA GTA AAA CGT ACG GGT GGC GGA GGG TCA GGT
TGA TTT CAT CTT CAT TTT GCA TGC CCA CCG CCT CCC AGT CCA
T   K   V   E   V   K   R   T   G   G   G   G   S   G>

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FIG. 4(contd.)

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      430      440      450      460
      *      *      *      *
GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA
CCG CCT CCC AGT CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT
G  G  G  S  G  G  G  G  S  G  G  G  G  S>

      470      480      490      500
      *      *      *      *
GGT GGC GGA GGG TCA CAG ATT CAG CTG GTG CAG TCT GGA GCA
CCA CCG CCT CCC AGT GTC TAA GTC GAC CAC GTC AGA CCT CGT
G  G  G  G  S  Q  I  Q  L  V  Q  S  G  A>

      510      520      530      540
      *      *      *      *
GAG GTG AAG AAG CCT GGA TCT TCT GTG AAG GTG TCT TGT AAG
CTC CAC TTC TTC GGA CCT AGA AGA CAC TTC CAC AGA ACA TTC
E  V  K  K  P  G  S  S  V  K  V  S  C  K>

      550      560      570      580
      *      *      *      *
GCA TCT GGA TAC ACC TTC ACC GAC TAC TAC ATT AAT TGG ATG
CGT AGA CCT ATG TGG AAG TGG CTG ATG ATG TAA TTA ACC TAC
A  S  G  Y  T  F  T  D  Y  Y  I  N  W  M>

      590      600      610      620      630
      *      *      *      *      *
AGA CAG GCA CCT GGA CAG GGA CTC GAG TGG ATT GGA TGG ATT
TCT GTC CGT GGA CCT GTC CCT GAG CTC ACC TAA CCT ACC TAA
R  Q  A  P  G  Q  G  L  E  W  I  G  W  I>

      640      650      660      670
      *      *      *      *
GAC CCT GGA TCT GGA AAT ACA AAG TAC AAT GAG AAG TTC AAG
CTG GGA CCT AGA CCT TTA TGT TTC ATG TTA CTC TTC AAG TTC
D  P  G  S  G  N  T  K  Y  N  E  K  F  K>

      680      690      700      710
      *      *      *      *
GGA AGA GCA ACA CTG ACA GTG GAC ACA TCC ACG AAT ACC GCC
CCT TCT CGT TGT GAC TGT CAC CTG TGT AGG TGC TTA TGG CCG
G  R  A  T  L  T  V  D  T  S  T  N  T  A>

      720      730      740      750
      *      *      *      *
TAC ATG GAG CTG TCT TCT CTG AGA TCT GAG GAC ACA GCA TTC
ATG TAC CTC GAC AGA AGA GAC TCT AGA CTC CTG TGT CGT AAG
Y  M  E  L  S  S  L  R  S  E  D  T  A  F>

      760      770      780      790
      *      *      *      *
TAC TTC TGT GCA AGA GAG AAG ACC ACC TAC TAC TAC GCA ATG
ATG AAG ACA CGT TCT CTC TTC TGG TGG ATG ATG ATG CGT TAC
Y  F  C  A  R  E  K  T  T  Y  Y  Y  A  M>

      800      810      820      830      840
      *      *      *      *      *
GAC TAC TGG GGA CAG GGA ACA CTG GTG ACA GTG TCT TCT GCC
CTG ATG ACC CCT GTC CCT TGT GAC CAC TGT CAC AGA AGA CGG
D  Y  W  G  Q  G  T  L  V  T  V  S  S  A>

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FIG. 4(contd.)

850				860				870				880			
*				*				*				*			
TCA	ACG	AAG	GGC	CCG	ACT	AGT	AAC	TCC	ATC	ATG	TAC	TTC	AGC		
AGT	TGC	TTC	CCG	GGC	TGA	TCA	TTG	AGG	TAG	TAC	ATG	AAG	TCG		
S	T	K	G	P	T	S	N	S	I	M	Y	F	S>		
890				900				910				920			
*				*				*				*			
CAC	TTC	GTG	CCG	GTC	TTC	CTG	CCA	GCG	AAG	CCC	ACC	ACG	ACG		
GTG	AAG	CAC	GGC	CAG	AAG	GAC	GGT	CGC	TTC	GGG	TGG	TGC	TGC		
H	F	V	P	V	F	L	P	A	K	P	T	T	T>		
930				940				950				960			
*				*				*				*			
CCA	GCG	CCG	CGA	CCA	CCA	ACA	CCG	GCG	CCC	ACC	ATC	GCG	TCG		
GGT	CGC	GGC	GCT	GGT	GGT	TGT	GGC	CGC	GGG	TGG	TAG	CGC	AGC		
P	A	P	R	P	P	T	P	A	P	T	I	A	S>		
970				980				990				1000			
*				*				*				*			
CAG	CCC	CTG	TCC	CTG	CGC	CCA	GAG	GCG	CAG	AGC	TTT	GGC	CTG		
GTC	GGG	GAC	AGG	GAC	GCG	GGT	CTC	CGC	GTC	TCG	AAA	CCG	GAC		
Q	P	L	S	L	R	P	E	A	Q	S	F	G	L>		
1010				1020				1030				1040			
*				*				*				*			
CTG	GAT	CCC	AAA	CTC	TGC	TAC	CTG	CTG	GAT	GGA	ATC	CTC	TTC		
GAC	CTA	GGG	TTT	GAG	ACG	ATG	GAC	GAC	CTA	CCT	TAG	GAG	AAG		
L	D	P	K	L	C	Y	L	L	D	G	I	L	F>		
1060				1070				1080				1090			
*				*				*				*			
ATC	TAT	GGT	GTC	ATT	CTC	ACT	GCC	TTG	TTC	CTG	AGA	GTG	AAG		
TAG	ATA	CCA	CAG	TAA	GAG	TGA	CGG	AAC	AAG	GAC	TCT	CAC	TTC		
I	Y	G	V	I	L	T	A	L	F	L	R	V	K>		
1100				1110				1120				1130			
*				*				*				*			
TTC	AGC	AGG	AGC	GCA	GAC	GCC	CCC	GCG	TAC	CAG	CAG	GGC	CAG		
AAG	TCG	TCC	TCG	CGT	CTG	CGG	GGG	CGC	ATG	GTC	GTC	CCG	GTC		
F	S	R	S	A	D	A	P	A	Y	Q	Q	G	Q>		
1140				1150				1160				1170			
*				*				*				*			
AAC	CAG	CTC	TAT	AAC	GAG	CTC	AAT	CTA	GGA	CGA	AGA	GAG	GAG		
TTG	GTC	GAG	ATA	TTG	CTC	GAG	TTA	GAT	CCT	GCT	TCT	CTC	CTC		
N	Q	L	Y	N	E	L	N	L	G	R	R	E	E>		
1180				1190				1200				1210			
*				*				*				*			
TAC	GAT	GTT	TTG	GAC	AAG	AGA	CGT	GCG	CGG	GAC	CCT	GAG	ATG		
ATG	CTA	CAA	AAC	CTG	TTC	TCT	GCA	CCG	GCC	CTG	GGA	CTC	TAC		
Y	D	V	L	D	K	R	R	G	R	D	P	E	M>		
1220				1230				1240				1250			
*				*				*				*			
GGG	GGA	AAG	CCG	AGA	AGG	AAG	AAC	CCT	CAG	GAA	GGC	CTG	TAC		
CCC	CCT	TTC	GGC	TCT	TCC	TTC	TTG	GGA	GTC	CTT	CCG	GAC	ATG		
G	G	K	P	R	R	K	N	P	Q	E	G	L	Y>		

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FIG. 4(contd)

1270	1280	1290	1300
*	*	*	*
AAT GAA CTG CAG AAA GAT AAG ATG GCG GAG GCC TAC AGT GAG			
TTA CTT GAC GTC TTT CTA TTC TAC CGC CTC CGG ATG TCA CTC			
N E L Q K D K M A E A Y S E>			
1310	1320	1330	1340
*	*	*	*
ATT GGG ATG AAA GGC GAG CGC CGG AGG GGC AAG GGG CAC GAT			
TAA CCC TAC TTT CCG CTC GCG GCC TCC CCG TTC CCC GTG CTA			
I G M K G E R R R G K G H D>			
1350	1360	1370	1380
*	*	*	*
GGC CTT TAC CAG GGT CTC AGT ACA GCC ACC AAG GAC ACC TAC			
CCG GAA ATG GTC CCA GAG TCA TGT CGG TGG TTC CTG TGG ATG			
G L Y Q G L S T A T K D T Y>			
1390	1400	1410	1420
*	*	*	*
GAC GCC CTT CAC ATG CAG GCC CTG CCC CCT CGC TAA			
CTG CGG GAA GTG TAC GTC CGG GAC GGG GGA GCG ATT			
D A L H M Q A L P P R *			

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FIG. 5

SEQUENCE OF hCTMO1 / CD8 / Zeta-CD28 FUSION RECOMBINANT
CHIMERIC RECEPTOR

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      10      20      30      40
      *      *      *      *
ATG TCT GTC CCC ACC CAA GTC CTC GGA CTC CTG CTG CTG TGG CTT ACA
TAC AGA CAG GGG TGG GTT CAG GAG CCT GAG GAC GAC GAC ACC GAA TGT
m s v p t q v l g l l l l w l t>

50      60      70      80      90
*      *      *      *      *
GAT GCC AGA TGC GAT ATC CAG ATG ACT CAG AGT CCA AGT ACT CTC AGT
CTA CGG TCT ACG CTA TAG GTC TAC TGA GTC TCA GGT TCA TGA GAG TCA
d a r c D I Q M T Q S P S T L S>

100      110      120      130      140
*      *      *      *      *
GCC AGT GTA GGT GAT AGG GTC ACC ATC ACT TGT AGG AGT AGT AAA AGT
CGG TCA CAT CCA CTA TCC CAG TGG TAG TGA ACA TCC TCA TCA TTT TCA
A S V G D R V T I T C R S S K S>

150      160      170      180      190
*      *      *      *      *
CTC CTC CAT AGT AAC GGT GAC ACC TTC CTC TAT TGG TTC CAG CAG AAA
GAG GAG GTA TCA TTG CCA CTG TGG AAG GAG ATA ACC AAG GTC GTC TTT
L L H S N G D T F L Y W F Q Q K>

200      210      220      230      240
*      *      *      *      *
CCA GGT AAA GCC CCA AAG CTC CTC ATG TAT AGG ATG AGT AAC CTC GCC
GGT CCA TTT CGG GGT TTC GAG GAG TAC ATA TCC TAC TCA TTG GAG CGG
P G K A P K L L M Y R M S N L A>

250      260      270      280
*      *      *      *
AGT GGT GTA CCA TCT AGA TTC AGT GGT AGT GGT AGT GGT ACT GAG TTC
TCA CCA CAT GGT AGA TCT AAG TCA CCA TCA CCA TCA CCA TGA CTC AAG
S G V P S R F S G S G S G T E F>

290      300      310      320      330
*      *      *      *      *
ACT CTC ACT ATC AGT AGT CTC CAG CCA GAT GAT TTC GCC ACT TAT TAT
TGA GAG TGA TAG TCA TCA GAG GTC GGT CTA CTA AAG CGG TGA ATA ATA
T L T I S S L Q P D D F A T Y Y>

340      350      360      370      380
*      *      *      *      *
TGT ATG CAG CAT CTC GAA TAT CCA TTC ACT TTC GGT CAG GGT ACT AAA
ACA TAC GTC GTA GAG CTT ATA GGT AAG TGA AAG CCA GTC CCA TGA TTT
C M Q H L E Y P F T F G Q G T K>

390      400      410      420      430
*      *      *      *      *
GTA GAA GTA AAA CGT ACG GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA
CAT CTT CAT TTT GCA TGC CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT
V E V K R T G G G G S G G G G S>

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FIG. 5(contd.)

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      440      450      460      470      480
      *      *      *      *      *
GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA CAG
CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT GTC
G  G  G  G  S  G  G  G  G  S  G  G  G  G  S  Q>

      490      500      510      520
      *      *      *      *
ATT CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG CCT GGA TCT TCT
TAA GTC GAC CAC GTC AGA CCT CGT CTC CAC TTC TTC GGA CCT AGA AGA
I  Q  L  V  Q  S  G  A  E  V  K  K  P  G  S  S>

530      540      550      560      570
*      *      *      *      *
GTG AAG GTG TCT TGT AAG GCA TCT GGA TAC ACC TTC ACC GAC TAC TAC
CAC TTC CAC AGA ACA TTC CGT AGA CCT ATG TGG AAG TGG CTG ATG ATG
V  K  V  S  C  K  A  S  G  Y  T  F  T  D  Y  Y>

      580      590      600      610      620
      *      *      *      *      *
ATT AAT TGG ATG AGA CAG GCA CCT GGA CAG GGA CTC GAG TGG ATT GGA
TAA TTA ACC TAC TCT GTC CGT GGA CCT GTC CCT GAG CTC ACC TAA CCT
I  N  W  M  R  Q  A  P  G  Q  G  L  E  W  I  G>

      630      640      650      660      670
      *      *      *      *      *
TGG ATT GAC CCT GGA TCT GGA AAT ACA AAG TAC AAT GAG AAG TTC AAG
ACC TAA CTG GGA CCT AGA CCT TTA TGT TTC ATG TTA CTC TTC AAG TTC
W  I  D  P  G  S  G  N  T  K  Y  N  E  K  F  K>

      680      690      700      710      720
      *      *      *      *      *
GGA AGA GCA ACA CTG ACA GTG GAC ACA TCC ACG AAT ACC GCC TAC ATG
CCT TCT CGT TGT GAC TGT CAC CTG TGT AGG TGC TTA TGG CGG ATG TAC
G  R  A  T  L  T  V  D  T  S  T  N  T  A  Y  M>

      730      740      750      760
      *      *      *      *
GAG CTG TCT TCT CTG AGA TCT GAG GAC ACA GCA TTC TAC TTC TGT GCA
CTC GAC AGA AGA GAC TCT AGA CTC CTG TGT CGT AAG ATG AAG ACA CGT
E  L  S  S  L  R  S  E  D  T  A  F  Y  F  C  A>

770      780      790      800      810
*      *      *      *      *
AGA GAG AAG ACC ACC TAC TAC TAC GCA ATG GAC TAC TGG GGA CAG GGA
TCT CTC TTC TGG TGG ATG ATG ATG CGT TAC CTG ATG ACC CCT GTC CCT
R  E  K  T  T  Y  Y  Y  A  M  D  Y  W  G  Q  G>

      820      830      840      850      860
      *      *      *      *      *
ACA CTG GTG ACA GTG TCT TCT GCC TCA ACG AAG GGC CCG ACT AGT AAC
TGT GAC CAC TGT CAC AGA AGA CGG AGT TGC TTC CCG GGC TGA TCA TTG
T  L  V  T  V  S  S  A  S  T  K  G  P  T  S  N>

      870      880      890      900      910
      *      *      *      *      *
TCC ATC ATG TAC TTC AGC CAC TTC GTG CCG GTC TTC CTG CCA GCG AAG
AGG TAG TAC ATG AAG TCG GTG AAG CAC GGC CAG AAG GAC GGT CGC TTC
S  I  M  Y  F  S  H  F  V  P  V  F  L  P  A  K>

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FIG. 5(contd.)

920	930	940	950	960
CCC ACC ACG ACG CCA GCG CCG CGA CCA CCA ACA CCG GCG CCC ACC ATC				
GGG TGG TGC TGC GGT CGC GGC GCT GGT GGT TGT GGC CGC GGG TGG TAG				
P T T T P A P R P P T P A P T I>				
970	980	990	1000	
GCG TCG CAG CCC CTG TCC CTG CGC CCA GAG GCG CAG AGC TTT GGC CTG				
CGC AGC GTC GGG GAC AGG GAC GCG GGT CTC CGC GTC TCG AAA CCG GAC				
A S Q P L S L R P E A Q S F G L>				
1010	1020	1030	1040	1050
CTG GAT CCC AAA CTC TGC TAC CTG CTG GAT GGA ATC CTC TTC ATC TAT				
GAC CTA GGG TTT GAG ACG ATG GAC GAC CTA CCT TAG GAG AAG TAG ATA				
L D P K L C Y L L D G I L F I Y>				
1060	1070	1080	1090	1100
GGT GTC ATT CTC ACT GCC TTG TTC CTG AGA GTG AAG TTC AGC AGG AGC				
CCA CAG TAA GAG TGA CGG AAC AAG GAC TCT CAC TTC AAG TCG TCC TCG				
G V I L T A L F L R V K F S R S>				
1110	1120	1130	1140	1150
GCA GAC GCC CCC GCG TAC CAG CAG GGC CAG AAC CAG CTC TAT AAC GAG				
CGT CTG CGG GGG CGC ATG GTC GTC CCG GTC TTG GTC GAG ATA TTG CTC				
A D A P A Y Q Q G Q N Q L Y N E>				
1160	1170	1180	1190	1200
CTC AAT CTA GGA CGA AGA GAG GAG TAC GAT GTT TTG GAC AAG AGA CGT				
GAG TTA GAT CCT GCT TCT CTC CTC ATG CTA CAA AAC CTG TTC TCT GCA				
L N L G R R E E Y D V L D K R R>				
1210	1220	1230	1240	
GGC CGG GAC CCT GAG ATG GGG GGA AAG CCG AGA AGG AAG AAC CCT CAG				
CCG GCC CTG GGA CTC TAC CCC CCT TTC GGC TCT TCC TTC TTG GGA GTC				
G R D P E M G G K P R R K N P Q>				
1250	1260	1270	1280	1290
GAA GGC CTG TAC AAT GAA CTG CAG AAA GAT AAG ATG GCG GAG GCC TAC				
CTT CCG GAC ATG TTA CTT GAC GTC TTT CTA TTC TAC CGC CTC CGG ATG				
E G L Y N E L Q K D K M A E A Y>				
1300	1310	1320	1330	1340
AGT GAG ATT GGG ATG AAA GGC GAG CGC CGG AGG GGC AAG GGG CAC GAT				
TCA CTC TAA CCC TAC TTT CCG CTC GCG GCC TCC CCG TTC CCC GTG CTA				
S E I G M K G E R R R G K G H D>				
1350	1360	1370	1380	1390
GGC CTT TAC CAG GGT CTC AGT ACA GCC ACC AAG GAC ACC TAC GAC GCC				
CCG GAA ATG GTC CCA GAG TCA TGT CCG TGG TTC CTG TGG ATG CTG CCG				
G L Y Q G L S T A T K D T Y D A>				

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FIG.5(contd.)

1400		1410		1420		1430		1440	
	*		*		*		*		*
CTT CAC ATG CAG GCC CTG CCC CCT CGC AGG AGT AAG AGG AGC AGG CTC									
GAA GTG TAC GTC CGG GAC GGG GGA GCG TCC TCA TTC TCC TCG TCC GAG									
L H M Q A L P P R R S K R S R L>									
		1450		1460		1470		1480	
		*		*		*		*	
CTG CAC AGT GAC TAC ATG AAC ATG ACT CCC CGC CGC CCC GGG CCC ACC									
GAC GTG TCA CTG ATG TAC TTG TAC TGA GGG GCG GCG GGG CCC GGG TGG									
L H S D Y M N M T P R R P G P T>									
1490		1500		1510		1520		1530	
*		*		*		*		*	
CGC AAG CAT TAC CAG CCC TAT GCC CCA CCA CGC GAC TTC GCA GCC TAT									
GCG TTC GTA ATG GTC GGG ATA CGG GGT GGT GCG CTG AAG CGT CGG ATA									
R K H Y Q P Y A P P R D F A A Y>									
		1540							
		*							
CGC TCC TGA									
GCG AGG ACT									
R S *									

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FIG. 6

SEQUENCE OF hCTMO1 /CD8 / CD28 RECOMBINANT CHIMERIC RECEPTOR

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      10      20      30      40
      *      *      *      *
ATG TCT GTC CCC ACC CAA GTC CTC GGA CTC CTG CTG CTG TGG
TAC AGA CAG GGG TGG GTT CAG GAG CCT GAG GAC GAC GAC ACC
M  S  V  P  T  Q  V  L  G  L  L  L  L  W>

      50      60      70      80
      *      *      *      *
CTT ACA GAT GCC AGA TGC GAT ATC CAG ATG ACT CAG AGT CCA
GAA TGT CTA CGG TCT ACG CTA TAG GTC TAC TGA GTC TCA GGT
L  T  D  A  R  C  D  I  Q  M  T  Q  S  P>

      90      100      110      120
      *      *      *      *
AGT ACT CTC AGT GCC AGT GTA GGT GAT AGG GTC ACC ATC ACT
TCA TGA GAG TCA CGG TCA CAT CCA CTA TCC CAG TGG TAG TGA
S  T  L  S  A  S  V  G  D  R  V  T  I  T>

      130      140      150      160
      *      *      *      *
TGT AGG AGT AGT AAA AGT CTC CTC CAT AGT AAC GGT GAC ACC
ACA TCC TCA TCA TTT TCA GAG GAG GTA TCA TTG CCA CTG TGG
C  R  S  S  K  S  L  L  H  S  N  G  D  T>

      170      180      190      200      210
      *      *      *      *      *
TTC CTC TAT TGG TTC CAG CAG AAA CCA GGT AAA GCC CCA AAG
AAG GAG ATA ACC AAG GTC GTC TTT GGT CCA TTT CGG GGT TTC
F  L  Y  W  F  Q  Q  K  P  G  K  A  P  K>

      220      230      240      250
      *      *      *      *
CTC CTC ATG TAT AGG ATG AGT AAC CTC GCC AGT GGT GTA CCA
GAG GAG TAC ATA TCC TAC TCA TTG GAG CGG TCA CCA CAT GGT
L  L  M  Y  R  M  S  N  L  A  S  G  V  P>

      260      270      280      290
      *      *      *      *
TCT AGA TTC AGT GGT AGT GGT AGT GGT ACT GAG TTC ACT CTC
AGA TCT AAG TCA CCA TCA CCA TCA CCA TGA CTC AAG TGA GAG
S  R  F  S  G  S  G  S  G  T  E  F  T  L>

      300      310      320      330
      *      *      *      *
ACT ATC AGT AGT CTC CAG CCA GAT GAT TTC GCC ACT TAT TAT
TGA TAG TCA TCA GAG GTC GGT CTA CTA AAG CGG TGA ATA ATA
T  I  S  S  L  Q  P  D  D  F  A  T  Y  Y>

      340      350      360      370
      *      *      *      *
TGT ATG CAG CAT CTC GAA TAT CCA TTC ACT TTC GGT CAG GGT
ACA TAC GTC GTA GAG CTT ATA GGT AAG TGA AAG CCA GTC CCA
C  M  Q  H  L  E  Y  P  F  T  F  G  Q  G>

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FIG. 6(contd.)

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380          390          400          410          420
*          *          *          *          *
ACT AAA GTA GAA GTA AAA CGT ACG GGT GGC GGA GGG TCA GGT
TGA TTT CAT CTT CAT TTT GCA TGC CCA CCG CCT CCC AGT CCA
T   K   V   E   V   K   R   T   G   G   G   G   S   G>

          430          440          450          460
          *          *          *          *
GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA
CCG CCT CCC AGT CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT
G   G   G   S   G   G   G   G   S   G   G   G   G   S>

          470          480          490          500
          *          *          *          *
GGT GGC GGA GGG TCA CAG ATT CAG CTG GTG CAG TCT GGA GCA
CCA CCG CCT CCC AGT GTC TAA GTC GAC CAC GTC AGA CCT CGT
G   G   G   G   S   Q   I   Q   L   V   Q   S   G   A>

          510          520          530          540
          *          *          *          *
GAG GTG AAG AAG CCT GGA TCT TCT GTG AAG GTG TCT TGT AAG
CTC CAC TTC TTC GGA CCT AGA AGA CAC TTC CAC AGA ACA TTC
E   V   K   K   P   G   S   S   V   K   V   S   C   K>

          550          560          570          580
          *          *          *          *
GCA TCT GGA TAC ACC TTC ACC GAC TAC TAC ATT AAT TGG ATG
CGT AGA CCT ATG TGG AAG TGG CTG ATG ATG TAA TTA ACC TAC
A   S   G   Y   T   F   T   D   Y   Y   I   N   W   M>

590          600          610          620          630
*          *          *          *          *
AGA CAG GCA CCT GGA CAG GGA CTC GAG TGG ATT GGA TGG ATT
TCT GTC CGT GGA CCT GTC CCT GAG CTC ACC TAA CCT ACC TAA
R   Q   A   P   G   Q   G   L   E   W   I   G   W   I>

          640          650          660          670
          *          *          *          *
GAC CCT GGA TCT GGA AAT ACA AAG TAC AAT GAG AAG TTC AAG
CTG GGA CCT AGA CCT TTA TGT TTC ATG TTA CTC TTC AAG TTC
D   P   G   S   G   N   T   K   Y   N   E   K   F   K>

          680          690          700          710
          *          *          *          *
GGA AGA GCA ACA CTG ACA GTG GAC ACA TCC ACG AAT ACC GCC
CCT TCT CGT TGT GAC TGT CAC CTG TGT AGG TGC TTA TGG CCG
G   R   A   T   L   T   V   D   T   S   T   N   T   A>

          720          730          740          750
          *          *          *          *
TAC ATG GAG CTG TCT TCT CTG AGA TCT GAG GAC ACA GCA TTC
ATG TAC CTC GAC AGA AGA GAC TCT AGA CTC CTG TGT CGT AAG
Y   M   E   L   S   S   L   R   S   E   D   T   A   F>

          760          770          780          790
          *          *          *          *
TAC TTC TGT GCA AGA GAG AAG ACC ACC TAC TAC TAC GCA ATG
ATG AAG ACA CGT TCT CTC TTC TGG TGG ATG ATG ATG CGT TAC
Y   F   C   A   R   E   K   T   T   Y   Y   Y   A   M>

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FIG. 6(contd.)

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800      810      820      830      840
*        *        *        *        *
GAC TAC TGG GGA CAG GGA ACA CTG GTG ACA GTG TCT TCT GCC
CTG ATG ACC CCT GTC CCT TGT GAC CAC TGT CAC AGA AGA CGG
D   Y   W   G   Q   G   T   L   V   T   V   S   S   A>

      850      860      870      880
      *        *        *        *
TCA ACG AAG GGC CCG ACT AGT AAC TCC ATC ATG TAC TTC AGC
AGT TGC TTC CCG GGC TGA TCA TTG AGG TAG TAC ATG AAG TCG
S   T   K   G   P   T   S   N   S   I   M   Y   F   S>

      890      900      910      920
      *        *        *        *
CAC TTC GTG CCG GTC TTC CTG CCA GCG AAG CCC ACC ACG ACG
GTG AAG CAC GGC CAG AAG GAC GGT CGC TTC GGG TGG TGC TGC
H   F   V   P   V   F   L   P   A   K   P   T   T   T

      930      940      950      960
      *        *        *        *
CCA GCG CCG CGA CCA CCA ACA CCG GCG CCC ACC ATC GCG TCG
GGT CGC GGC GCT GGT GGT TGT GGC CGC GGG TGG TAG CGC AGC
P   A   P   R   P   P   T   P   A   P   T   I   A   S>

      970      980      990      1000
      *        *        *        *
CAG CCC CTG TCC CTG CGC CCA GAG GCG GGA TCC AAG CCC TTT
GTC GGG GAC AGG GAC GCG GGT CTC CGC CCT AGG TTC GGG AAA
Q   P   L   S   L   R   P   E   A   G   S   K   P   F>

1010      1020      1030      1040      1050
*        *        *        *        *
TGG GTG CTG GTG GTG GTT GGT GGA GTC CTG GCT TGC TAT AGC
ACC CAC GAC CAC CAC CAA CCA CCT CAG GAC CGA ACG ATA TCG
W   V   L   V   V   V   G   G   V   L   A   C   Y   S>

      1060      1070      1080      1090
      *        *        *        *
TTG CTA GTA ACA GTG GCC TTT ATT ATT TTC TGG GTG AGG AGT
AAC GAT CAT TGT CAC CGG AAA TAA TAA AAG ACC CAC TCC TCA
L   L   V   T   V   A   F   I   I   F   W   V   R   S>

      1100      1110      1120      1130
      *        *        *        *
AAG AGG AGC AGG CTC CTG CAC AGT GAC TAC ATG AAC ATG ACT
TTC TCC TCG TCC GAG GAC GTG TCA CTG ATG TAC TTG TAC TGA
K   R   S   R   L   L   H   S   D   Y   M   N   M   T>

      1140      1150      1160      1170
      *        *        *        *
CCC CGC CGC CCC GGG CCC ACC CGC AAG CAT TAC CAG CCC TAT
GGG GCG GCG GGG GGG CCC GGG TGG GCG TTC GTA ATG GTC GGG ATA
P   R   R   P   G   P   T   R   K   H   Y   Q   P   Y>

      1180      1190      1200      1210
      *        *        *        *
GCC CCA CCA CGC GAC TTC GCA GCC TAT CGC TCC TGA
CGG GGT GGT GCG CTG AAG CGT CGG ATA GCG AGG ACT
A   P   P   R   D   F   A   A   Y   R   S   *

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FIG. 7
SEQUENCE OF hCTMO1 / G1 / ZETA RECOMBINANT CHIMERIC
RECEPTOR

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      10      20      30      40
      *      *      *      *
ATG TCT GTC CCC ACC CAA GTC CTC GGA CTC CTG CTG CTG TGG CTT ACA
TAC AGA CAG GGG TGG GTT CAG GAG CCT GAG GAC GAC GAC ACC GAA TGT
M  S  V  P  T  Q  V  L  G  L  L  L  L  W  L  T>

50      60      70      80      90
  *      *      *      *      *
GAT GCC AGA TGC GAT ATC CAG ATG ACT CAG AGT CCA AGT ACT CTC AGT
CTA CGG TCT ACG CTA TAG GTC TAC TGA GTC TCA GGT TCA TGA GAG TCA
D  A  R  C  D  I  Q  M  T  Q  S  P  S  T  L  S>

100     110     120     130     140
  *      *      *      *      *
GCC AGT GTA GGT GAT AGG GTC ACC ATC ACT TGT AGG AGT AGT AAA AGT
CGG TCA CAT CCA CTA TCC CAG TGG TAG TGA ACA TCC TCA TCA TTT TCA
A  S  V  G  D  R  V  T  I  T  C  R  S  S  K  S>

150     160     170     180     190
  *      *      *      *      *
CTC CTC CAT AGT AAC GGT GAC ACC TTC CTC TAT TGG TTC CAG CAG AAA
GAG GAG GTA TCA TTG CCA CTG TGG AAG GAG ATA ACC AAG GTC GTC TTT
L  L  H  S  N  G  D  T  F  L  Y  W  F  Q  Q  K>

200     210     220     230     240
  *      *      *      *      *
CCA GGT AAA GCC CCA AAG CTC CTC ATG TAT AGG ATG AGT AAC CTC GCC
GGT CCA TTT CGG GGT TTC GAG GAG TAC ATA TCC TAC TCA TTG GAG CGG
P  G  K  A  P  K  L  L  M  Y  R  M  S  N  L  A>

250     260     270     280
  *      *      *      *
AGT GGT GTA CCA TCT AGA TTC AGT GGT AGT GGT AGT GGT ACT GAG TTC
TCA CCA CAT GGT AGA TCT AAG TCA CCA TCA CCA TCA CCA TGA CTC AAG
S  G  V  P  S  R  F  S  G  S  G  S  G  T  E  F>

290     300     310     320     330
  *      *      *      *      *
ACT CTC ACT ATC AGT AGT CTC CAG CCA GAT GAT TTC GCC ACT TAT TAT
TGA GAG TGA TAG TCA TCA GAG GTC GGT CTA CTA AAG CGG TGA ATA ATA
T  L  T  I  S  S  L  Q  P  D  D  F  A  T  Y  Y>

340     350     360     370     380
  *      *      *      *      *
TGT ATG CAG CAT CTC GAA TAT CCA TTC ACT TTC GGT CAG GGT ACT AAA
ACA TAC GTC GTA GAG CTT ATA GGT AAG TGA AAG CCA GTC CCA TGA TTT
C  M  Q  H  L  E  Y  P  F  T  F  G  Q  G  T  K>

390     400     410     420     430
  *      *      *      *      *
GTA GAA GTA AAA CGT ACG GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA
CAT CTT CAT TTT GCA TGC CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT
V  E  V  K  R  T  G  G  G  G  S  G  G  G  G  S>

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FIG. 7 (contd.)

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      440      450      460      470      480
      *      *      *      *      *
GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA CAG
CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT GTC
G  G  G  G  S  G  G  G  G  S  G  G  G  G  S  Q>

      490      500      510      520
      *      *      *      *
ATT CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG CCT GGA TCT TCT
TAA GTC GAC CAC GTC AGA CCT CGT CTC CAC TTC TTC GGA CCT AGA AGA
I  Q  L  V  Q  S  G  A  E  V  K  K  P  G  S  S>

530      540      550      560      570
*      *      *      *      *
GTG AAG GTG TCT TGT AAG GCA TCT GGA TAC ACC TTC ACC GAC TAC TAC
CAC TTC CAC AGA ACA TTC CGT AGA CCT ATG TGG AAG TGG CTG ATG ATG
V  K  V  S  C  K  A  S  G  Y  T  F  T  D  Y  Y>

      580      590      600      610      620
      *      *      *      *      *
ATT AAT TGG ATG AGA CAG GCA CCT GGA CAG GGA CTC GAG TGG ATT GGA
TAA TTA ACC TAC TCT GTC CGT GGA CCT GTC CCT GAG CTC ACC TAA CCT
I  N  W  M  R  Q  A  P  G  Q  G  L  E  W  I  G>

      630      640      650      660      670
      *      *      *      *      *
TGG ATT GAC CCT GGA TCT GGA AAT ACA AAG TAC AAT GAG AAG TTC AAG
ACC TAA CTG GGA CCT AGA CCT TTA TGT TTC ATG TTA CTC TTC AAG TTC
W  I  D  P  G  S  G  N  T  K  Y  N  E  K  F  K>

      680      690      700      710      720
      *      *      *      *      *
GGA AGA GCA ACA CTG ACA GTG GAC ACA TCC ACG AAT ACC GCC TAC ATG
CCT TCT CGT TGT GAC TGT CAC CTG TGT AGG TGC TTA TGG CCG ATG TAC
G  R  A  T  L  T  V  D  T  S  T  N  T  A  Y  M>

      730      740      750      760
      *      *      *      *
GAG CTG TCT TCT CTG AGA TCT GAG GAC ACA GCA TTC TAC TTC TGT GCA
CTC GAC AGA AGA GAC TCT AGA CTC CTG TGT CGT AAG ATG AAG ACA CGT
E  L  S  S  L  R  S  E  D  T  A  F  Y  F  C  A>

770      780      790      800      810
*      *      *      *      *
AGA GAG AAG ACC ACC TAC TAC TAC GCA ATG GAC TAC TGG GGA CAG GGA
TCT CTC TTC TGG TGG ATG ATG ATG CGT TAC CTG ATG ACC CCT GTC CCT
R  E  K  T  T  Y  Y  Y  A  M  D  Y  W  G  Q  G>

      820      830      840      850      860
      *      *      *      *      *
ACA CTG GTG ACA GTG TCT TCT GCC TCA ACG AAG GGC CCG ACT AGT GAC
TGT GAC CAC TGT CAC AGA AGA CGG AGT TGC TTC CCG GGC TGA TCA CTG
T  L  V  T  V  S  S  A  S  T  K  G  P  T  S  D>

      870      880      890      900      910
      *      *      *      *      *
AAA ACT CAC ACA TGC CCA CCG TGC CCA GCA CCT GAA CTC CTG GGG GGA
TTT TGA GTG TGT ACG GGT GGC ACG GGT CGT GGA CTT GAG GAC CCC CCT
K  T  H  T  C  P  P  C  P  A  P  E  L  L  G  G>

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FIG. 7 (contd.)

920	930	940	950	960
CCG TCA GTC TTC CTC	TTC CCC CCA AAA CCC AAG GAC ACC CTC ATG ATC			
GGC AGT CAG AAG GAG AAG GGG GGT TTT GGG TTC CTG TGG GAG TAC TAG				
P S V F L F P P K P K D T L M I>				
970	980	990	1000	
TCC CGG ACC CCT GAG GTC ACA TGC GTG GTG GTG GAC GTG AGC CAC GAA				
AGG GCC TGG GGA CTC CAG TGT ACG CAC CAC CAC CTG CAC TCG GTG CTT				
S R T P E V T C V V V D V S H E>				
1010	1020	1030	1040	1050
GAC CCT GAG GTC AAG TTC AAC TGG TAC GTG GAC GGC GTG GAG GTG CAT				
CTG GGA CTC CAG TTC AAG TTG ACC ATG CAC CTG CCG CAC CTC CAC GTA				
D P E V K F N W Y V D G V E V H>				
1060	1070	1080	1090	1100
AAT GCC AAG ACA AAG CCG CGG GAG GAG CAG TAC AAC AGC ACG TAC CGT				
TTA CGG TTC TGT TTC GGC GCC CTC CTC GTC ATG TTG TCG TGC ATG GCA				
N A K T K P R E E Q Y N S T Y R>				
1110	1120	1130	1140	1150
GTG GTC AGC GTC CTC ACC GTC CTG CAC CAG GAC TGG CTG AAT GGC AAG				
CAC CAG TCG CAG GAG TGG CAG GAC GTG GTC CTG ACC GAC TTA CCG TTC				
V V S V L T V L H Q D W L N G K>				
1160	1170	1180	1190	1200
GAG TAC AAG TGC AAG GTC TCC AAC AAA GCC CTC CCA GCC CCC ATC GAG				
CTC ATG TTC ACG TTC CAG AGG TTG TTT CGG GAG GGT CGG GGG TAG CTC				
E Y K C K V S N K A L P A P I E>				
1210	1220	1230	1240	
AAA ACC ATC TCC AAA GCC AAA GGG CAG CCC CGA GAA CCA CAG GTG TAC				
TTT TGG TAG AGG TTT CGG TTT CCC GTC GGG GCT CTT GGT GTC CAC ATG				
K T I S K A K G Q P R E P Q V Y>				
1250	1260	1270	1280	1290
ACC CTG CCC CCA TCC CGG GAG GAG ATG ACC AAG AAC CAG GTC AGC CTG				
TGG GAC GGG GGT AGG GCC CTC CTC TAC TGG TTC TTG GTC CAG TCG GAC				
T L P P S R E E M T K N Q V S L>				
1300	1310	1320	1330	1340
ACC TGC CTG GTC AAA GGC TTC TAT CCC AGC GAC ATC GCC GTG GAG TGG				
TGG ACG GAC CAG TTT CCG AAG ATA GGG TCG CTG TAG CGG CAC CTC ACC				
T C L V K G F Y P S D I A V E W>				
1350	1360	1370	1380	1390
GAG AGC AAT GGG CAG CCG GAG AAC AAC TAC AAG ACC ACG CCT CCC GTG				
CTC TCG TTA CCC GTC GGC CTC TTG TTG ATG TTC TGG TGC GGA GGG CAC				
E S N G Q P E N N Y K T T P P V>				
1400	1410	1420	1430	1440
CTG GAC TCC GAC GGC TCC TTC TTC CTC TAC AGC AAG CTC ACC GTG GAC				
GAC CTG AGG CTG CCG AGG AAG AAG GAG ATG TCG TTC GAG TGG CAC CTG				
L D S D G S F F L Y S K L T V D>				

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1450	1460	1470	1480
AAG AGC AGG TGG CAG	CAG GGG AAC GTC	TTC TCA TGC TCC	GTG ATG CAT
TTC TCG TCC ACC GTC	GTC CCC TTG CAG	AAG AGT ACG AGG	CAC TAC GTA
K S R W Q Q	G N V F S C	S V M H>	
1490	1500	1510	1520
GAG GCT CTG CAC AAC	CAC TAC ACG CAG	AAG AGC CTC TCC	CTG TCT CCG
CTC CGA GAC GTG TTG	GTG ATG TGC GTC	TTC TCG GAG AGG	GAC AGA GGC
E A L H N H	Y T Q K S L S	L S P>	
1540	1550	1560	1570
GGT AAA CTG GAT CCC	AAA CTC TGC TAC	CTG CTG GAT GGA	ATC CTC TTC
CCA TTT GAC CTA GGG	TTT GAG ACG ATG	GAC GAC CTA CCT	TAG GAG AAG
G K L D P K	L C Y L L D	G I L F>	
1590	1600	1610	1620
ATC TAT GGT GTC ATT	CTC ACT GCC TTG	TTC CTG AGA GTG	AAG TTC AGC
TAG ATA CCA CAG TAA	GAG TGA CCG AAC	AAG GAC TCT CAC	TTC AAG TCG
I Y G V I L T A	L F L R V K F	S>	
1640	1650	1660	1670
AGG AGC GCA GAC GCC	CCC GCG TAC CAG	CAG GGC CAG AAC	CAG CTC TAT
TCC TCG CGT CTG CCG	GGG CGC ATG GTC	GTC CCG GTC TTG	GTC GAG ATA
R S A D A P A	Y Q Q G Q N	Q L Y>	
1690	1700	1710	1720
AAC GAG CTC AAT CTA	GGA CGA AGA GAG	GAG TAC GAT GTT	TTG GAC AAG
TTG CTC GAG TTA GAT	CCT GCT TCT CTC	CTC ATG CTA CAA	AAC CTG TTC
N E L N L G R R	E E Y D V L D	K>	
1730	1740	1750	1760
AGA CGT GGC CGG GAC	CCT GAG ATG GGG	GGA AAG CCG AGA	AGG AAG AAC
TCT GCA CCG GCC CTG	GGA CTC TAC CCC	CCT TTC GGC TCT	TCC TTC TTG
R R G R D P E M	G G K P R R K	N>	
1780	1790	1800	1810
CCT CAG GAA GGC CTG	TAC AAT GAA CTG	CAG AAA GAT AAG	ATG GCG GAG
GGA GTC CTT CCG GAC	ATG TTA CTT GAC	GTC TTT CTA TTC	TAC CCG CTC
P Q E G L Y N E	L Q K D K M A	E>	
1830	1840	1850	1860
GCC TAC AGT GAG ATT	GGG ATG AAA GGC	GAG CGC CCG AGG	GGC AAG GGG
CGG ATG TCA CTC TAA	CCC TAC TTT CCG	CTC GCG GCC TCC	CCG TTC CCC
A Y S E I G M K	G E R R R G K	G>	
1880	1890	1900	1910
CAC GAT GGC CTT TAC	CAG GGT CTC AGT	ACA GCC ACC AAG	GAC ACC TAC
GTG CTA CCG GAA ATG	GTC CCA GAG TCA	TGT CCG TGG TTC	CTG TGG ATG
H D G L Y Q G L	S T A T K D T	Y>	
1930	1940	1950	
GAC GCC CTT CAC ATG	CAG GCC CTG CCC	CCT CGC TAA	
CTG CCG GAA GTG TAC	GTC CCG GAC GGG	GGA GCG ATT	
D A L H M Q A L	P P R *		

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FIG. 8

SEQUENCE OF hCTMO1/G1/ZETA-CD28 FUSION RECOMBINANT
CHIMERIC RECEPTOR

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      10      20      30      40
      *      *      *      *
ATG TCT GTC CCC ACC CAA GTC CTC GGA CTC CTG CTG CTG TGG CTT ACA
TAC AGA CAG GGG TGG GTT CAG GAG CCT GAG GAC GAC GAC ACC GAA TGT
M S V P T Q V L G L L L L W L T>

50      60      70      80      90
*      *      *      *      *
GAT GCC AGA TGC GAT ATC CAG ATG ACT CAG AGT CCA AGT ACT CTC AGT
CTA CGG TCT ACG CTA TAG GTC TAC TGA GTC TCA GGT TCA TGA GAG TCA
D A R C D I Q M T Q S P S T L S>

100      110      120      130      140
*      *      *      *      *
GCC AGT GTA GGT GAT AGG GTC ACC ATC ACT TGT AGG AGT AGT AAA AGT
CGG TCA CAT CCA CTA TCC CAG TGG TAG TGA ACA TCC TCA TCA TTT TCA
A S V G D R V T I T C R S S K S>

150      160      170      180      190
*      *      *      *      *
CTC CTC CAT AGT AAC GGT GAC ACC TTC CTC TAT TGG TTC CAG CAG AAA
GAG GAG GTA TCA TTG CCA CTG TGG AAG GAG ATA ACC AAG GTC GTC TTT
L L H S N G D T F L Y W F Q Q K>

200      210      220      230      240
*      *      *      *      *
CCA GGT AAA GCC CCA AAG CTC CTC ATG TAT AGG ATG AGT AAC CTC GCC
GGT CCA TTT CGG GGT TTC GAG GAG TAC ATA TCC TAC TCA TTG GAG CGG
P G K A P K L L M Y R M S N L A>

250      260      270      280
*      *      *      *
AGT GGT GTA CCA TCT AGA TTC AGT GGT AGT GGT AGT GGT ACT GAG TTC
TCA CCA CAT GGT AGA TCT AAG TCA CCA TCA CCA TCA TGA TGA TGA AAG
S G V P S R F S G S G S G T E F>

290      300      310      320      330
*      *      *      *      *
ACT CTC ACT ATC AGT AGT CTC CAG CCA GAT GAT TTC GCC ACT TAT TAT
TGA GAG TGA TAG TCA TCA GAG GTC GGT CTA CTA AAG CGG TGA ATA ATA
T L T I S S L Q P D D F A T Y Y>

340      350      360      370      380
*      *      *      *      *
TGT ATG CAG CAT CAG GAA TAT CCA TTC ACT TTC GGT CAG GGT ACT AAA
ACA TAC GTC GTA GAG CTT ATA GGT AAG TGA AAG CCA GTC CCA TGA TTT
C M Q H L E Y P F T F G Q G T K>

390      400      410      420      430
*      *      *      *      *
GTA GAA GTA AAA CGT ACG GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA
CAT CTT CAT TTT GCA TGC CCA CCG CCT CCC AGT CCA CCG CCT CCC TCA
V E V K R T G G G G S G G G G S>

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FIG. 8(contd.)

440	450	460	470	480
* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>
490	500	510	520	
* ATT CAG CTG GTG CAG TCT TAA GTC GAC CAC GTC AGA I Q L V Q S G A E V K K P G S S>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	* GGT GGC GGA GGG TCA CCA CCG CCT CCC AGT G G G G S G G G S G G G S Q>	
530	540	550	560	570
* GTG AAG GTG TCT TGT AAG CAC TTC CAC AGA ACA TTC V K V S C K A S G Y T F T D Y Y>	* GTG AAG GTG TCT TGT AAG CAC TTC CAC AGA ACA TTC V K V S C K A S G Y T F T D Y Y>	* GTG AAG GTG TCT TGT AAG CAC TTC CAC AGA ACA TTC V K V S C K A S G Y T F T D Y Y>	* GTG AAG GTG TCT TGT AAG CAC TTC CAC AGA ACA TTC V K V S C K A S G Y T F T D Y Y>	* GTG AAG GTG TCT TGT AAG CAC TTC CAC AGA ACA TTC V K V S C K A S G Y T F T D Y Y>
580	590	600	610	620
* ATT AAT TGG ATG AGA CAG TAA TTA ACC TAC TCT GTC I N W M R Q A P G Q G L E W I G>	* ATT AAT TGG ATG AGA CAG TAA TTA ACC TAC TCT GTC I N W M R Q A P G Q G L E W I G>	* ATT AAT TGG ATG AGA CAG TAA TTA ACC TAC TCT GTC I N W M R Q A P G Q G L E W I G>	* ATT AAT TGG ATG AGA CAG TAA TTA ACC TAC TCT GTC I N W M R Q A P G Q G L E W I G>	* ATT AAT TGG ATG AGA CAG TAA TTA ACC TAC TCT GTC I N W M R Q A P G Q G L E W I G>
630	640	650	660	670
* TGG ATT GAC CCT GGA TCT ACC TAA CTG GGA CCT AGA W I D P G S G N T K Y N E K F K>	* TGG ATT GAC CCT GGA TCT ACC TAA CTG GGA CCT AGA W I D P G S G N T K Y N E K F K>	* TGG ATT GAC CCT GGA TCT ACC TAA CTG GGA CCT AGA W I D P G S G N T K Y N E K F K>	* TGG ATT GAC CCT GGA TCT ACC TAA CTG GGA CCT AGA W I D P G S G N T K Y N E K F K>	* TGG ATT GAC CCT GGA TCT ACC TAA CTG GGA CCT AGA W I D P G S G N T K Y N E K F K>
680	690	700	710	720
* GGA AGA GCA ACA CTG ACA CCT TCT CGT TGT GAC TGT G R A T L T V D T S T N T A Y M>	* GGA AGA GCA ACA CTG ACA CCT TCT CGT TGT GAC TGT G R A T L T V D T S T N T A Y M>	* GGA AGA GCA ACA CTG ACA CCT TCT CGT TGT GAC TGT G R A T L T V D T S T N T A Y M>	* GGA AGA GCA ACA CTG ACA CCT TCT CGT TGT GAC TGT G R A T L T V D T S T N T A Y M>	* GGA AGA GCA ACA CTG ACA CCT TCT CGT TGT GAC TGT G R A T L T V D T S T N T A Y M>
730	740	750	760	
* GAG CTG TCT TCT CTG AGA CTC GAC AGA AGA GAC TCT E L S S L R S E D T A F Y F C A>	* GAG CTG TCT TCT CTG AGA CTC GAC AGA AGA GAC TCT E L S S L R S E D T A F Y F C A>	* GAG CTG TCT TCT CTG AGA CTC GAC AGA AGA GAC TCT E L S S L R S E D T A F Y F C A>	* GAG CTG TCT TCT CTG AGA CTC GAC AGA AGA GAC TCT E L S S L R S E D T A F Y F C A>	
770	780	790	800	810
* AGA GAG AAG ACC ACC TAC TCT CTC TTC TGG TGG ATG R E K T T Y Y Y A M D Y W G Q G>	* AGA GAG AAG ACC ACC TAC TCT CTC TTC TGG TGG ATG R E K T T Y Y Y A M D Y W G Q G>	* AGA GAG AAG ACC ACC TAC TCT CTC TTC TGG TGG ATG R E K T T Y Y Y A M D Y W G Q G>	* AGA GAG AAG ACC ACC TAC TCT CTC TTC TGG TGG ATG R E K T T Y Y Y A M D Y W G Q G>	* AGA GAG AAG ACC ACC TAC TCT CTC TTC TGG TGG ATG R E K T T Y Y Y A M D Y W G Q G>
820	830	840	850	860
* ACA CTG GTG ACA GTG TCT TGT GAC CAC TGT CAC AGA T L V T V S S A S T K G P T S D>	* ACA CTG GTG ACA GTG TCT TGT GAC CAC TGT CAC AGA T L V T V S S A S T K G P T S D>	* ACA CTG GTG ACA GTG TCT TGT GAC CAC TGT CAC AGA T L V T V S S A S T K G P T S D>	* ACA CTG GTG ACA GTG TCT TGT GAC CAC TGT CAC AGA T L V T V S S A S T K G P T S D>	* ACA CTG GTG ACA GTG TCT TGT GAC CAC TGT CAC AGA T L V T V S S A S T K G P T S D>
870	880	890	900	910
* AAA ACT CAC ACA TGC CCA TTT TGA GTG TGT ACG GGT K T H T C P P C P A P E L L G G>	* AAA ACT CAC ACA TGC CCA TTT TGA GTG TGT ACG GGT K T H T C P P C P A P E L L G G>	* AAA ACT CAC ACA TGC CCA TTT TGA GTG TGT ACG GGT K T H T C P P C P A P E L L G G>	* AAA ACT CAC ACA TGC CCA TTT TGA GTG TGT ACG GGT K T H T C P P C P A P E L L G G>	* AAA ACT CAC ACA TGC CCA TTT TGA GTG TGT ACG GGT K T H T C P P C P A P E L L G G>

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FIG. 8(contd.)

920	930	940	950	960
CCG TCA GTC TTC CTC TTC CCC CCA AAA CCC AAG GAC ACC CTC ATG ATC				
GGC AGT CAG AAG GAG AAG GGG GGT TTT GGG TTC CTG TGG GAG TAC TAG				
P S V F L F P P K P K D T L M I>				
970	980	990	1000	
TCC CGG ACC CCT GAG GTC ACA TGC GTG GTG GTG GAC GTG AGC CAC GAA				
AGG GCC TGG GGA CTC CAG TGT ACG CAC CAC CAC CTG CAC TCG GTG CTT				
S R T P E V T C V V V D V S H E>				
1010	1020	1030	1040	1050
GAC CCT GAG GTC AAG TTC AAC TGG TAC GTG GAC GGC GTG GAG GTG CAT				
CTG GGA CTC CAG TTC AAG TTG ACC ATG CAC CTG CCG CAC CTC CAC GTA				
D P E V K F N W Y V D G V E V H>				
1060	1070	1080	1090	1100
AAT GCC AAG ACA AAG CCG CGG GAG GAG CAG TAC AAC AGC ACG TAC CGT				
TTA CGG TTC TGT TTC GGC GCC CTC CTC GTC ATG TTG TCG TGC ATG GCA				
N A K T K P R E E Q Y N S T Y R>				
1110	1120	1130	1140	1150
GTG GTC AGC GTC CTC ACC GTC CTG CAC CAG GAC TGG CTG AAT GGC AAG				
CAC CAG TCG CAG GAG TGG CAG GAC GTG GTC CTG ACC GAC TTA CCG TTC				
V V S V L T V L H Q D W L N G K>				
1160	1170	1180	1190	1200
GAG TAC AAG TGC AAG GTC TCC AAC AAA GCC CTC CCA GCC CCC ATC GAG				
CTC ATG TTC ACG TTC CAG AGG TTG TTT CGG GAG GGT CGG GGG TAG CTC				
E Y K C K V S N K A L P A P I E>				
1210	1220	1230	1240	
AAA ACC ATC TCC AAA GCC AAA GGG CAG CCC CGA GAA CCA CAG GTG TAC				
TTT TGG TAG AGG TTT CGG TTT CCC GTC GGG GCT CTT GGT GTC CAC ATG				
K T I S K A K G Q P R E P Q V Y>				
1250	1260	1270	1280	1290
ACC CTG CCC CCA TCC CGG GAG GAG ATG ACC AAG AAC CAG GTC AGC CTG				
TGG GAC GGG GGT AGG GCC CTC CTC TAC TGG TTC TTG GTC CAG TCG GAC				
T L P P S R E E M T K N Q V S L>				
1300	1310	1320	1330	1340
ACC TGC CTG GTC AAA GGC TTC TAT CCC AGC GAC ATC GCC GTG GAG TGG				
TGG ACG GAC CAG TTT CCG AAG ATA GGG TCG CTG TAG CGG CAC CTC ACC				
T C L V K G F Y P S D I A V E W>				
1350	1360	1370	1380	1390
GAG AGC AAT GGG CAG CCG GAG AAC AAC TAC AAG ACC ACG CCT CCC GTG				
CTC TCG TTA CCC GTC GGC CTC TTG TTG ATG TTC TGG TGC GGA GGG CAC				
E S N G Q P E N N Y K T T P P V>				
1400	1410	1420	1430	1440
CTG GAC TCC GAC GGC TCC TTC TTC CTC TAC AGC AAG CTC ACC GTG GAC				
GAC CTG AGG CTG CCG AGG AAG AAG GAG ATG TCG TTC GAG TGG CAC CTG				
L D S D G S F F L Y S K L T V D>				

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FIG. 8(contd.)

1450	1460	1470	1480
AAG AGC AGG TGG CAG CAG GGG AAC GTC TTC TCA TGC TCC GTG ATG CAT			
TTC TCG TCC ACC GTC GTC CCC TTG CAG AAG AGT ACG AGG CAC TAC GTA			
K S R W Q Q G N V F S C S V M H>			
1490	1500	1510	1520
GAG GCT CTG CAC AAC CAC TAC ACG CAG AAG AGC CTC TCC CTG TCT CCG			
CTC CGA GAC GTG TTG GTG ATG TGC GTC TTC TCG GAG AGG GAC AGA GGC			
E A L H N H Y T Q K S L S L S P>			
1540	1550	1560	1570
GGT AAA CTG GAT CCC AAA CTC TGC TAC CTG CTG GAT GGA ATC CTC TTC			
CCA TTT GAC CTA GGG TTT GAG ACG ATG GAC GAC CTA CCT TAG GAG AAG			
G K L D P K L C Y L L D G I L F>			
1590	1600	1610	1620
ATC TAT GGT GTC ATT CTC ACT GCC TTG TTC CTG AGA GTG AAG TTC AGC			
TAG ATA CCA CAG TAA GAG TGA CGG AAC AAG GAC TCT CAC TTC AAG TCG			
I Y G V I L T A L F L R V K F S>			
1640	1650	1660	1670
AGG AGC GCA GAC GCC CCC GCG TAC CAG CAG GGC CAG AAC CAG CTC TAT			
TCC TCG CGT CTG CGG GGG CGC ATG GTC GTC CCG GTC TTG GTC GAG ATA			
R S A D A P A Y Q Q G Q N Q L Y>			
1690	1700	1710	1720
AAC GAG CTC AAT CTA GGA CGA AGA GAG GAG TAC GAT GTT TTG GAC AAG			
TTG CTC GAG TTA GAT CCT GCT TCT CTC CTC ATG CTA CAA AAC CTG TTC			
N E L N L G R R E E Y D V L D K>			
1730	1740	1750	1760
AGA CGT GGC CGG GAC CCT GAG ATG GGG GGA AAG CCG AGA AGG AAG AAC			
TCT GCA CCG GCC CTG GGA CTC TAC CCC CCT TTC GGC TCT TCC TTC TTG			
R R G R D P E M G G K P R R K N>			
1780	1790	1800	1810
CCT CAG GAA GGC CTG TAC AAT GAA CTG CAG AAA GAT AAG ATG GCG GAG			
GGA GTC CTT CCG GAC ATG TTA CTT GAC GTC TTT CTA TTC TAC CGC CTC			
P Q E G L Y N E L Q K D K M A E>			
1830	1840	1850	1860
GCC TAC AGT GAG ATT GGG ATG AAA GGC GAG CGC CGG AGG GGC AAG GGG			
CGG ATG TCA CTC TAA CCC TAC TTT CCG CTC GCG GCC TCC CCG TTC CCC			
A Y S E I G M K G E R R R G K G>			
1880	1890	1900	1910
CAC GAT GGC CTT TAC CAG GGT CTC AGT ACA GCC ACC AAG GAC ACC TAC			
GTG CTA CCG GAA ATG GTC CCA GAG TCA TGT CCG TGG TTC CTG TGG ATG			
H D G L Y Q G L S T A T K D T Y>			

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	1930				1940				1950				1960								
	*				*				*				*								
	GAC	GCC	CTT	CAC	ATG	CAG	GCC	CTG	CCC	CCT	CGC	AGG	AGT	AAG	AGG	AGC					
	CTG	CGG	GAA	GTG	TAC	GTC	CGG	GAC	GGG	GGA	GCG	TCC	TCA	TTC	TCC	TCG					
	D	A	L	H	M	Q	A	L	P	P	R	R	S	K	R	S>					
1970					1980				1990				2000				2010				
	*				*				*				*				*				
	AGG	CTC	CTG	CAC	AGT	GAC	TAC	ATG	AAC	ATG	ACT	CCC	CGC	CGC	CCC	GGG					
	TCC	GAG	GAC	GTG	TCA	CTG	ATG	TAC	TTG	TAC	TGA	GGG	GCG	GCG	GGG	CCC					
	R	L	L	H	S	D	Y	M	N	M	T	P	R	R	P	G>					
	2020				2030				2040				2050				2060				
	*				*				*				*				*				
	CCC	ACC	CGC	AAG	CAT	TAC	CAG	CCC	TAT	GCC	CCA	CCA	CGC	GAC	TTC	GCA					
	GGG	TGG	GCG	TTC	GTA	ATG	GTC	GGG	ATA	CGG	GGT	GGT	GCG	CTG	AAG	CGT					
	P	T	R	K	H	Y	Q	P	Y	A	P	P	R	D	F	A>					
	2070																				
	*																				
	GCC	TAT	CGC	TCC	TGA																
	CGG	ATA	GCG	AGG	ACT																
	A	Y	R	S	*																

FIG. 8 (contd.)

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FIG. 9

SEQUENCE OF hCTMO1 / h / CD28 RECOMBINANT CHIMERIC
RECEPTOR

```

          10          20          30          40
          *          *          *          *
ATG TCT GTC CCC ACC CAA GTC CTC GGA CTC CTG CTG CTG TGG CTT ACA
TAC AGA CAG GGG TGG GTT CAG GAG CCT GAG GAC GAC GAC ACC GAA TGT
M   S   V   P   T   Q   V   L   G   L   L   L   L   W   L   T>

50          60          70          80          90
*          *          *          *          *
GAT GCC AGA TGC GAT ATC CAG ATG ACT CAG AGT CCA AGT ACT CTC AGT
CTA CGG TCT ACG CTA TAG GTC TAC TGA GTC TCA GGT TCA TGA GAG TCA
D   A   R   C   D   I   Q   M   T   Q   S   P   S   T   L   S>

100         110         120         130         140
*          *          *          *          *
GCC AGT GTA GGT GAT AGG GTC ACC ATC ACT TGT AGG AGT AGT AAA AGT
CGG TCA CAT CCA CTA TCC CAG TGG TAG TGA ACA TCC TCA TCA TTT TCA
A   S   V   G   D   R   V   T   I   T   C   R   S   S   K   S>

150         160         170         180         190
*          *          *          *          *
CTC CTC CAT AGT AAC GGT GAC ACC TTC CTC TAT TGG TTC CAG CAG AAA
GAG GAG GTA TCA TTG CCA CTG TGG AAG GAG ATA ACC AAG GTC GTC TTT
L   L   H   S   N   G   D   T   F   L   Y   W   F   Q   Q   K>

200         210         220         230         240
*          *          *          *          *
CCA GGT AAA GCC CCA AAG CTC CTC ATG TAT AGG ATG AGT AAC CTC GCC
GGT CCA TTT CGG GGT TTC GAG GAG TAC ATA TCC TAC TCA TTG GAG CGG
P   G   K   A   P   K   L   L   M   Y   R   M   S   N   L   A>

250         260         270         280
*          *          *          *
AGT GGT GTA CCA TCT AGA TTC AGT GGT AGT GGT AGT GGT ACT GAG TTC
TCA CCA CAT GGT AGA TCT AAG TCA CCA TCA CCA TCA CCA TGA CTC AAG
S   G   V   P   S   R   F   S   G   S   G   S   G   T   E   F>

290         300         310         320         330
*          *          *          *          *
ACT CTC ACT ATC AGT AGT CTC CAG CCA GAT GAT TTC GCC ACT TAT TAT
TGA GAG TGA TAG TCA TCA GAG GTC GGT CTA CTA AAG CGG TGA ATA ATA
T   L   T   I   S   S   L   Q   P   D   D   F   A   T   Y   Y>

340         350         360         370         380
*          *          *          *          *
TGT ATG CAG CAT CTC GAA TAT CCA TTC ACT TTC GGT CAG GGT ACT AAA
ACA TAC GTC GTA GAG CTT ATA GGT AAG TGA AAG CCA GTC CCA TGA TTT
C   M   Q   H   L   E   Y   P   F   T   F   G   Q   G   T   K>

390         400         410         420         430
*          *          *          *          *
GTA GAA GTA AAA CGT ACG GGT GGC GGA GGG TCA GGT GGC GGA GGC TCA
CAT CTT CAT TTT GCA TGC CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT
V   E   V   K   R   T   G   G   G   G   S   G   G   G   G   S>

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FIG. 9(contd.)

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      440      450      460      470      480
      *      *      *      *      *
GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA GGT GGC GGA GGG TCA CAG
CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT CCA CCG CCT CCC AGT GTC
G G G G S G G G S G G G G S Q>

      490      500      510      520
      *      *      *      *
ATT CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG CCT GGA TCT TCT
TAA GTC GAC CAC GTC AGA CCT CGT CTC CAC TTC TTC GGA CCT AGA AGA
I Q L V Q S G A E V K K P G S S>

530      540      550      560      570
*      *      *      *      *
GTG AAG GTG TCT TGT AAG GCA TCT GGA TAC ACC TTC ACC GAC TAC TAC
CAC TTC CAC AGA ACA TTC CGT AGA CCT ATG TGG AAG TGG CTG ATG ATG
V K V S C K A S G Y T F T D Y Y>

      580      590      600      610      620
      *      *      *      *      *
ATT AAT TGG ATG AGA CAG GCA CCT GGA CAG GGA CTC GAG TGG ATT GGA
TAA TTA ACC TAC TCT GTC CGT GGA CCT GTC CCT GAG CTC ACC TAA CCT
I N W M R Q A P G Q G L E W I G>

      630      640      650      660      670
      *      *      *      *      *
TGG ATT GAC CCT GGA TCT GGA AAT ACA AAG TAC AAT GAG AAG TTC AAG
ACC TAA CTG GGA CCT AGA CCT TTA TGT TTC ATG TTA CTC TTC AAG TTC
W I D P G S G N T K Y N E K F K>

      680      690      700      710      720
      *      *      *      *      *
GGA AGA GCA ACA CTG ACA GTG GAC ACA TCC ACG AAT ACC GCC TAC ATG
CCT TCT CGT TGT GAC AGT CAC CTG TGT AGG TGC TTA TGG CGG ATG TAC
G R A T L T V D T S T N T A Y M>

      730      740      750      760
      *      *      *      *
GAG CTG TCT TCT CTG AGA TCT GAG GAC ACA GCA TTC TAC TTC TGT GCA
CTC GAC AGA AGA GAC TCT AGA CTC CTG TGT CGT AAG ATG AAG ACA CGT
E L S S L R S E D T A F Y F C A>

770      780      790      800      810
*      *      *      *      *
AGA GAG AAG ACC ACC TAC TAC TAC GCA ATG GAC TAC TGG GGA CAG GGA
TCT CTC TTC TGG TGG ATG ATG ATG CGT TAC CTG ATG ACC CCT GTC CCT
R E K T T Y Y Y A M D Y W G Q G>

      820      830      840      850      860
      *      *      *      *      *
ACA CTG GTG ACA GTG TCT TCT GCC TCA ACG AAG GGC CCG ACT AGT GAC
TGT GAC CAC TGT CAC AGA AGA CGG AGT TGC TTC CCG GCC TGA TCA CTG
T L V T V S S A S T K G P T S D>

      870      880      890      900      910
      *      *      *      *      *
AAA ACT CAC ACA TGC CCA CCG TGC CCA AAA GGG AAA CAC CTT TGT CCA
TTT TGA GTG TGT ACG GGT GGC ACG GGT TTT CCC TTT GTG GAA ACA GGT
K T H T C P P C P K G K H L C P>

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          920          930          940          950          960
          *          *          *          *          *
AGT CCC CTA TTT CCC GGA CCT TCT AAG CCC TTT TGG GTG CTG GTG GTG
TCA GGG GAT AAA GGG CCT GGA AGA TTC GGG AAA ACC CAC GAC CAC CAC
S   P   L   F   P   G   P   S   K   P   F   W   V   L   V   V>

          970          980          990          1000
          *          *          *          *
GTT GGT GGA GTC CTG GCT TGC TAT AGC TTG CTA GTA ACA GTG GCC TTT
CAA CCA CCT CAG GAC CGA ACG ATA TCG AAC GAT CAT TGT CAC CGG AAA
V   G   G   V   L   A   C   Y   S   L   L   V   T   V   A   F>

1010          1020          1030          1040          1050
*          *          *          *          *
ATT ATT TTC TGG GTG AGG AGT AAG AGC AGC AGG CTC CTG CAC AGT GAC
TAA TAA AAG ACC CAC TCC TCA TTC TCC TCG TCC GAG GAC GTG TCA CTG
I   I   F   W   V   R   S   K   R   S   R   L   L   H   S   D>

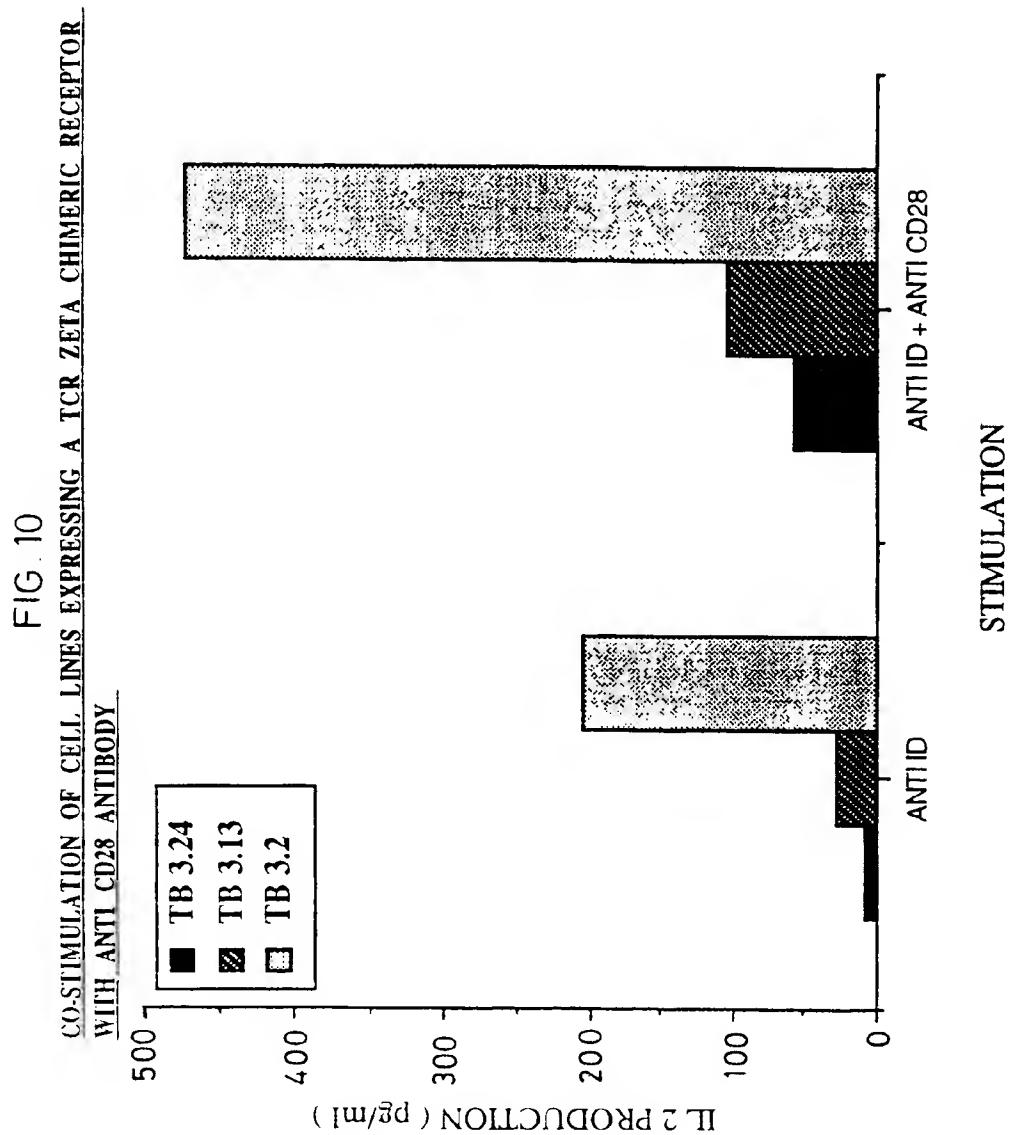
          1060          1070          1080          1090          1100
          *          *          *          *          *
TAC ATG AAC ATG ACT CCC CGC CGC CCC GGG CCC ACC CGC AAG CAT TAC
ATG TAC TTG TAC TGA GGG GCG GCG GGG CCC GGG TGG GCG TTC GTA ATG
Y   M   N   M   T   P   R   R   P   G   P   T   R   K   H   Y>

          1110          1120          1130          1140
          *          *          *          *
CAG CCC TAT GCC CCA CCA CGC GAC TTC GCA GCC TAT CGC TCC TGA
GTC GGG ATA CGG GGT GGT GCG CTG AAG CGT CGG ATA GCG AGG ACT
Q   P   Y   A   P   P   R   D   F   A   A   Y   R   S   *

```

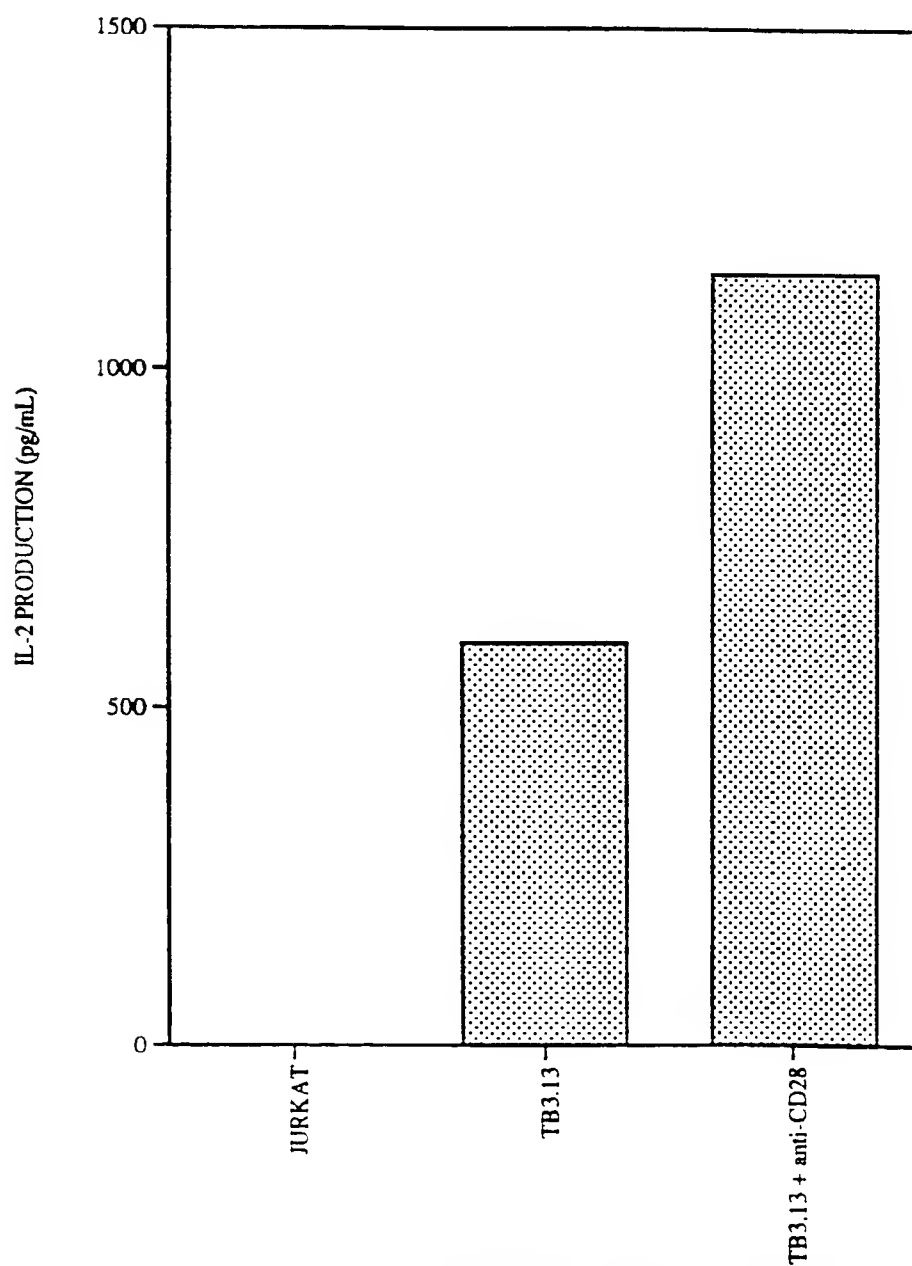
FIG. 9(contd.)

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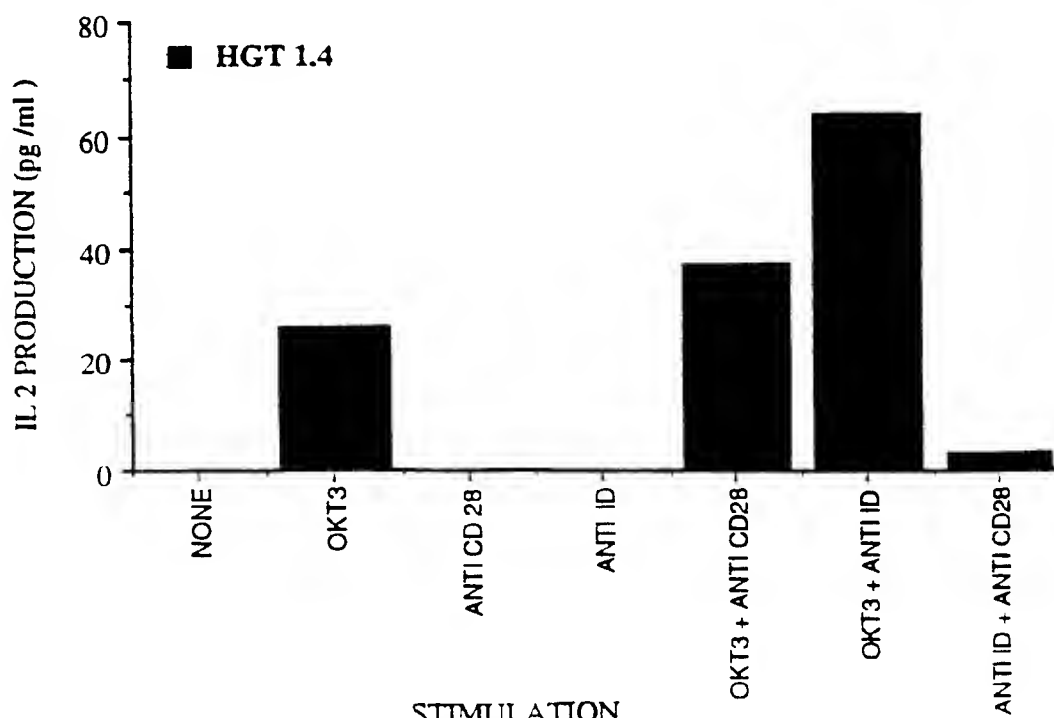
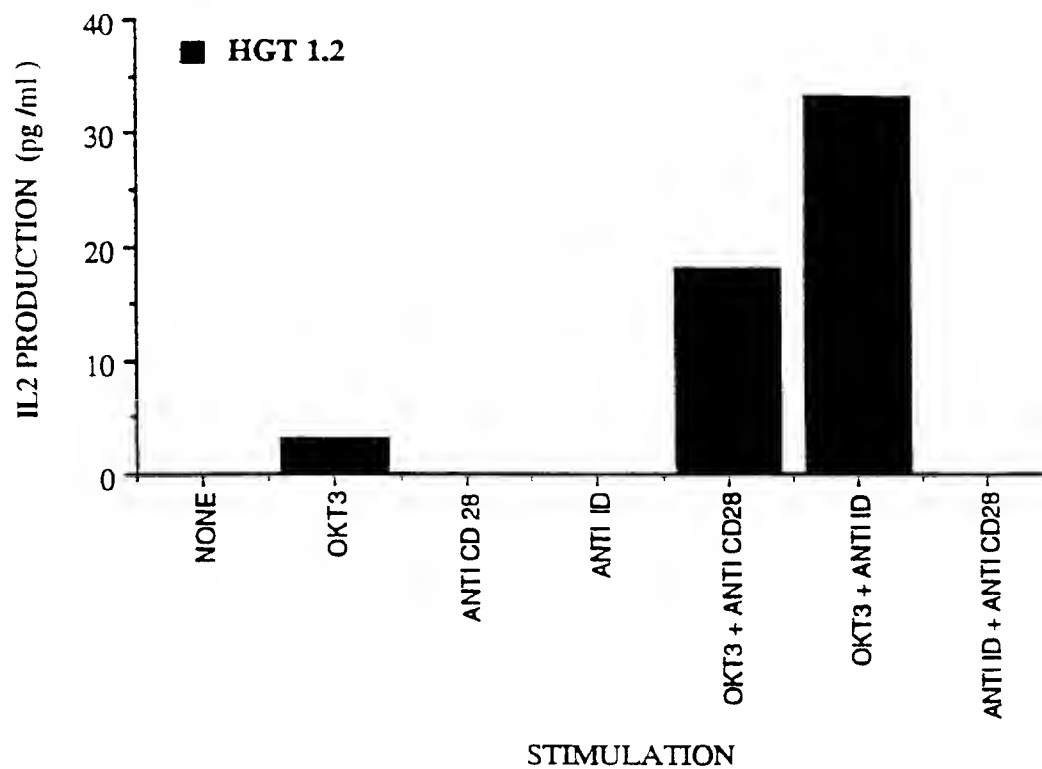
FIG. 11
STIMULATION WITH ANTIGEN POSITIVE CELLS.MCF-7



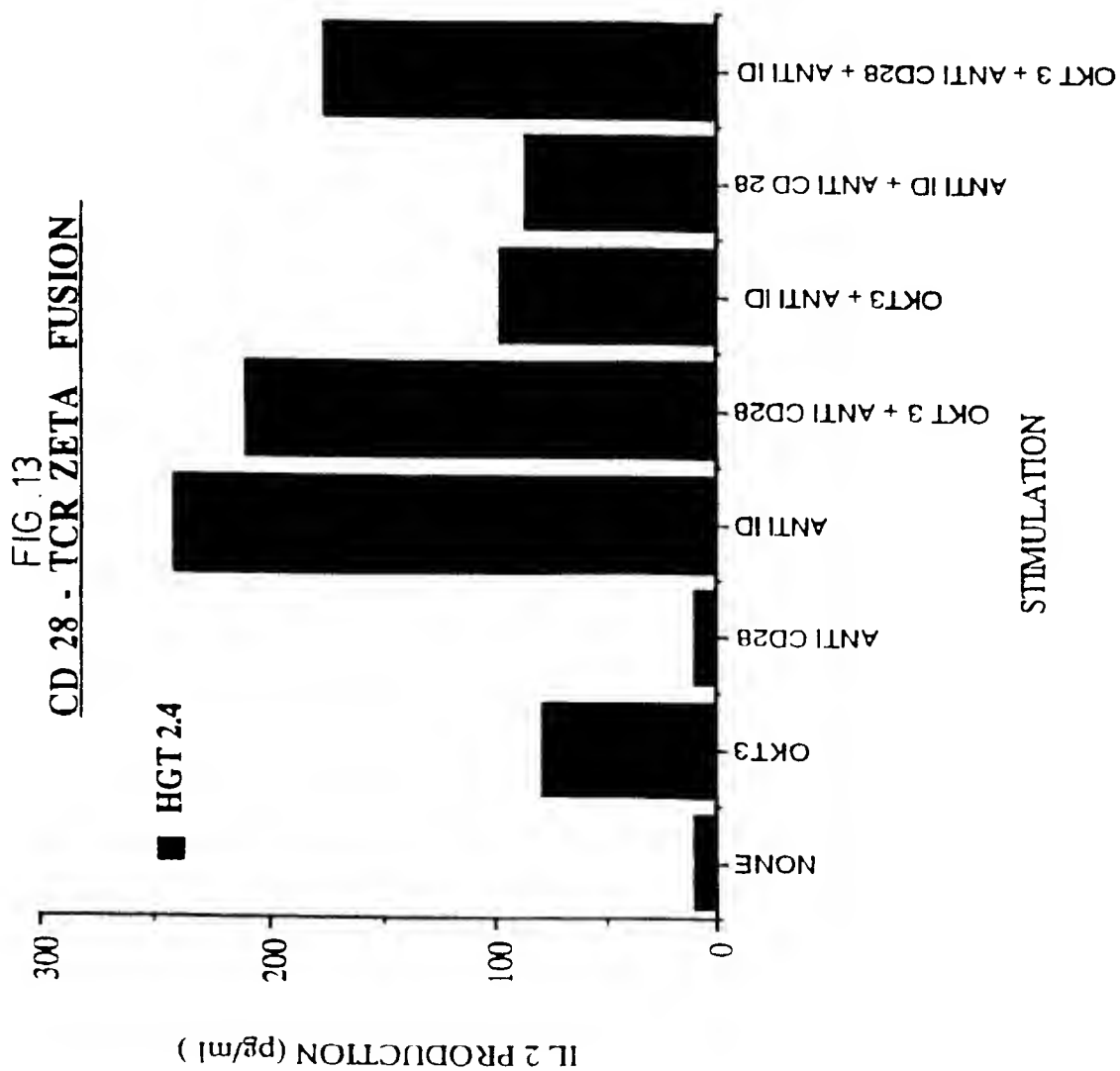
SUBSTITUTE SHEET (RULE 26)

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FIG. 12

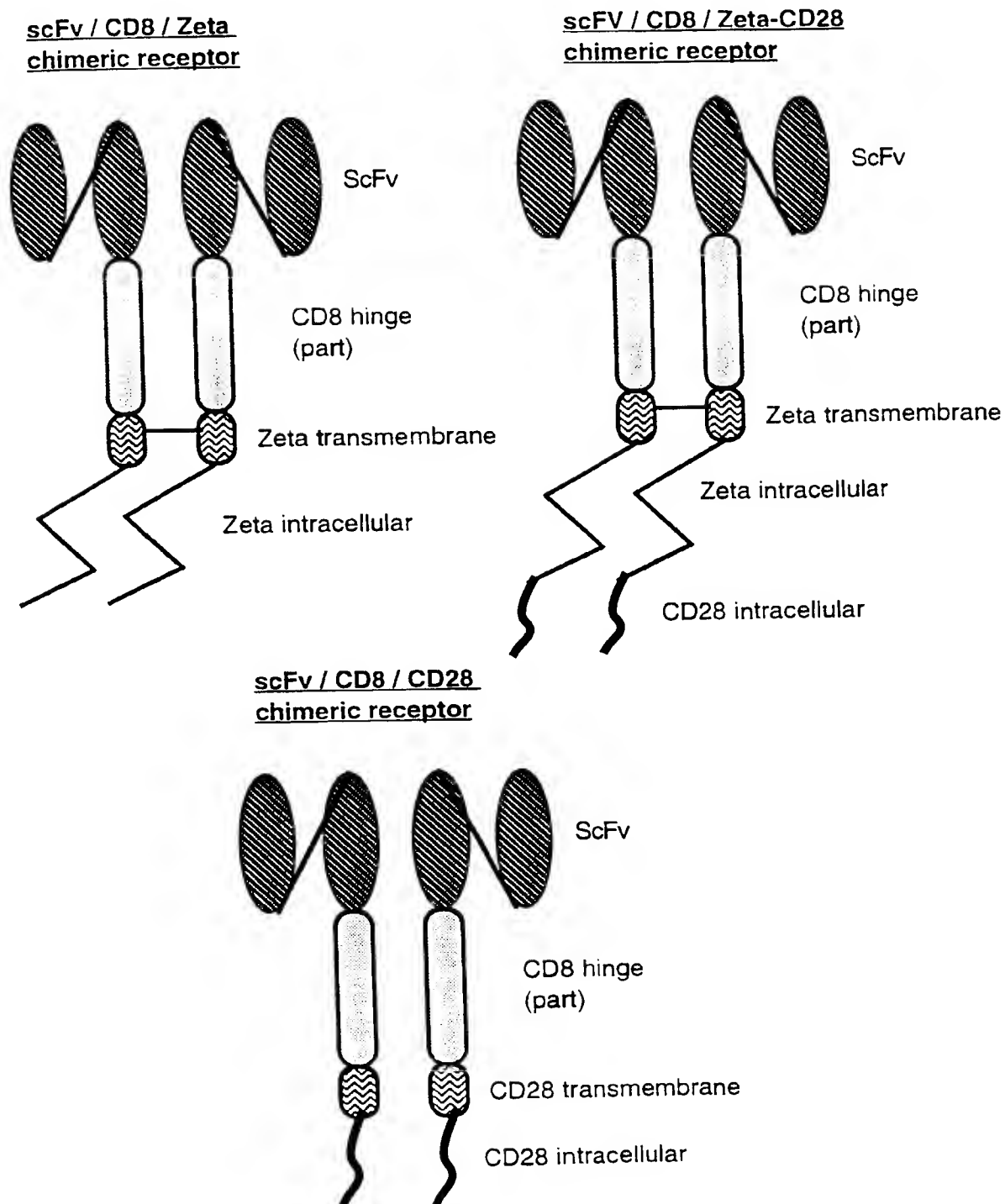
IL2 PRODUCTION IN RESPONSE TO VARIOUS STIMULI

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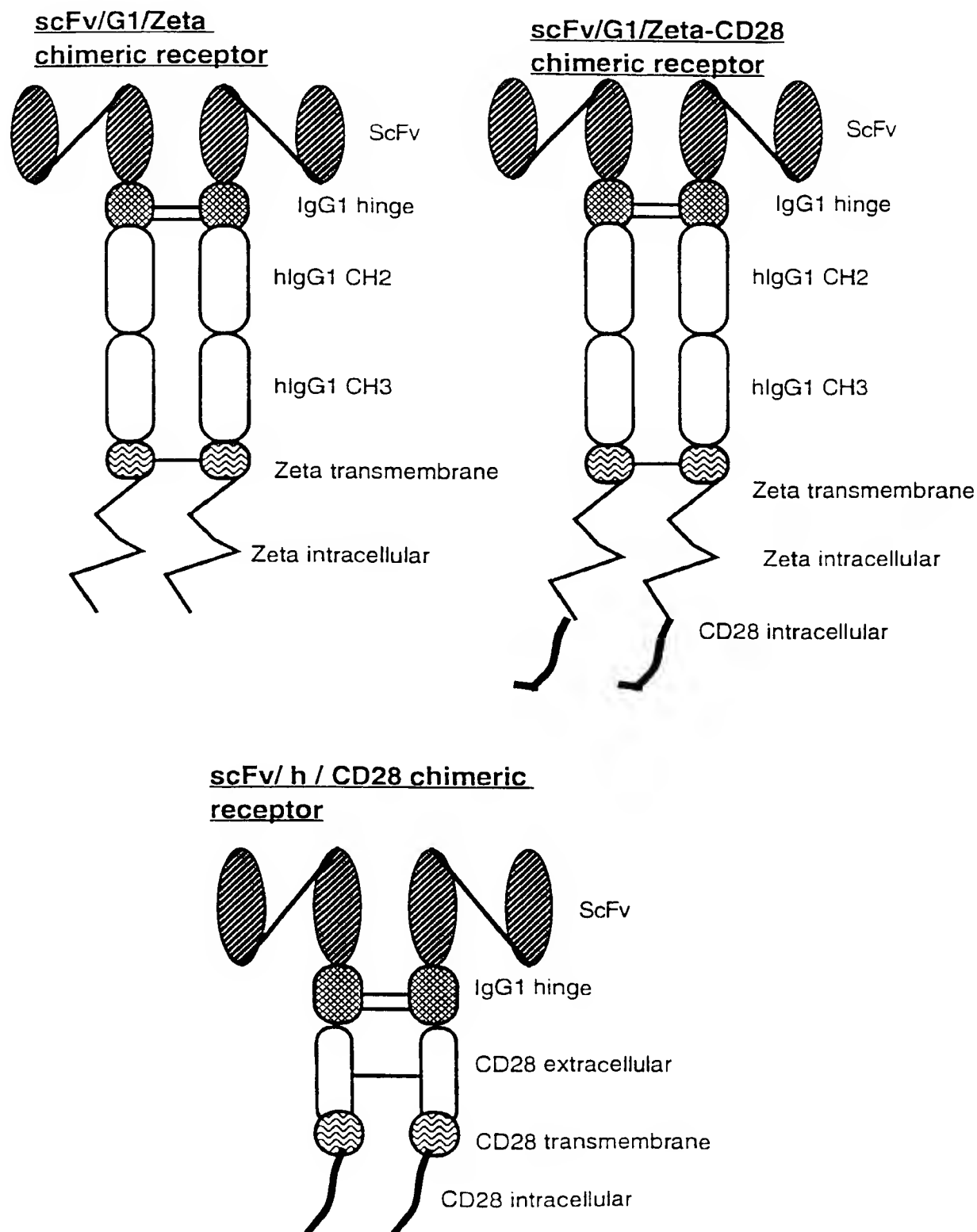
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FIG. 14



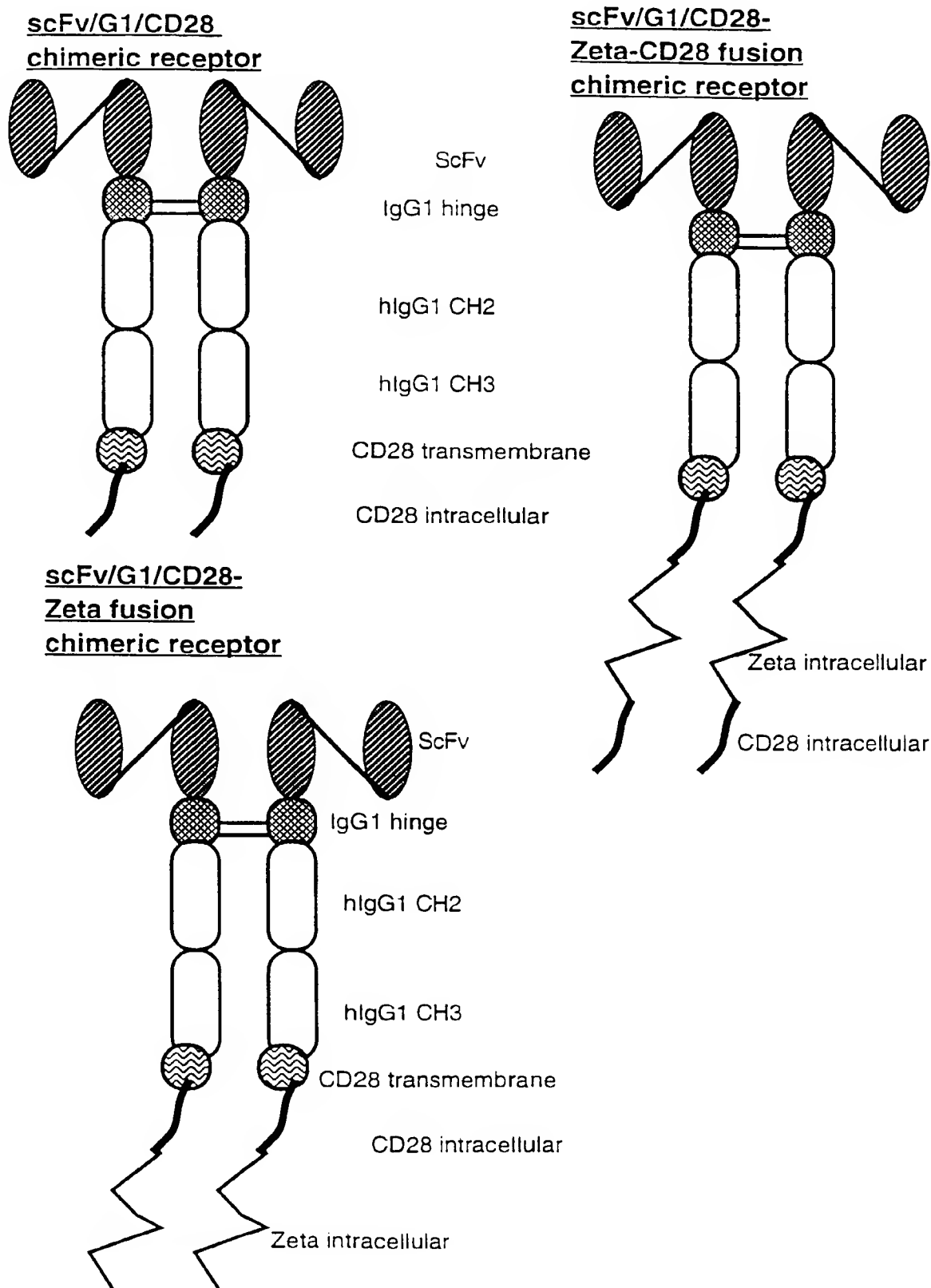
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FIG. 15



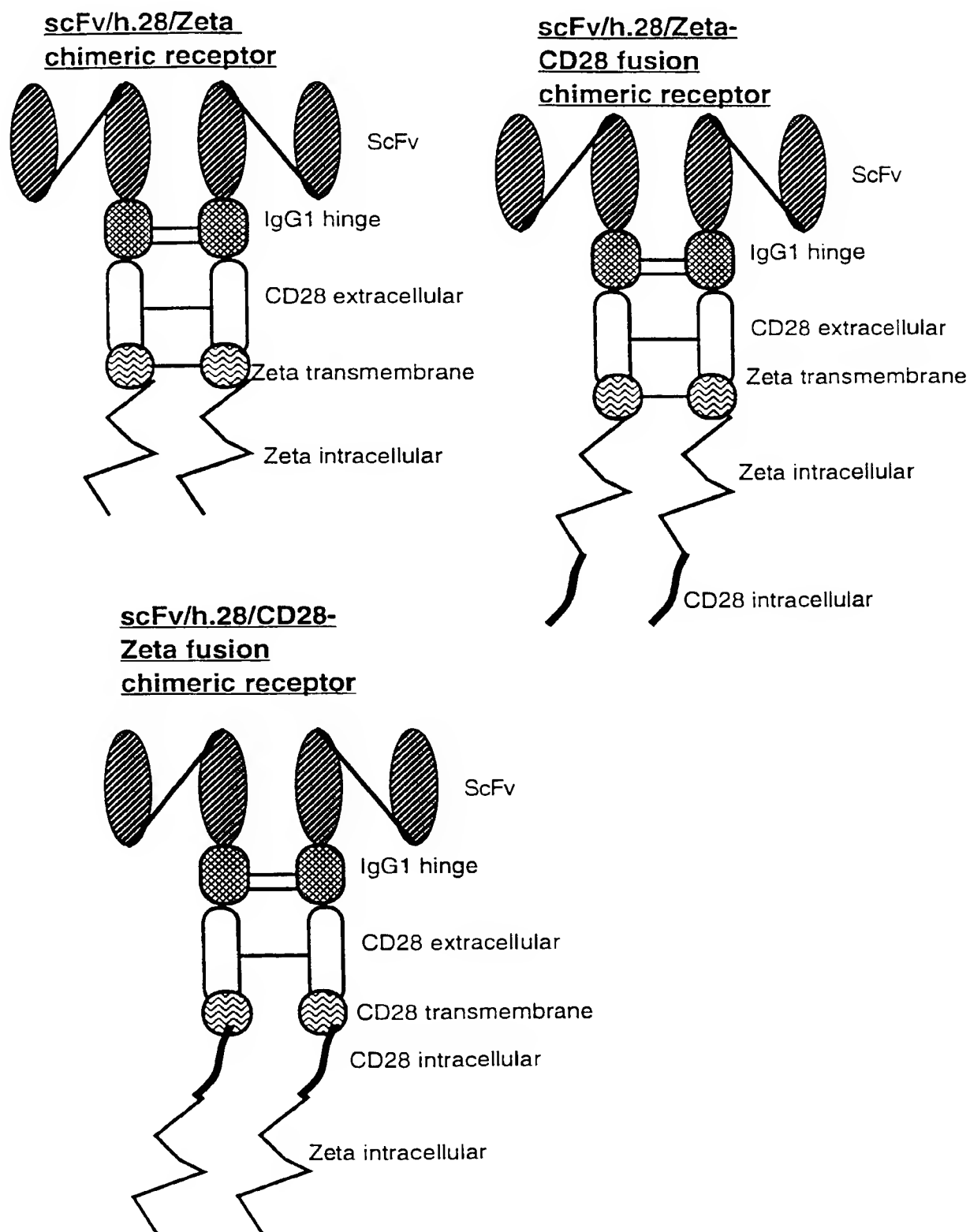
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FIG. 16



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FIG. 17



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FIG. 18
Surface expression of CD28-chimeras
in transfected Jurakat cell lines determined
by FITC-CD33 staining

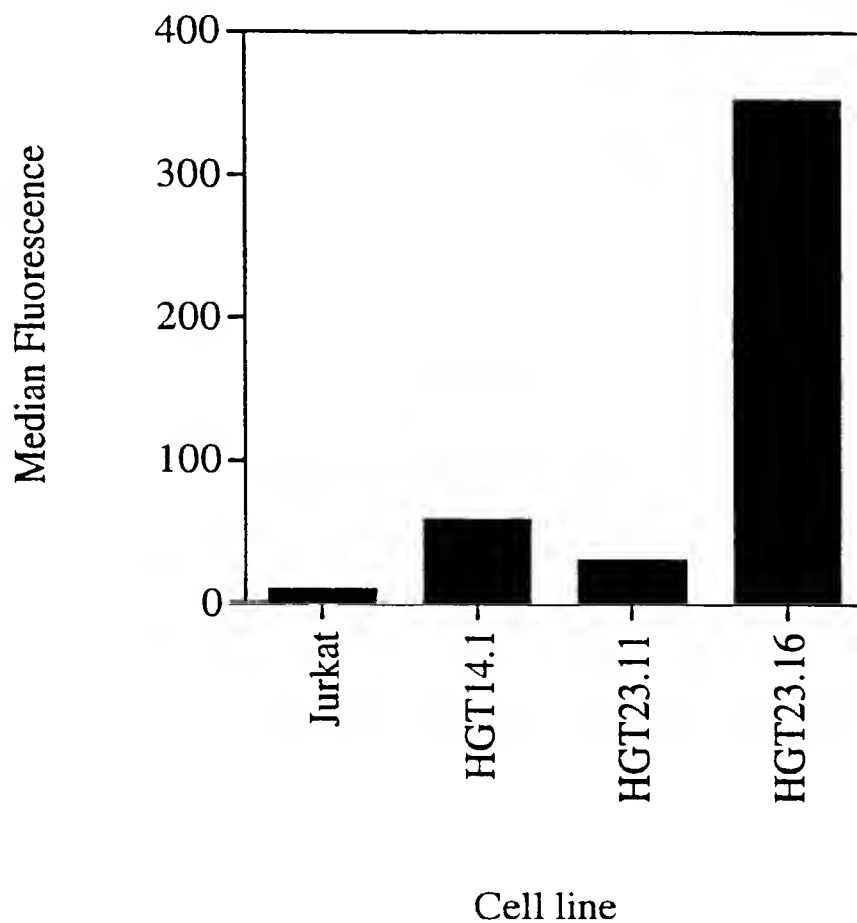
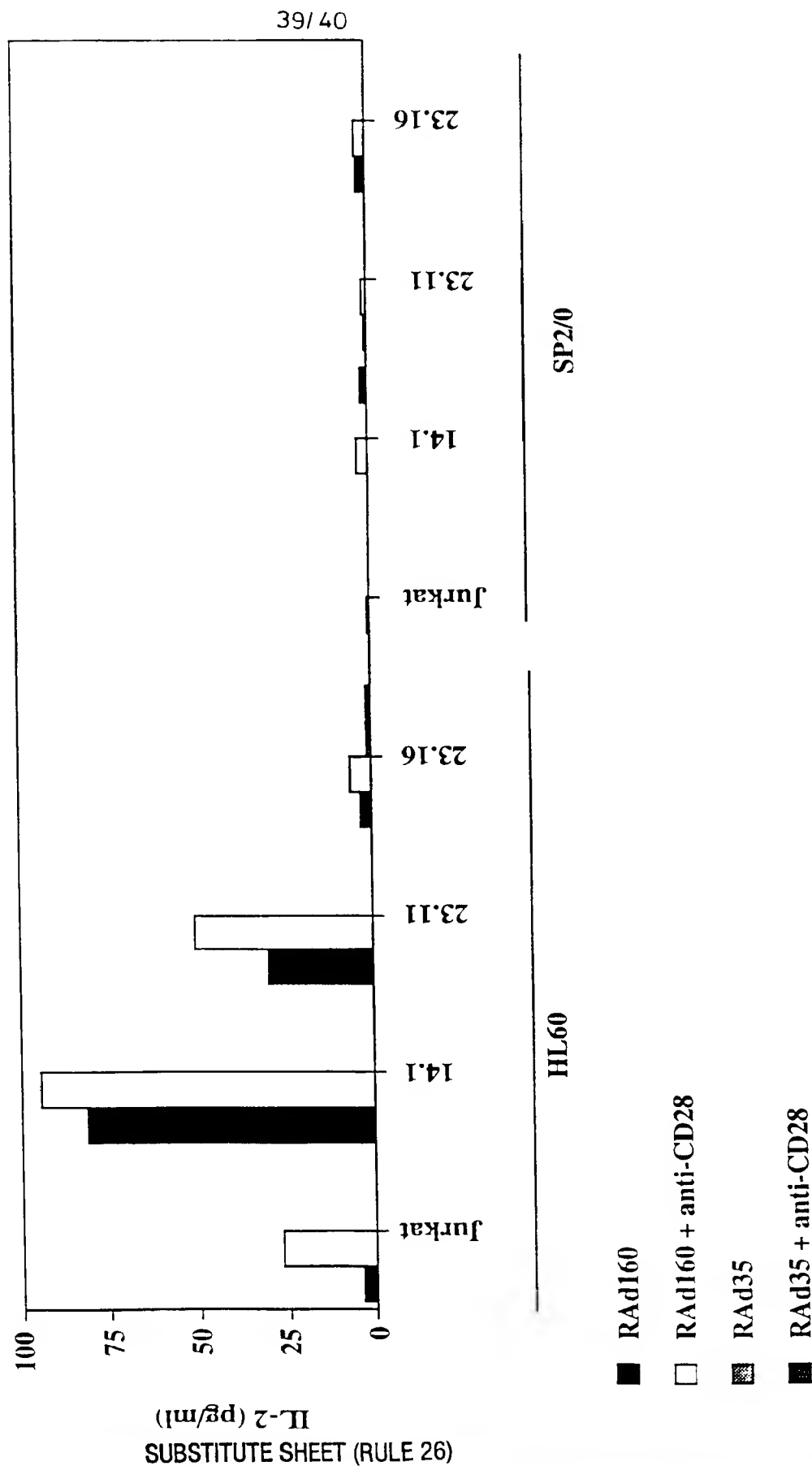
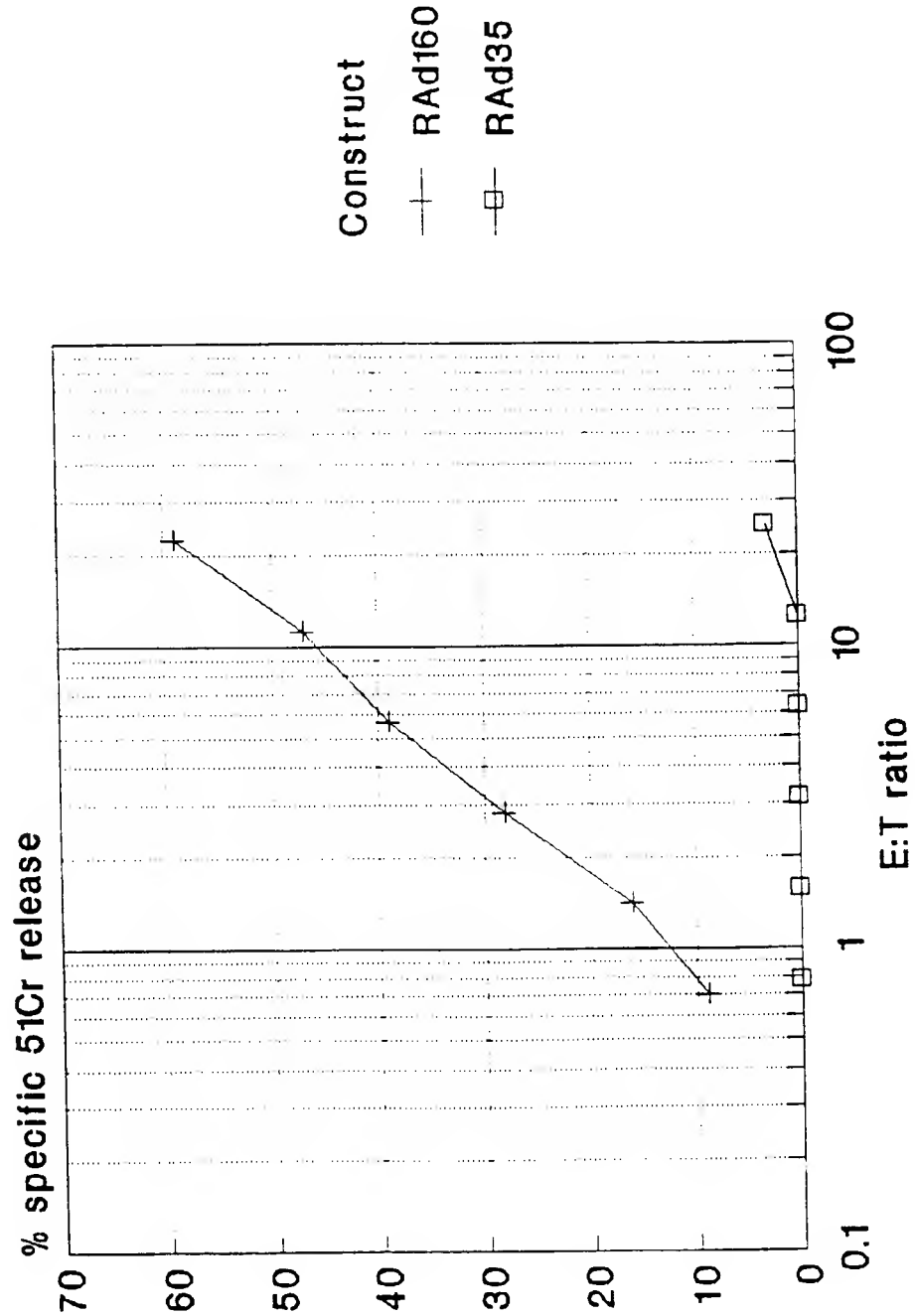


FIG. 19
IL-2 production by Jurkat cell lines expressing
p67-CD28 chimeras on infection with RAd160
stimulated with target cells



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FIG. 20
51Cr Release Assay
Adenovirus infected CD8+ve peripheral
blood lymphocytes with HL60 target cells





INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/GB96/03209 (22) International Filing Date: 23 December 1996 (23.12.96) (30) Priority Data: 9526131.9 21 December 1995 (21.12.95) GB (71) Applicant (for all designated States except US): CELLTECH THERAPEUTICS LTD. [GB/GB]; 216 Bath Road, Slough, Berkshire SL1 4EN (GB). (72) Inventors; and (75) Inventors/Applicants (for US only): BEBBINGTON, Christo- pher, Robert [GB/GB]; Berry Cottage, Westbrook, Boxford, Newbury, Berkshire RG20 8DJ (GB). LAWSON, Alastair, David, Griffiths [GB/GB]; Holden Farm, Cheriton, Alres- ford, Hants. SO24 0NX (GB). WEIR, Andrew, Neil, Charles [GB/GB]; 7 Willow Drive, Twyford, Berkshire RG10 9DD (GB). FINNEY, Helene, Margaret [GB/GB]; 64 Clare Road, Maidenhead, Berkshire SL6 4DQ (GB). (74) Agent: HALLYBONE, Huw, George; Carpmaels & Ransford, 43 Bloomsbury Square, London WC1A 2RA (GB).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the</i> <i>claims and to be republished in the event of the receipt of</i> <i>amendments.</i> (88) Date of publication of the international search report: 21 August 1997 (21.08.97)
(54) Title: CELL ACTIVATION PROCESS AND REAGENTS THEREFOR		
(57) Abstract <p>A cell activation process is described in which an effector cell is transformed with DNA coding for a chimeric receptor containing two or more different cytoplasmic signalling components. The activated cell may be of use in medicine for example in the treatment of diseases such as cancer.</p>		

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INTERNATIONAL SEARCH REPORT

Inten al Application No
PCT/GB 96/03209

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/12 C07K14/705 C12N15/62 C07K16/00 C12N5/10
A61K35/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 95 02686 A (THE GENERAL HOSPITAL CORPORATION) 26 January 1995 cited in the application see page 12, line 15 - page 16, line 14 see page 36, line 29 - page 51, line 16 ---	1-52
A	WO 93 19163 A (YEDA RESEARCH AND DEVELOPMENT CO. LTD.) 30 September 1993 cited in the application see page 7, line 9 - page 9, line 2 see page 16, line 14 - page 22, line 9 ---	1-52
P,X	WO 96 24671 A (CELL GENESYS, INC.) 15 August 1996 see page 3, line 17 - line 28 see page 10, line 24 - page 26, line 18 see page 28, line 11 - page 29, line 19 see page 30, line 25 - page 34, line 4 --- -/--	1-52

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	WO 96 23814 A (CELL GENESYS INC) 8 August 1996 see page 9, line 28 - page 10, line 31 see page 11, line 20 - page 12, line 9 see page 13, line 11 - page 23, line 26 see page 26, line 1 - line 12 see page 27, line 30 - page 34, line 28; examples -----	1-5, 7-13,15, 17-31, 36-52

INTERNATIONAL SEARCH REPORT

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